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<p>(54) Title: SYNTHETIC PEPTIDE INHIBITORS OF HIV TRANSMISSION (57) Abstract  The present invention relates to peptides which exhibit potent anti-retroviral activity. The peptides of the invention comprise DP-178 (SEQ ID:1) peptide corresponding to amino acids 638 to 673 of the HIV-1<sub>LAI</sub> gp41 protein, and fragments, analogs and homologs of DP-178. The invention further relates to the uses of such peptides as inhibitory of human and non-human retroviral, especially HIV, transmission to uninfected cells.</p>		

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SYNTHETIC PEPTIDE INHIBITORS OF HIV TRANSMISSION1. INTRODUCTION

The present invention relates to DP-178 (SEQ ID:1), a peptide corresponding to amino acids 638 to 673 of the HIV-1<sub>LAI</sub> transmembrane protein (TM) gp41, and portions, analogs, and homologs of DP-178 (SEQ ID:1), all of which exhibit anti-viral activity. Such anti-viral activity includes, but is not limited to, the inhibition of HIV transmission to uninfected CD-4<sup>+</sup> cells. Further, the invention relates to the use of DP-178 (SEQ ID:1) and DP-178 fragments and/or analogs or homologs as inhibitors of human and non-human retroviral, especially HIV, transmission to uninfected cells. Still further, the invention relates to the use of DP-178 as a HIV subtype-specific diagnostic. The present invention also relates to antiviral peptides analogous to DP-107, a peptide corresponding to amino acids 558 to 595 of the HIV-1<sub>LAI</sub> transmembrane protein (TM) gp41, that are present in other enveloped viruses. The present invention further relates to methods for identifying antiviral compounds that disrupt the interaction between DP-178 and DP-107, and/or between DP-107-like and DP-178-like peptides. The invention is demonstrated by way of a working example wherein DP-178 (SEQ ID:1), and a peptide whose sequence is homologous to DP-178 are each shown to be potent, non-cytotoxic inhibitors of HIV-1 transfer to uninfected CD-4<sup>+</sup> cells. The invention is further demonstrated by working examples wherein peptides having antiviral and/or structural similarity to DP-107 and DP-178 are identified.

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## 2. BACKGROUND OF THE INVENTION

### 2.1. THE HUMAN IMMUNODEFICIENCY VIRUS

The human immunodeficiency virus (HIV) has been implicated as the primary cause of the slowly degenerative immune system disease termed acquired  
5 immune deficiency syndrome (AIDS) (Barre-Sinoussi, F. et al., 1983, Science 220:868-870; Gallo, R. et al., 1984, Science 224:500-503). there are at least two distinct types of HIV: HIV-1 (Barre-Sinoussi, F. et al., 1983, Science 220:868-870; Gallo R. et al., 1984,  
10 Science 224:500-503) and HIV-2 (Clavel, F. et al., 1986, Science 233:343-346; Guyader, M. et al., 1987, Nature 326:662-669). Further, a large amount of genetic heterogeneity exists within populations of each of these types. Infection of human CD-4<sup>+</sup> T-  
15 lymphocytes with an HIV virus leads to depletion of the cell type and eventually to opportunistic infections, neurological dysfunctions, neoplastic growth, and ultimately death.

HIV is a member of the lentivirus family of  
20 retroviruses (Teich, N. et al., 1984, RNA Tumor Viruses, Weiss, R. et al., eds., CSH-Press, pp. 949-956). Retroviruses are small enveloped viruses that contain a diploid, single-stranded RNA genome, and replicate via a DNA intermediate produced by a  
25 virally-encoded reverse transcriptase, an RNA-dependent DNA polymerase (Varmus, H., 1988, Science 240:1427-1439). Other retroviruses include, for example, oncogenic viruses such as human T-cell leukemia viruses (HTLV-I, -II, -III), and feline  
30 leukemia virus.

The HIV viral particle consists of a viral core, composed of capsid proteins, that contains the viral RNA genome and those enzymes required for early  
35 replicative events. Myristylated Gag protein forms an



outer viral shell around the viral core, which is, in turn, surrounded by a lipid membrane envelope derived from the infected cell membrane. The HIV envelope surface glycoproteins are synthesized as a single 160 Kd precursor protein which is cleaved by a cellular protease during viral budding into two glycoproteins, gp41 and gp120. gp41 is a transmembrane protein and gp120 is an extracellular protein which remains non-covalently associated with gp41, possibly in a trimeric or multimeric form (Hammarskjold, M. and Rekosh, D., 1989, Biochem. Biophys. Acta 989:269-280).

HIV is targeted to CD-4<sup>+</sup> cells because the CD-4 cell surface protein acts as the cellular receptor for the HIV-1 virus (Dalglish, A. et al., 1984, Nature 312:763-767; Klatzmann et al., 1984, Nature 312:767-768; Maddon et al., 1986, Cell 47:333-348). Viral entry into cells is dependent upon gp120 binding the cellular CD-4<sup>+</sup> receptor molecules (McDougal, J.S. et al., 1986, Science 231:382-385; Maddon, P.J. et al., 1986, Cell 47:333-348) and thus explains HIV's tropism for CD-4<sup>+</sup> cells, while gp41 anchors the envelope glycoprotein complex in the viral membrane.

## 2.2. HIV TREATMENT

HIV infection is pandemic and HIV associated diseases represent a major world health problem. Although considerable effort is being put into the successful design of effective therapeutics, currently no curative anti-retroviral drugs against AIDS exist. In attempts to develop such drugs, several stages of the HIV life cycle have been considered as targets for therapeutic intervention (Mitsuya, H. et al., 1991, FASEB J. 5:2369-2381). For example, virally encoded reverse transcriptase has been one focus of drug development. A number of reverse-transcriptase-

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targeted drugs, including 2',3'-dideoxynucleoside analogs such as AZT, ddI, ddC, and d4T have been developed which have been shown to be active against HIV (Mitsuya, H. et al., 1991, Science 249:1533-1544). While beneficial, these nucleoside analogs are not  
5 curative, probably due to the rapid appearance of drug resistant HIV mutants (Lander, B. et al., 1989, Science 243:1731-1734). In addition, the drugs often exhibit toxic side effects such as bone marrow suppression, vomiting, and liver function  
10 abnormalities.

Attempts are also being made to develop drugs which can inhibit viral entry into the cell, the earliest stage of HIV infection. Here, the focus has thus far been on CD4, the cell surface receptor for  
15 HIV. Recombinant soluble CD4, for example, has been shown to inhibit infection of CD-4<sup>+</sup> T-cells by some HIV-1 strains (Smith, D.H. et al., 1987, Science 238:1704-1707). Certain primary HIV-1 isolates, however, are relatively less sensitive to inhibition  
20 by recombinant CD-4 (Daar, E. et al., 1990, Proc. Natl. Acad. Sci. USA 87:6574-6579). In addition, recombinant soluble CD-4 clinical trials have produced inconclusive results (Schooley, R. et al., 1990, Ann. Int. Med. 112:247-253; Kahn, J.O. et al., 1990, Ann.  
25 Int. Med. 112:254-261; Yarchoan, R. et al., 1989, Proc. Vth Int. Conf. on AIDS, p. 564, MCP 137).

The late stages of HIV replication, which involve crucial virus-specific secondary processing of certain viral proteins, have also been suggested as possible  
30 anti-HIV drug targets. Late stage processing is dependent on the activity of a viral protease, and drugs are being developed which inhibit this protease (Erickson, J., 1990, Science 249:527-533). The

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clinical outcome of these candidate drugs is still in question.

Attention is also being given to the development of vaccines for the treatment of HIV infection. The HIV-1 envelope proteins (gp160, gp120, gp41) have been shown to be the major antigens for anti-HIV antibodies present in AIDS patients (Barin, et al., 1985, Science 228:1094-1096). Thus far, therefore, these proteins seem to be the most promising candidates to act as antigens for anti-HIV vaccine development. To this end, several groups have begun to use various portions of gp160, gp120, and/or gp41 as immunogenic targets for the host immune system. See for example, Ivanoff, L. et al., U.S. Pat. No. 5,141,867; Saith, G. et al., WO 92/22,654; Shafferman, A., WO 91/09,872; Formoso, C. et al., WO 90/07,119. Clinical results concerning these candidate vaccines, however, still remain far in the future.

Thus, although a great deal of effort is being directed to the design and testing of anti-retroviral drugs, a truly effective, non-toxic treatment is still needed.

### 3. SUMMARY OF THE INVENTION

The present invention relates to DP-178 (SEQ ID:1), a 36-amino acid synthetic peptide corresponding to amino acids 638 to 673 of the transmembrane protein (TM) gp41 from the HIV-1 isolate LAI, which exhibits potent anti-HIV-1 activity. As evidenced by the example presented below, in Section 6, the DP-178 (SEQ ID:1) anti-viral activity is so high that, on a weight basis, no other known anti-HIV agent is effective at concentrations as low as those at which DP-178 (SEQ ID:1) exhibits its inhibitory effects. The invention further relates to those portions, analogs, and

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homologs of DP-178 which also show such antiviral activity. The antiviral activity of such DP-178 portions, analogs, and homologs, includes, but is not limited to the inhibition of HIV transmission to uninfected CD-4<sup>+</sup> cells. The invention relates to the  
5 use of DP-178 (SEQ ID:1) and DP-178 fragments and/or analogs or homologs. Such uses may include, but are not limited to, the use of the peptides as inhibitors of human and non-human retroviral, especially HIV, transmission to uninfected cells, and as type and/or  
10 subtype-specific diagnostic tools.

An embodiment of the invention is demonstrated below wherein an extremely low concentration of DP-178 (SEQ ID:1), and very low concentrations of a DP-178 homolog (SEQ ID:3) are shown to be potent inhibitors  
15 of HIV-1 mediated CD-4<sup>+</sup> cell-cell fusion (*i.e.*, syncytial formation) and infection of CD-4<sup>+</sup> cells by cell-free virus. Further, it is shown that DP-178 (SEQ ID:1) is not toxic to cells, even at concentrations 3 logs higher than the inhibitory  
20 DP-178 (SEQ ID:1) concentration.

The invention also relates to analogous DP178 peptides in other enveloped viruses that demonstrate similar antiviral properties.

The invention further relates to peptides  
25 analogous to DP-107, a peptide corresponding to amino acids 558-595 of the HIV-1<sub>LA1</sub> transmembrane protein (TM) of gp41, that are present in other enveloped viruses, and demonstrate antiviral properties. The present invention is based, in part, on the surprising  
30 discovery that the DP-107 and DP-108 domains of the gp41 protein non-covalently complex with each other, and that their interaction is necessary for the normal activity of the virus. The invention, therefore, further relates to methods for identifying antiviral  
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compounds that disrupt the interaction between DP-107 and DP-178, and/or between DP-107-like and DP-178-like peptides.

Embodiments of the invention are demonstrated, below, wherein peptides having structural and/or  
5 similarity to DP-107 and DP-178 are identified.

### 3.1. DEFINITIONS

Peptides are defined herein as organic compounds comprising two or more amino acids covalently joined  
10 by peptide bonds. Peptides may be referred to with respect to the number of constituent amino acids, i.e., a dipeptide contains two amino acid residues, a tripeptide contains three, etc. Peptides containing  
15 ten or fewer amino acids may be referred to as oligopeptides, while those with more than ten amino acid residues are polypeptides.

Peptide sequences defined herein are represented by one-letter symbols for amino acid residues as follows:

20 A (alanine)  
R (arginine)  
N (asparagine)  
D (aspartic acid)  
C (cysteine)  
25 Q (glutamine)  
E (glutamic acid)  
G (glycine)  
H (histidine)  
I (isoleucine)  
30 L (leucine)  
K (lysine)  
M (methionine)  
F (phenylalanine)  
P (proline)  
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S (serine)  
T (threonine)  
W (tryptophan)  
Y (tyrosine)  
V (valine)

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#### 4. BRIEF DESCRIPTION OF THE FIGURES

FIG. 1. Amino acid sequence of DP-178 (SEQ ID:1) derived from HIV<sub>LAI</sub>; DP-178 homologs derived from HIV-1<sub>SF2</sub> (DP-185; SEQ ID:3), HIV-1<sub>RF</sub> (SEQ ID:4), and  
10 HIV-1<sub>MN</sub> (SEQ ID:5); DP-178 homologs derived from amino acid sequences of two prototypic HIV-2 isolates, namely, HIV-2<sub>rod</sub> (SEQ ID:6) and HIV-2<sub>NH2</sub> (SEQ ID:7); control peptides: DP-180 (SEQ ID:2), a peptide incorporating the amino acid residues of DP-178 in a  
15 scrambled sequence; DP-118 (SEQ ID:10) unrelated to DP-178, which inhibits HIV-1 cell free virus infection; DP-125 (SEQ ID:8), unrelated to DP-178, was also previously shown to inhibit HIV-1 cell free virus infection (Wild et al., 1992, Proc. Natl. Acad. Sci  
20 USA 89:10,537-10,541); DP-116 (SEQ ID:9), unrelated to DP-178 had previously been shown to be negative for inhibition of HIV-1 infection using the cell-free virus infection assay (Wild, et al., 1992, Proc. Natl. Acad. Sci USA 89:10,537-10,541). Throughout the  
25 figures, the one letter amino acid code is used.

FIG. 2. Inhibition of HIV-1 cell-free virus infection by synthetic peptides. IC50 refers to the concentration of peptide that inhibits RT production from infected cells by 50% compared to the untreated  
30 control. Control: the level of RT produced by untreated cell cultures infected with the same level of virus as treated cultures.

FIG. 3. Inhibition of HIV-1 and HIV-2 cell-free virus infection by the synthetic peptide DP-178 (SEQ  
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ID:1). IC50: concentration of peptide that inhibits RT production by 50% compared to the untreated control. Control: Level of RT produced by untreated cell cultures infected with the same level of virus as treated cultures.

5       FIG. 4A. Fusion Inhibition Assay. DP-178 (SEQ ID:1) inhibition of HIV-1 prototypic isolate-mediated syncytia formation. Data represents the number of virus-induced syncytia per cell.

10       FIG. 4B. Fusion Inhibition Assay. DP-180 (SEQ ID:2): scrambled control peptide. DP-185 (SEQ ID:3): DP-178 homolog derived from HIV-1<sub>SP2</sub> isolate. Control: number of syncytia produced in the absence of peptide.

15       FIG. 5. Fusion inhibition assay: HIV-1 vs. HIV-2. Data represents the number of virus-induced syncytia per well. ND: not done.

      FIG. 6. Cytotoxicity study of DP-178 (SEQ ID:1) and DP-116 (SEQ ID:9) on CEM cells. Cell proliferation data is shown.

20       FIG. 7. Schematic representation of HIV-gp41 and maltose binding protein (MBP)-gp41 fusion proteins. DP107 and DP178 are synthetic peptides based on the two putative helices of gp41. The letter P in the DP107 boxes denotes an Ile to Pro mutation at amino acid number 578. Amino acid residues are  
25       numbered according to Meyers et al., Human Retroviruses and AIDS, 1991, Theoret. Biol. and Biophys. Group, Los Alamos Natl. Lab., Los Alamos, NM.

      FIG. 8. A point mutation alters the conformation and anti-HIV activity of M41.

30       FIG. 9. Abrogation of DP178 anti-HIV activity. Cell fusion assays were carried out in the presence of 10 nM DP178 and various concentrations of M41Δ178 or M41PA178.

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FIG. 10. Binding of DP178 to leucine zipper of gp41 analyzed by ELISA.

FIG. 11A-B. Models for a structural transition in the HIV-1 TM protein. Two models are proposed which indicate a structural transition from a native oligomer to a fusogenic state following a trigger event (possibly gp120 binding to CD4). Common features of both models include (1) the native state is held together by noncovalent protein-protein interactions to form the heterodimer of gp120/41 and other interactions, principally through gp41 interactive sites, to form homo-oligomers on the virus surface of the gp120/41 complexes; (2) shielding of the hydrophobic fusogenic peptide at the N-terminus (F) in the native state; and (3) the leucine zipper domain (DP107) exists as a homo-oligomer coiled coil only in the fusogenic state. The major differences in the two models include the structural state (native or fusogenic) in which the DP107 and DP178 domains are complexed to each other. In the first model (A; FIG. 11A) this interaction occurs in the native state and in B during the fusogenic state. When triggered, the fusion complex in the model depicted in (A) is generated through formation of coiled-coil interactions in homologous DP107 domains resulting in an extended  $\alpha$ -helix. This conformational change positions the fusion peptide for interaction with the cell membrane. In the second model (B; FIG. 11B), the fusogenic complex is stabilized by the association of the DP178 domain with the DP107 coiled-coil.

FIG. 12. Motif design using heptad repeat positioning of amino acids of known coiled-coils.

FIG. 13. Motif design using proposed heptad repeat positioning of amino acids of DP-107 and DP-178.



FIG. 14. Hybrid motif design crossing GCN4 and DP-107.

FIG. 15. Hybrid motif design crossing GCN4 and DP-178.

5 FIG. 16. Hybrid motif design 107x178x4, crossing DP-107 and DP-178. This motif was found to be the most consistent at identifying relevant DP-107-like and DP-178-like peptide regions.

10 FIG. 17. Hybrid motif design ALLMOTI5, crossing GCN4, DP-107, and DP-178.

FIG. 18. Hybrid motif design crossing GCN4, DP-107, DP-178, c-Fos c-Jun, c-Myc, and Flu Loop 36.

FIG. 19. Motifs designed to identify N-terminal proline-leucine zipper motifs.

15 FIG. 20. Search results for HIV-1 (BRU isolate) envelope protein gp41. Sequence search motif designations: Spades (♠): 107x178x4; Hearts (♥) ALLMOTI5; Clubs (♣): PLZIP; Diamonds (♦): transmembrane region (the putative transmembrane domains were identified using a PC/Gene program  
20 designed to search for such peptide regions). Asterisk (\*): Lupas method. The amino acid sequences identified by each motif are bracketed by the respective characters. Representative sequences chosen based on all searches are underlined and in  
25 bold. DP-107 and DP-178 sequences are marked, and additionally double-underlined and italicized.

30 FIG. 21. Search results for human respiratory syncytial virus (RSV) strain A2 fusion glycoprotein F1. Sequence search motif designations are as in FIG. 20.

FIG. 22. Search results for simian immunodeficiency virus (SIV) envelope protein gp41 (AGM3 isolate). Sequence search motif designations are as in FIG. 20.

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FIG. 23. Search results for canine distemper virus (strain Onderstepoort) fusion glycoprotein 1. Sequence search motif designations are as in FIG. 20.

5 FIG. 24. Search results for newcastle disease virus (strain Australia-Victoria/32) fusion glycoprotein F1. Sequence search motif designations are as in FIG. 20.

10 FIG. 25. Search results for human parainfluenza 3 virus (strain NIH 47885) fusion glycoprotein F1. Sequence search motif designations are as in FIG. 20.

15 FIG. 26. Search results for influenza A virus (strain A/AICHI/2/68) hemagglutinin precursor HA2. Sequence search designations are as in FIG. 20.

20 FIG. 27. Coiled-coil structural similarity and anti-RSV antiviral activity of 35-mer peptides synthesized utilizing the sequence of a 48-amino acid RSV F2 peptide which spans sequences identified utilizing the computer-assisted searches described herein. For the exact location and motifs utilized, see FIG. 21. "+" symbols are relative indicators of either structural similarity or antiviral activity, with a greater number of "+" symbols indicating a higher relative similarity or antiviral activity.

25 FIG. 28. Coiled-coil structural similarity and anti-RSV antiviral activity of 35-mer peptides synthesized utilizing the sequence of a 53-amino acid RSV F1 peptide which spans sequences identified utilizing the computer-assisted searches described herein. See FIG. 21 for the exact location and motifs used. "+" symbols are as described for FIG. 27.

30 FIG. 29. Coiled-coil structural similarity and anti-human parainfluenza 3 virus (HPF3) antiviral activity of 35-mer peptides synthesized utilizing the  
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sequence of a 56-amino acid HPF3 peptide which spans sequences identified utilizing computer-assisted searches described herein. For the exact location and motifs utilized, see FIG. 25. "+" symbols are as described in FIG. 27.

5                   FIG. 30. Coiled-coil structural similarity and anti-HPF3 antiviral activity of 35-mer peptides synthesized utilizing the sequence of a 70-amino acid HPF3 peptide which spans sequences identified  
10                   utilizing the computer-assisted searches described herein. For the exact location and motifs utilized, see FIG. 25. "+" symbols are as described in FIG. 27.

#### 5.     DETAILED DESCRIPTION OF THE INVENTION

Described herein are peptides that exhibit potent  
15                   antiviral activity. These peptides include DP-178 (SEQ ID:1), a gp41-derived 36 amino acid peptide, fragments and/or analogs of DP-178, and peptides which are homologous to DP-178. In addition, these peptides may include peptides exhibiting anti-viral activity  
20                   which are analogous to DP-107, a 38 amino acid peptide corresponding to residues 558 to 595 of the HIV-1<sub>LAI</sub> transmembrane (TM) gp41 protein, and which are present in other enveloped viral proteins. Also described here are assays for testing the antiviral activities  
25                   of such peptides. The present invention is based, in part, of the surprising discovery that the DP-107 and DP-178 domains of the gp41 protein complex with each other via non-covalent protein-protein interactions which are necessary for normal activity of the virus.  
30                   As such, methods are described for the identification of antiviral compounds that disrupt the interaction between DP-107 and DP-178 peptides, and between DP-107-like and DP-178-like peptides. Finally, the use of the peptides of the invention as inhibitors of non-  
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human and human viral and retroviral, especially HIV, transmission are detailed, as is the use of the peptides as diagnostic indicators of the presence of specific, viruses, especially retroviruses.

While not limited to any theory of operation, the following model is proposed to explain the potent anti-HIV activity of DP178, based, in part, on the experiments described in the working examples, infra. In the viral protein, gp41, DP178 corresponds to a putative  $\alpha$ -helix region located in the C-terminal end of the gp41 ectodomain, and appears to associate with a distal site on gp41 whose interactive structure is influenced by the leucine zipper motif, a coiled-coil structure, referred to as DP107. The association of these two domains may reflect a molecular linkage or "molecular clasp" intimately involved in the fusion process. It is of interest that mutations in the C-terminal  $\alpha$ -helix motif of gp41 (i.e., the D178 domain) tend to enhance the fusion ability of gp41, whereas mutations in the leucine zipper region (i.e., the DP107 domain) decrease or abolish the fusion ability of the viral protein. It may be that the leucine zipper motif is involved in membrane fusion while the C-terminal  $\alpha$ -helix motif serves as a molecular safety to regulate the availability of the leucine zipper during virus-induced membrane fusion.

On the basis of the foregoing, two models are proposed of gp41-mediated membrane fusion which are schematically shown in FIG. 11A-B. The reason for proposing two models is that the temporal nature of the interaction between the regions defined by DP107 and DP178 cannot, as yet, be pinpointed. Each model envisions two conformations for gp41 - one in a "native" state as it might be found on a resting virion. The other in a "fusogenic" state to reflect

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conformational changes triggered following binding of gp120 to CD4 and just prior to fusion with the target cell membrane. The strong binding affinity between gp120 and CD4 may actually represent the trigger for the fusion process obviating the need for a pH change such as occurs for viruses that fuse within intracellular vesicles. The two major features of both models are: (1) the leucine zipper sequences (DP107) in each chain of oligomeric envelope are held apart in the native state and are only allowed access to one another in the fusogenic state so as to form the extremely stable coiled-coils, and (2) association of the DP178 and DP107 sites as they exist in gp41 occur either in the native or fusogenic state. FIG. 11A depicts DP178/DP107 interaction in the native state as a molecular class. On the other hand, if one assumes that the most stable form of the envelope occurs in the fusogenic state, the model in FIG. 11B can be considered.

When synthesized as peptides, both DP107 and DP178 are potent inhibitors of HIV infection and fusion, probably by virtue of their ability to form complexes with viral gp41 and interfere with its fusogenic process; e.g., during the structural transition of the viral protein from the native structure to the fusogenic state, the DP178 and DP107 peptides may gain access to their respective binding sites on the viral gp41, and exert a disruptive influence. DP107 peptides which demonstrate anti-HIV activity are described in Applicants' co-pending application Serial No. 07/927,532, filed August 7, 1992, which is incorporated by reference herein in its entirety.

As shown in the working examples, infra, a truncated recombinant gp41 protein corresponding the

ectodomain of gp41 containing both DP107 and DP178 domains (excluding the fusion peptide, transmembrane region and cytoplasmic domain of gp41) did not inhibit HIV-1 induced fusion. However, when a single mutation was introduced to disrupt the coiled-coil structure of the DP107 domain -- a mutation which results in a total loss of biological activity of DP107 peptides -- the inactive recombinant protein was transformed to an active inhibitor of HIV-1 induced fusion. This transformation may result from liberation of the potent DP178 domain from a molecular clasp with the leucine zipper, DP107 domain.

For clarity of discussion, the invention will be described for DP178 peptide inhibitors of HIV. However, the principles may be analogously applied to other fusogenic enveloped viruses, including but not limited to those viruses containing the peptides listed in Tables V through X, below.

#### 5.1. DP-178 AND DP-178-LIKE PEPTIDES

The peptide DP-178 (SEQ ID:1) of the invention corresponds to amino acid residues 638 to 673 of the transmembrane protein gp41 from the HIV-1<sub>LAI</sub> isolate, and has the 36 amino acid sequence (reading from amino to carboxy terminus):

NH<sub>2</sub>-YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-COOH (SEQ ID:1)

In addition to the full-length DP-178 (SEQ ID:1) 36-mer, the peptides of the invention may include truncations of the DP-178 (SEQ ID:1) peptide which exhibit antiviral activity. Such truncated DP-178 (SEQ ID:1) peptides may comprise peptides of between 3 and 36 amino acid residues (*i.e.*, peptides ranging in size from a tripeptide to a 36-mer polypeptide), and

may include but are not limited to those listed in Tables I and II, below. Peptide sequences in these tables are listed from amino (left) to carboxy (right) terminus. "X" may represent an amino group ( $-NH_2$ ) and "Z" may represent a carboxyl ( $-COOH$ ) group.

- 5 Alternatively, as described below, "X" and/or "Z" may represent a hydrophobic group, an acetyl group, a FMOC group, an amido group, or a covalently attached macromolecule.

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TABLE I  
DP-178 (SEQ ID:1) CARBOXY TRUNCATIONS

X-YTS-Z  
 X-YTSL-Z  
 X-YTSLI-Z  
 X-YTSLIH-Z  
 5 X-YTSLIHS-Z  
 X-YTSLIHSL-Z  
 X-YTSLIHSLI-Z  
 X-YTSLIHSLIE-Z  
 X-YTSLIHSLIEE-Z  
 X-YTSLIHSLIEES-Z  
 X-YTSLIHSLIEESQ-Z  
 10 X-YTSLIHSLIEESQN-Z  
 X-YTSLIHSLIEESQNNQ-Z  
 X-YTSLIHSLIEESQNNQQ-Z  
 X-YTSLIHSLIEESQNNQQE-Z  
 X-YTSLIHSLIEESQNNQQEK-Z  
 X-YTSLIHSLIEESQNNQQEKN-Z  
 X-YTSLIHSLIEESQNNQQEKNE-Z  
 X-YTSLIHSLIEESQNNQQEKNEQ-Z  
 15 X-YTSLIHSLIEESQNNQQEKNEQE-Z  
 X-YTSLIHSLIEESQNNQQEKNEQEL-Z  
 X-YTSLIHSLIEESQNNQQEKNEQELL-Z  
 X-YTSLIHSLIEESQNNQQEKNEQELLE-Z  
 X-YTSLIHSLIEESQNNQQEKNEQELLEL-Z  
 X-YTSLIHSLIEESQNNQQEKNEQELLELD-Z  
 X-YTSLIHSLIEESQNNQQEKNEQELLELDK-Z  
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKW-Z  
 20 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWA-Z  
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWAS-Z  
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWASLW-Z  
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWASLWN-Z  
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWASLWNW-Z  
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWASLWNWF-Z

25 The one letter amino acid code is used.

Additionally,

30 "X" may represent an amino group, a hydrophobic group, including but not limited to carbobenzoxyl, dansyl, or T-butyloxycarbonyl; an acetyl group; a 9-fluorenylmethoxy-carbonyl (FMOC) group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

35 "Z" may represent a carboxyl group; an amido group; a T-butyloxycarbonyl group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.



TABLE II  
DP-178 (SEQ ID:1) AMINO TRUNCATIONS

	X-NWF-Z
	X-WNWF-Z
	X-LWNWF-Z
	X-SLWNWF-Z
5	X-ASLWNWF-Z
	X-WASLWNWF-Z
	X-KWASLWNWF-Z
	X-DKWASLWNWF-Z
	X-LDKWASLWNWF-Z
	X-ELDKWASLWNWF-Z
	X-LELDKWASLWNWF-Z
10	X-LLELDKWASLWNWF-Z
	X-ELLELDKWASLWNWF-Z
	X-QELLELDKWASLWNWF-Z
	X-EQELLELDKWASLWNWF-Z
	X-NEQELLELDKWASLWNWF-Z
	X-KNEQELLELDKWASLWNWF-Z
	X-EKNEQELLELDKWASLWNWF-Z
	X-QEKNEQELLELDKWASLWNWF-Z
15	X-QQEKNEQELLELDKWASLWNWF-Z
	X-NQQEKNEQELLELDKWASLWNWF-Z
	X-QNQEKNEQELLELDKWASLWNWF-Z
	X-SQNQQEKNEQELLELDKWASLWNWF-Z
	X-ESQNQQEKNEQELLELDKWASLWNWF-Z
	X-EESQNQQEKNEQELLELDKWASLWNWF-Z
	X-IEESQNQQEKNEQELLELDKWASLWNWF-Z
20	X-LIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-SLIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-HSLIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-IHSLIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-LIHSLIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-SLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-TSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-Z
25	X-YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-Z

The one letter amino acid code is used.

Additionally,

"X" may represent an amino group, a hydrophobic group, including but not limited to carbobenzoxyl, dansyl, or T-butyloxycarbonyl; an acetyl group; a 9-fluorenylmethoxy-carbonyl group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

"Z" may represent a carboxyl group; an amido group; a T-butyloxycarbonyl group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

The antiviral peptides of the invention also include analogs of DP-178 and/or DP-178 truncations which may include, but are not limited to, peptides comprising the DP-178 (SEQ ID:1) sequence, or DP-178 truncated sequence, containing one or more amino acid  
5 substitutions, insertions and/or deletions. Analogs of DP-178 homologs, described below, are also within the scope of the invention. The DP-178 analogs of the invention exhibit antiviral activity, and may, further, possess additional advantageous features,  
10 such as, for example, increased bioavailability, and/or stability, or reduced host immune recognition.

HIV-1 and HIV-2 envelope proteins are structurally distinct, but there exists a striking amino acid conservation within the DP-178-  
15 corresponding regions of HIV-1 and HIV-2. The amino acid conservation is of a periodic nature, suggesting some conservation of structure and/or function. Therefore, one possible class of amino acid substitutions would include those amino acid changes  
20 which are predicted to stabilize the structure of the DP-178 peptides of the invention.

Amino acid substitutions may be of a conserved or non-conserved nature. Conserved amino acid substitutions consist of replacing one or more amino  
25 acids of the DP-178 (SEQ ID:1) peptide sequence with amino acids of similar charge, size, and/or hydrophobicity characteristics, such as, for example, a glutamic acid (E) to aspartic acid (D) amino acid substitution. When only conserved substitutions are  
30 made, the resulting peptide is functionally equivalent to DP-178 (SEQ ID:1) or the DP-178 peptide from which it is derived. Non-conserved substitutions consist of replacing one or more amino acids of the DP-178 (SEQ  
ID:1) peptide sequence with amino acids possessing  
35 dissimilar charge, size, and/or hydrophobicity

characteristics, such as, for example, a glutamic acid (E) to valine (V) substitution.

Amino acid insertions may consist of single amino acid residues or stretches of residues ranging from 2 to 15 amino acids in length. One or more insertions  
5 may be introduced into DP-178 (SEQ ID:1), DP-178 fragments, analogs and/or DP-178 homologs (described below).

Deletions of DP-178 (SEQ ID:1), DP-178 fragments, analogs, and/or DP-178 homologs (described below) are  
10 also within the scope of the invention. Such deletions consist of the removal of one or more amino acids from the DP-178 or DP-178-like peptide sequence, with the lower limit length of the resulting peptide sequence being 4 to 6 amino acids. Such deletions may  
15 involve a single contiguous or greater than one discrete portion of the peptide sequences.

The peptides of the invention may further include homologs of DP-178 (SEQ ID:1) and/or DP-178 truncations which exhibit antiviral activity. Such  
20 DP-178 homologs are peptides whose amino acid sequences are comprised of the amino acid sequences of peptide regions of other (*i.e.*, other than HIV-1<sub>LA1</sub>) viruses that correspond to the gp41 peptide region from which DP-178 (SEQ ID:1) was derived. Such  
25 viruses may include, but are not limited to, other HIV-1 isolates and HIV-2 isolates. DP-178 homologs derived from the corresponding gp41 peptide region of other (*i.e.*, non HIV-1<sub>LA1</sub>) HIV-1 isolates may include, for example, peptide sequences as shown below.

30

NH<sub>2</sub>-YTNTIYTLLEESQNQQEKNEQEELLELDKWASLWNWF-COOH (DP-185; SEQ ID:3);

35

NH<sub>2</sub>-YTGIIYNLLEESQNQQEKNEQEELLELDKWANLWNWF-COOH (SEQ ID:4);

NH<sub>2</sub>-YTSLIYSLLEKSQIQQEKNEQELLELDKWASLWNWF-COOH (SEQ ID:5).

SEQ ID:3 (DP-185), SEQ ID:4, and SEQ ID:5 are derived from HIV-1<sub>SP2</sub>, HIV-1<sub>RF</sub>, and HIV-1<sub>MN</sub> isolates, respectively. Underlined amino acid residues refer to those residues that differ from the corresponding position in the DP-178 (SEQ ID:1) peptide. One such DP-178 homolog, DP-185 (SEQ ID:3), is described in the Working Example presented in Section 6, below, where it is demonstrated that DP-185 (SEQ ID:3) exhibits antiviral activity. The DP-178 homologs of the invention may also include truncations, amino acid substitutions, insertions, and/or deletions, as described above.

In addition, striking similarities, as shown in FIG. 1, exist within the regions of HIV-1 and HIV-2 isolates which correspond to the DP-178 sequence. A DP-178 homolog derived from the HIV-2<sub>NH2</sub> isolate has the 36 amino acid sequence (reading from amino to carboxy terminus):

NH<sub>2</sub>-LEANISQSLEQAQIQQEKNMVELQKLNSWDVFTNWL-COOH (SEQ ID:7)

Table III and Table IV show some possible truncations of the HIV-2<sub>NH2</sub> DP-178 homolog, which may comprise peptides of between 3 and 36 amino acid residues (i.e., peptides ranging in size from a tripeptide to a 36-mer polypeptide). Peptide sequences in these tables are listed from amino (left) to carboxy (right) terminus. "X" may represent an amino group (-NH<sub>2</sub>) and "Z" may represent a carboxyl (-COOH) group. Alternatively, as described below, "X" and/or "Z" may represent a hydrophobic group, an acetyl group, a FMOC group, an amido group, or a covalently attached macromolecule, as described below.

TABLE III

HIV-2<sub>NH2</sub> DP-178 homolog carboxy truncations.

	X-LEA-Z
	X-LEAN-Z
	X-LEANI-Z
	X-LEANIS-Z
5	X-LEANISQ-Z
	X-LEANISQS-Z
	X-LEANISQSL-Z
	X-LEANISQSLE-Z
	X-LEANISQSLEQ-Z
	X-LEANISQSLEQA-Z
	X-LEANISQSLEQAQ-Z
10	X-LEANISQSLEQAQI-Z
	X-LEANISQSLEQAQIQ-Z
	X-LEANISQSLEQAQIQQ-Z
	X-LEANISQSLEQAQIQQE-Z
	X-LEANISQSLEQAQIQQEK-Z
	X-LEANISQSLEQAQIQQEKN-Z
	X-LEANISQSLEQAQIQQEKNM-Z
	X-LEANISQSLEQAQIQQEKNMY-Z
15	X-LEANISQSLEQAQIQQEKNMYE-Z
	X-LEANISQSLEQAQIQQEKNMYEL-Z
	X-LEANISQSLEQAQIQQEKNMYELQ-Z
	X-LEANISQSLEQAQIQQEKNMYELQK-Z
	X-LEANISQSLEQAQIQQEKNMYELQKL-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLN-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNS-Z
20	X-LEANISQSLEQAQIQQEKNMYELQKLNSW-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWD-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDV-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVF-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFT-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTN-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNW-Z
25	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z

The one letter amino acid code is used.

Additionally,

"X" may represent an amino group, a hydrophobic group, including but not limited to carbobenzoxyl, dansyl, or T-butyloxycarbonyl; an acetyl group; a 9-fluorenylmethoxy-carbonyl (Fmoc) group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

"Z" may represent a carboxyl group; an amido group; a T-butyloxycarbonyl group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

TABLE IV

HIV-2<sub>NDZ</sub> DP-178 homolog amino truncations.

	X-NWL-Z
	X-TNWL-Z
	X-FTNWL-Z
	X-VFTNWL-Z
5	X-DVFTNWL-Z
	X-WDVFTNWL-Z
	X-SWDVFTNWL-Z
	X-NSWDVFTNWL-Z
	X-LNSWDVFTNWL-Z
	X-KLNSWDVFTNWL-Z
	X-QKLNSWDVFTNWL-Z
10	X-LQKLNSWDVFTNWL-Z
	X-ELQKLNSWDVFTNWL-Z
	X-YELQKLNSWDVFTNWL-Z
	X-MYELQKLNSWDVFTNWL-Z
	X-NMYELQKLNSWDVFTNWL-Z
	X-KNMYELQKLNSWDVFTNWL-Z
	X-EKNMYELQKLNSWDVFTNWL-Z
	X-QEKNMYELQKLNSWDVFTNWL-Z
15	X-QQEKNMYELQKLNSWDVFTNWL-Z
	X-IQQEKNMYELQKLNSWDVFTNWL-Z
	X-QIQQEKNMYELQKLNSWDVFTNWL-Z
	X-AQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-QAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-EQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-LEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
20	X-SLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-QSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-SQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-ISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-NISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-ANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-EANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
25	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z

The one letter amino acid code is used.

Additionally,

"X" may represent an amino group, a hydrophobic group, including but not limited to carbobenzoxyl, dansyl, or T-butyloxycarbonyl; an acetyl group; a 9-fluorenylmethoxy-carbonyl (Fmoc) group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

"Z" may represent a carboxyl group; an amido group; a T-butyloxycarbonyl group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

## 5.2. DP-107 and DP-178 ANALOGOUS ANTIVIRAL PEPTIDES

Peptid sequences functionally corresponding, and thus analogous to, the DP-178 sequences of the invention, described, above, in Section 5.1 may be found in other, non-HIV-1 envelope viruses. Further, peptide sequences functionally corresponding, and thus analogous to, DP-107, an HIV-1-derived antiviral peptide, may also be found in other, non-HIV-1 envelope viruses. DP-107 is a 38 amino acid peptide corresponding to residues 558 to 595 of HIV-1<sub>LA1</sub> transmembrane (TM) gp41 protein, which exhibits potent anti-viral activity. DP-107 is more fully described in Applicant's co-pending U.S. Patent Application Ser. No. 07/927,532. These DP-107-like and DP-178-like analogous peptides and present in TM proteins of envelope viruses and preferably exhibit antiviral activity, most preferably antiviral activity which is specific to the virus in which their native sequences are found.

DP-107-like and DP-178-like peptides may be identified, for example, by utilizing a computer-assisted search strategy such as that described and demonstrated, below, in the Examples presented in Sections 9 through 16. The search strategy identifies regions in other viruses that are similar in predicted secondary structure to DP-107 and DP-178.

This search strategy is described fully, below, in the Example presented in Section 9. While this search strategy is based, in part, on a primary amino acid motif deduced from DP-107 and DP-178, it is not based solely on searching for primary amino acid sequence homologies, as such protein sequence homologies exist within, but not between major groups of viruses. For example, primary amino acid sequence homology is high within the TM protein of different

strains of HIV-1 or within the TM protein of different isolates of simian immunodeficiency virus (SIV).

Primary amino acid sequence homology between HIV-1 and SIV, however, is low enough so as not to be useful.

It is not possible, therefore, to find DP-107 or DP-  
5 178-like peptides within other viruses, whether structurally, or otherwise, based on primary sequence homology, alone.

Further, while it would be potentially useful to identify primary sequence arrangements of amino acids  
10 based on the physical chemical characteristics of different classes of amino acids rather than based on the specific amino acids themselves, for instance, a by concentrating on the coiled-coil nature of the peptide sequence, a computer algorithm designed by  
15 Lupas et al. to identify such coiled-coil propensities of regions within proteins (Lupas, A., et al., 1991 Science 252:1162-1164) is inadequate for identifying protein regions analogous to DP-107 or DP-178.

Specifically, analysis of HIV-1 gp160 (containing  
20 both gp120 and gp41) using the Lupas algorithm does not identify the coiled-coil region within DP-107. It does, however, identify a region within DP-178 beginning eight amino acids N-terminal to the start of DP-178 and ending eight amino acids from the C-  
25 terminus. The DP-107 peptide has been shown experimentally to form a stable coiled coil. A search based on the Lupas search algorithm, therefore, would not have identified the DP-107 coiled-coil region. Conversely, the Lupas algorithm identified the DP-178  
30 region as a potential coiled-coil motif. However, the peptide DP-178 derived from this region failed to form a coiled coil in solution. A possible explanation for the inability of the Lupas search algorithm to accurately identify coiled-coil sequences within the  
35 HIV-1 TM, is that the Lupas algorithm is based on the



structure of coiled c ils from proteins that are not structurally or functionally similar to the TM proteins of viruses, antiviral peptides (e.g. DP-107 and DP-178) of which are an object of this invention.

5 The computer search strategy of the invention, as demonstrated in the Examples presented below, in Sections 9 through 16, successfully identifies regions of viral TM proteins similar to DP-107 or DP-178. This search strategy was designed to be used with a commercially-available sequence database packages,  
10 preferably PC/Gene. A series of motifs were designed and engineered to range in stringency from very strict to very broad, as discussed in Section 9.

Among the protein sequence seach motifs which may be utilized in such a computer-assisted DP-107-like  
15 and DP-178-like antiviral peptide search are the 107x178x4 motif, the ALLMOTI5 motif, and the PLZIP series of motifs, each of which is described in the Example presented in Section 9, below, with 107x178x4 being preferred.

20 Coiled-coiled sequences are thought to consist of heptad amino acid repeats. For ease of description, the amino acid positions within the heptad repeats are sometimes referred to as A through G, with the first position being A, the second B, etc. The motifs used  
25 to identify DP-107-like and DP-178-like sequences herein are desined to specifically search for and identify such heptad repeats. In the descriptions of each of the motifs described, below, amino acids enclosed by brackets , i.e., [], designate the only  
30 amino acid residues that are acceptable at the given position, while amino acids enclosed by braces, i.e., {}, designate the only amino acids which are unacceptable at the given heptad position. When a set of bracketed or braced amino acids is followed by a  
35 number in parentheses i.e., (), it refers to the

number of subsequent amino acid positions for which the designated set of amino acids hold, e.g., a (2) means "for the next two heptad amino acid positions."

The ALLMOTI5 is written as follows:

5 {CDGHP}-{CFP}(2)-{CDGHP}-{CFP}(3)-  
 {CDGHP}-{CFP}(2)-{CDGHP}-{CFP}(3)-  
 {CDGHP}-{CFP}(2)-{CDGHP}-{CFP}(3)-  
 {CDGHP}-{CFP}(2)-{CDGHP}-{CFP}(3)-  
 {CDGHP}-{CFP}(2)-{CDGHP}-{CFP}(3)-

Translating this motif, it would read: "at the first (A) position of the heptad, any amino acid residue except C, D, G, H, or P is acceptable, at the next two (B,C) amino acid positions, any amino acid residue except C, F, or P is acceptable, at the fourth heptad position (D), any amino acid residue except C, D, G, H, or P is acceptable, at the next three (E, F, G) amino acid positions, any amino acid residue except C, F, or P is acceptable. This motif is designed to search for five consecutive heptad repeats (thus the repeat of the first line five times), meaning that it searches for 35-mer sized peptides. It may also be designed to search for 28-mers, by only repeating the initial motif four times. With respect to the ALLMOTI5 motif, a 35-mer search is preferred. Those viral sequences identified via such an ALLMOTI5 motif are listed in Table V, below, at the end of this Section. The viral sequences listed in Table V potentially exhibit antiviral activity, may be useful in the identification of antiviral compounds, and are intended to be within the scope of the invention.

The 107x178x4 motif is written as follows:

30 [EIKLNQSTVWY]-{CFMP}(2)-[EIKLNQSTVWY]-{CFMP}(3)-  
 [EIKLNQSTVWY]-{CFMP}(2)-[EIKLNQSTVWY]-{CFMP}(3)-  
 [EIKLNQSTVWY]-{CFMP}(2)-[EIKLNQSTVWY]-{CFMP}(3)-  
 [EIKLNQSTVWY]-{CFMP}(2)-[EIKLNQSTVWY]-{CFMP}(3)-

Translating this motif, it would read: "at the first (A) position of the heptad, any amino acid residue except E, F, I, K, L, N, Q, S, T, V, W, or Y

is acceptable, at the next two (B,C) amino acid positions, any amino acid residue except C, F, M or P is acceptable, at the fourth position (D), any amino acid residue except E, F, I, K, L, N, Q, S, T, V, W, or Y is acceptable, at the next three (E, F, G) amino acid positions, any amino acid residue except C, F, M or P is acceptable. This motif is designed to search for four consecutive heptad repeats (thus the repeat of the first line four times), meaning that it searches for 28-mer sized peptides. It may also be designed to search for 35-mers, by repeating the initial motif five times. With respect to the 107x178x4 motif, a 28-mer search is preferred. Those viral sequences identified via such a 107x178x4 motif are listed in Table V, below, at the end of this Section. The viral sequences listed in Table V potentially exhibit antiviral activity, may be useful in the the identification of antiviral compounds, and are intended to be within the scope of the invention.

The PLZIP series of motifs are as listed in FIG. 19. These motifs are designed to identify leucine zipper coiled-coil like heptads wherein at least one proline residue is present at some predefined distance N-terminal to the repeat. These PLZIP motifs find regions of proteins with similarities to HIV-1 DP-178 generally located just N-terminal to the transmembrane anchor. These motifs may be translated according to the same convention described above. Each line depicted in FIG. 19 represents a single, complete search motif. "X" in these motifs refers to any amino acid residue. In instances wherein a motif contains two numbers within parentheses, this refers to a variable number of amino acid residues. For example, X (1,12) is translated to "the next one to twelve amino acid residues, inclusive, may be any amino acid".

Tables VI through X, below, at the end of this

Section, list hits from such PLZIP motifs. The viral sequences listed in Table VI through X potentially exhibit antiviral activity, may be useful in the identification of antiviral compounds, and are intended to be within the scope of the invention.

5       The Examples presented in Sections 17 and 18, below, demonstrate that respiratory syncytial virus and parainfluenza virus sequences identified via such a computer search exhibit antiviral and/or structural characteristics similar to those of DP-107 or DP-178.

10       The DP-107-like and DP-178-like analogous peptides may, further, contain any of the additional groups described for DP-178, above, in Section 5.1. For example, these peptides may include any of the additional amino-terminal groups which "X" of Tables I  
15       through IV may represent, and may also include any of the carboxy-terminal groups which "Z" of Tables I through IV may represent.

          Additionally, such DP-107-like and DP-178-like peptides may further include DP-107-like or DP-178-like  
20       peptides, such as those listed in Tables V through X, above, containing one or more amino acid substitutions, insertions, and/or deletions. Also, analogs of such DP-107-like and DP-178-like peptides are intended to be within the scope of the invention.  
25       Such analogs of the invention may exhibit increased antiviral activity, and may, further, possess increased bioavailability, and/or stability, or reduced immune recognition.

          The DP-107-like and DP-178-like amino acid  
30       substitutions, insertions and deletions, are as described for DP-178, above, in Section 5.1. Analog modifications are as described, below, in Section 5.3.

35

**TABLE V**

**Search Results Summary for 107x178x4 and  
ALLMOTI5 Motifs**

[illegible]

PENV HV1H3	545-594	631-663	781-818			PENV HV1BN	801-580	809-708	783-831			
PENV HV1J3	556-606	842-694	802-828			PENV HV1BR	510-589	618-717	772-841			
PENV HV1JR		822-875	783-811			PENV HV1C4	810-608	628-724	778-855			
PENV HV1K8	555-586	637-677	776-824			PENV HV1EL	502-581	607-708	785-828			
PENV HV1MA	547-595	633-707	784-828			PENV HV1H2	505-584	610-712	767-838			
PENV HV1MF	543-592	828-681	789-818			PENV HV1H3	505-584	610-712	767-843			
PENV HV1MN	507-586	632-684	781-818			PENV HV1J3	517-605	622-723	778-843			
PENV HV1ND	536-583	821-673	783-813			PENV HV1JR	487-586	603-704	768-835			
PENV HV1OY	544-583	630-704	789-820			PENV HV1KB	511-545	555-589	618-718	772-848		
PENV HV1PV	545-584	831-683	781-818			PENV HV1MA	507-586	617-714	770-828			
PENV HV1RH	584-602	640-692	800-832			PENV HV1MF	503-592	622-710	765-841			
PENV HV1S1	538-585	822-674	782-808			PENV HV1MN	505-585	617-713	774-841			
PENV HV1S3	541-589	627-679	787-815			PENV HV1ND	485-584	601-702	767-826			
PENV HV18C	545-593	631-683				PENV HV1OY	487-583	610-711	768-842			
PENV HV1W1	545-593	631-683	781-818			PENV HV1PV	505-584	610-712	767-843			
PENV HV1W2	536-584	622-674	782-808			PENV HV1RH	507-603	618-721	770-852			
PENV HV1Z3	542-581	628-680	780-820			PENV HV1S1	496-586	602-703	768-830			
PENV HV1Z6	545-593	630-682	782-822			PENV HV1S3	494-580	607-708	763-837			
PENV HV1Z8	573-601	634-678	767-828			PENV HV18C	489-584	611-712	767-834			
PENV HV1ZH	545-584	627-668	781-823			PENV HV1W1	485-584	611-712	767-836			
PENV HV2BE	532-591	621-648	863-697			PENV HV1W2	489-584	602-703	768-827			
PENV HV2CA	534-593	623-650	855-689			PENV HV1Z2	502-591	607-708	764-831			
PENV HV2D1	523-580	568-582	844-688			PENV HV1Z6	504-583	608-711	768-840			
PENV HV2G1	524-581	550-583	813-640	845-693		PENV HV1Z8	512-601	617-675	882-718	774-831		
PENV HV2NZ	524-581	550-583	813-640	862-689		PENV HV1ZB	522-584	612-712	777-838			
PENV HV2RO	533-582	622-688				PENV HV2BE	510-585	617-680				
PENV HV282	527-584	559-588	848-682			PENV HV2CA	512-697	619-709				
PENV HV28B	557-584	614-673				PENV HV2D1	501-586	608-688				
PENV HV28T	527-584	559-588	848-682			PENV HV2G1	502-587	609-689				
PENV MCFF	473-512					PENV HV2NZ	488-587	609-688				
PENV MCFF3	489-516					PENV HV2RO	511-589	616-708				
PENV MLVAV	517-544					PENV HV282	505-590	612-702				
PENV MLVC8	810-538					PENV HV28B	520-588	614-700				
PENV MLVF8	523-583					PENV HV28T	505-590	612-702				
PENV MLVFF	523-563					PENV IPMAE	367-422	485-527				
PENV MLVFP	523-563					PENV JSRV	403-455	571-605				
PENV MLVHO	810-540					PENV MCFF	473-525	537-571				
PENV MLVK1	40-81					PENV MCFF3	474-526	538-572				
PENV MLVNO	802-543					PENV MLVAV	503-555	587-601				
PENV MLVRD	487-538					PENV MLVCS	498-550	582-586				
PENV MLVRK	487-538					PENV MLVFS	520-584	576-610				
PENV MMTVB	488-485	562-588				PENV MLVFF	520-584	576-610				
PENV MMTVG	458-485	562-588				PENV MLVFP	520-584	576-610				
PENV MPMV	422-470					PENV MLVHO	504-551	563-587				
PENV MSVB	57-84					PENV MLVK1	40-82	104-138				
PENV OMVVB	42-68	190-223	780-807			PENV MLVMO	502-554	566-600				
PENV RMCPV	487-517					PENV MLVRD	487-549	581-585				

PENV 8FV1	14-41	868-801				PENV MLVRK	497-549	581-586					
PENV 8FV3L	18-45	318-357	673-700	863-888		PENV MMTVB	477-539	558-612					
PENV 8IV1	581-588	582-619	652-678	687-724		PENV MMTVG	477-539	558-612					
PENV 8IVAQ	588-593	587-624	688-685	703-730		PENV MPNV	408-474						
PENV 8IVAI	548-603	634-708				PENV MSVFB	43-98	107-141					
PENV 8IVAT	580-617	661-678				PENV OMVVB	22-84	185-223	684-748	780-816			
PENV 8IVCZ	528-584	627-664				PENV RMCV	484-528	540-574					
PENV 8IVGB	588-650	784-816				PENV RGVV	342-376						
PENV 8IVM1	550-609	671-715				PENV 8FV1	1-41	101-140	164-206	321-365	583-651	658-804	
PENV 8IVM2	185-215	277-289				PENV 8FV3L	5-46	158-208	318-357	680-708	883-901		
PENV 8IVMK	583-608					PENV 8IV1	289-310	581-623	643-683				
PENV 8IVML	548-608					PENV 8IVAG	558-628	651-688	808-852				
PENV 8IV84	583-612	642-689	691-718			PENV 8IVAT	287-291	338-370	538-607	627-684	782-840		
PENV 8IV8P	584-585	646-722				PENV 8IVAT	204-288	548-621	644-692	786-833			
PENV 8IVRH	400-462					PENV 8IVCZ	263-291	330-368	612-684	889-703	803-837		
PENV 8RV1	408-471					PENV 8IVGB	588-654	677-725					
PENV 8RV1	773-800					PENV 8IVM1	114-181	488-508	528-613	635-725	809-864		
PENV 8RV1	780-807					PENV 8IVM2	71-116	134-218	245-331				
PENV 8RV1	782-809					PENV 8IVM2	484-505	540-612	638-724				
PENV 8RV1	208-242					PENV 8IVMK	484-505	540-612	638-724				
PENV 8RV1	208-242					PENV 8IVML	484-505	540-612	638-724				
PENV 8RV1	208-242					PENV 8IV84	488-508	517-618	638-728	812-853			
PENV 8RV1	208-242					PENV 8IV8P	470-513	521-620	642-732	811-848			
PENV 8RV1	208-242					PENV 8IVRH	400-488						
PENV 8RV1	387-453					PENV 8RV1	408-476						
PENV 8RV1	371-437					PENV 8RV1	21-82	184-222	637-740	773-808			
PENV 8RV1	381-451					PENV 8RV1	21-82	184-222	643-748	780-816			
PENV 8RV1	381-451					PENV 8RV1	21-82	184-222	648-748	782-818			
PENV 8RV1	381-451					PENV 8RV1	21-82	184-222					
PENV 8RV1	382-441	484-528				PENV 8RV1	208-242						
PENV 8RV1	388-428					PENV 8RV1	208-242						
PENV 8RV1	388-428					PENV 8RV1	208-242						
PENV 8RV1	384-443					PENV 8RV1	208-242						
PENV 8RV1	381-451					PENV 8RV1	380-468						
PENV 8RV1	423-453	488-543				PENV 8RV1	384-440						
PENV 8RV1	387-453					PENV 8RV1	378-454						
PENV 8RV1	418-478					PENV 8RV1	378-454						
PENV 8RV1	381-451					PENV 8RV1	108-142	376-475	494-528				
PENV 8RV1	402-453	508-533				PENV 8RV1	380-452	487-532					
PENV 8RV1	371-437					PENV 8RV1	380-452	487-532					
PENV 8RV1	371-437					PENV 8RV1	377-469	504-649					
PENV 8RV1	371-437					PENV 8RV1	112-148	377-469					
PENV 8RV1	371-437					PENV 8RV1	377-454						
PENV 8RV1	371-437					PENV 8RV1	377-478	495-547					
PENV 8RV1	371-437					PENV 8RV1	380-453						
PENV 8RV1	371-437					PENV 8RV1	378-478	508-548					
PENV 8RV1	418-445					PENV 8RV1	378-454						
PENV 8RV1	387-453					PENV 8RV1	21-55	377-472					
PENV 8RV1	381-451					PENV 8RV1	384-440						



[illegible]

[illegible]

PHEMA PI1HW	70-110	368-393				PHEMA INCP2	471-559
PHEMA PI3B	80-93					PHEMA INCP3	471-559
PHEMA PI3H4	27-61					PHEMA INGTA	471-559
PHEMA PI3HA	27-61					PHEMA INCYA	471-559
PHEMA PI3HT	27-76					PHEMA MEAGE	48-90
PHEMA PI3HU	23-70					PHEMA MEAGH	48-90
PHEMA PI3HV	27-61					PHEMA MEABI	48-87
PHEMA PI3HW	27-61					PHEMA MEABY	48-87
PHEMA PI3HX	27-61					PHEMA MUMPM	34-99
PHEMA RACVI	166-214					PHEMA MUNPR	34-99
PHEMA 8END6	70-106	268-283				PHEMA MUMPB	34-99
PHEMA 8ENDF	70-106					PHEMA NDVA	8-52 477-529
PHEMA 8ENDH	70-106					PHEMA NDVB	1-49
PHEMA 8ENDJ	70-106					PHEMA NDVD	1-49
PHEMA 8ENDZ	70-106					PHEMA NDVM	1-49
PHEMA 8V41	22-62	364-421				PHEMA NDVQ	1-49
PHEMA VACC	119-146	176-202	210-243			PHEMA NDVTG	1-49
PHERA VACCI	108-146	176-202	210-243			PHEMA NDVV	1-49
PHEMA VACTT	119-146	176-202	210-243			PHEMA PHODV	30-73
PHERA VACCV	108-146	176-202	215-242			PHEMA PI1HW	60-110
PVENV DHV11	319-366					PHEMA PI2H	247-281
PVENV EAV	120-147					PHEMA PI2HT	247-281
PVENV THOGV	313-347					PHEMA PI3B	38-93
PVF03 VACCC	71-110	186-212				PHEMA PI3H4	13-110 394-428
PVF03 VACCV	71-110	188-212				PHEMA PI3HA	20-110 394-428
PVF06 VACCP	33-60					PHEMA PI3HT	13-110 394-428
PVF08 VACCV	33-60					PHEMA PI3HU	13-110 394-428
PVE11 VACCC	274-321					PHEMA PI3HV	13-110 394-428
PVF11 VACCP	270-317					PHEMA PI3HW	13-110 394-428
PVF12 VACCC	10-37	113-140	554-581			PHEMA PI3HX	13-110 394-428
PVF12 VACCP	10-37		554-581			PHEMA PI4HA	84-86
PVF16 VACCC	35-62	162-179				PHEMA RACVI	166-214 268-280
PVF16 VACCP	35-62	162-179				PHEMA RINDK	48-87
PVPF4 POWPV	140-173					PHEMA RINDL	48-87 191-226
PVFU8 ORFRZ	50-86					PHEMA 8END6	67-110
PVFUB VACCC	37-64					PHEMA 8ENDP	67-110
PVFUB VACCV	37-64					PHEMA 8ENDH	67-110
PVG01 VACCC	226-262	301-335				PHEMA 8ENDJ	67-110
PVG01 VACCV	104-191	240-274				PHEMA 8ENDZ	87-110
PVG01 VARV	226-262	301-335				PHEMA 8V41	18-52 387-421
PVG02 VACCV	96-123					PHEMA 8V6	27-82
PVG02 VARV	96-123					PHEMA 8V6LN	27-82
PVG03 H8VEB	140-176					PVENV BEV	195-229
PVG03 H8VEK	140-176					PVENV DHV11	318-366
PVG06 VACCC	48-75	131-161	226-269	355-389		PVENV MGV1	262-286
PVG06 VARV	48-75	124-161	255-289	355-389		PVENV MCV2	262-286
PVG07 H8V11	71-86					PVENV THOGV	313-364

PVG09 VACCC	308-338					PVENV VACCC	267-285			
PVG09 VACCV	271-301					PVENV VACCI	267-285			
PVG09 VACCV	308-338					PVENV VACCP	267-285			
PVG09 VARV	11-46					PVENV VAGCV	267-285			
PVG12 BPVIR	177-204					PVF01 VAGCC	46-80	124-168		
PVG17 HSVI	177-204					PVF01 VAGCV	46-80	124-168		
PVG18 HSVI	174-208					PVF03 VACCC	71-110			
PVG18 HSVI	260-287					PVF03 VACCV	71-110			
PVG1 SPVIR	267-314	383-410				PVF06 VACCC	81-129	282-320		
PVG1 SPV4	373-400	591-622	668-706	768-824		PVF06 VACCV	81-129	282-320		
PVG22 HSVI	31-58					PVF05 VACCP	81-129	283-321		
PVG24 HSVI	283-290	497-528				PVF05 VACCV	217-258	268-316		
PVG26 HSVI	33-84	81-118				PVF11 VACCC	213-254	265-311		
PVG28 AMEPV	285-328					PVF11 VACCP	1-67	102-143	198-236	544-581
PVG2 SPVIR	146-173	175-205	262-310			PVF12 VACCC	1-67	102-143	198-236	544-581
PVG2 BPV4	88-122					PVF16 VACCC	155-194			
PVG34 HSVI	442-469					PVF16 VACCV	155-184			
PVG37 HSVI	981-678	1089-1116				PVF16 VACCV	155-184			
PVG39 HSVI	2-29					PVF16 VACCV	155-184			
PVG3L AMEPV	18-49					PVF16 VACCV	155-184			
PVG3 SPVIR	18-52	87-148				PVF16 VACCV	155-184			
PVG3 SPV4	139-165					PVF16 VACCV	155-184			
PVG48 HSVI	142-169	346-373	987-924	973-1007		PVF16 VACCV	155-184			
PVG48 HSVI	360-394					PVF16 VACCV	155-184			
PVG48 HSVI	4-31					PVF16 VACCV	155-184			
PVG4 SPVIR	116-146					PVF16 VACCV	155-184			
PVG51 HSVI	34-61	87-114				PVF16 VACCV	155-184			
PVG62 HSVI	47-74					PVF16 VACCV	155-184			
PVG66 HSVI	582-608					PVF16 VACCV	155-184			
PVG6 SPVIR	68-92					PVF16 VACCV	155-184			
PVG6 SPV4	68-93					PVF16 VACCV	155-184			
PVG6 HSVI	580-584					PVF16 VACCV	155-184			
PVG6 HSVI	477-504					PVF16 VACCV	155-184			
PVG6 HSVI	1213-1264					PVF16 VACCV	155-184			
PVG6 HSVI	362-406					PVF16 VACCV	155-184			
PVG6 HSVI	1342-1368					PVF16 VACCV	155-184			
PVG6 HSVI	261-289					PVF16 VACCV	155-184			
PVG72 HSVI	447-481					PVF16 VACCV	155-184			
PVG75 HSVI	388-422					PVF16 VACCV	155-184			
PVG76 HSVI	200-227					PVF16 VACCV	155-184			
PVG7 SPV4	14-44					PVF16 VACCV	155-184			
PVG1 IBV8	1230-1260	2408-2435				PVF16 VACCV	155-184			
PVG12 CVBF	388-426	842-676	1022-1084	1278-1305		PVF16 VACCV	155-184			
PVG12 CVBL9	399-426		1022-1084	1278-1305		PVF16 VACCV	155-184			
PVG12 CVBL9	389-426	842-676	1022-1084	1278-1305		PVF16 VACCV	155-184			
PVG12 CVBL9	399-426	842-676	1022-1084	1278-1305		PVF16 VACCV	155-184			
PVG12 CVBL9	399-426	842-676	1022-1084	1278-1305		PVF16 VACCV	155-184			
PVG12 CVBL9	399-426	842-676	1022-1084	1278-1305		PVF16 VACCV	155-184			</

PVGL2 CVH22	770-787	808-878	1056-1112		PVG1 SPV4	267-321			
PVGL2 CVMA	843-884	1030-1082			PVG22 HSV11	117-158	437-629	889-1055	
PVGL2 CVMA5	38-63	581-632	978-1040		PVG24 HSV11	7-72	74-108		
PVGL2 CVMAH	502-543	888-851			PVG27 HSV11	164-219			
PVGL2 CYPFS	89-110	882-733	1072-1145	1353-1389	PVG28 HSV11	263-280			
PVGL2 CYPFU	69-107	880-731	1067-1143	1351-1387	PVG2R AMEPV	28-63	184-218		
PVGL2 CYPFR	488-509	845-921	1128-1165		PVG2 8PV1R	222-256	285-328		
PVGL2 CYPFRM	488-509	845-921	1128-1165		PVG2 8PV4	255-310			
PVGL2 EBV	68-102				PVG33 HSV11	149-183			
PVGL2 FIPV	188-233	454-481	709-738	1072-1148	PVG34 HSV11	345-378			
PVGL2 IBV6	808-838	878-903	1057-1081		PVG36 HSV11	17-80			
PVGL2 IBV8	808-835	878-902	1056-1080		PVG37 HSV11	435-472			
PVGL2 IBVD2	808-838	878-903	1057-1081		PVG38 HSV11	84-118			
PVGL2 IBVK	808-838	878-902	1056-1080		PVG38 HSV11	124-158	288-300		
PVGL2 IBVM	808-838	878-902	1056-1080		PVG3 SPV1R	8-49	182-188	203-244	
PVGLB EBV	95-122	931-858			PVG3 8PV4	8-54	87-121		
PVGLB HCMVA	28-88	397-424	440-497	851-878	PVG43 HSV11	116-150	262-296	324-361	843-877
PVGLB HCMVT	50-88	387-424	435-482	852-878	PVG45 HSV8A	121-182			
PVGLB HSB1	427-454				PVG46 HSV11	48-88	938-1078		
PVGLB HSB2	447-474				PVG48 HSV11	188-207			
PVGLB HSB2C	428-453				PVG48 HSB8A	360-417	611-866	733-787	
PVGLB HSBV1	443-470	834-861			PVG49 HSB8A	68-102			
PVGLB HSBV4	443-470	816-843			PVG49 HSB8A	68-102			
PVGLB HSBV5A	443-470	834-861			PVG4R AMEPV	4-38			
PVGLB HSBV6	443-470	834-861			PVG4 8PV4	88-130			
PVGLB HSBV6L	443-470	834-861			PVG51 HSV11	34-73	88-123		
PVGLB HSBVMD	93-120	352-378			PVG51 HSB8A	28-70	123-157	182-196	
PVGLB HCMVB	381-408	441-475			PVG53 HSV11	67-127			
PVGLC H9V11	488-510				PVG54 HSB8A	355-398			
PVGLC H9V1K	488-510				PVG55 HSB8A	101-135			
PVGLC H9VB	124-151				PVG55 HSB8A	128-178			
PVGLC H9VB	63-87				PVG56 HSB8A	151-182	578-612	760-784	848-880
PVGLC H9VMG	62-86				PVG59 HSB8A	10-72	88-123		
PVGLC H9VMH	63-87				PVG59 HSB8A	188-209			
PVGLC H9VMH	63-87				PVG5 8PV1R	65-103			
PVGLC H9VMH	63-87				PVG5 8PV1R	285-299			
PVGLC H9VMH	63-87				PVG53 HSB8A	548-584			
PVGLC H9VMH	63-87				PVG56 HSB8A	1213-1264			
PVGLC H9VMH	63-87				PVG56 HSB8A	154-188	328-410		
PVGLC H9VMH	63-87				PVG56 HSB8A	378-413	501-548	1321-1388	1478-1541
PVGLC H9VMH	63-87				PVG56 HSB8A	245-288			
PVGLC H9VMH	63-87				PVG56 HSB8A	447-484	723-767	912-948	
PVGLC H9VMH	63-87				PVG56 HSB8A	271-305	388-422		
PVGLC H9VMH	63-87				PVG56 HSB8A	5-51			
PVGLC H9VMH	63-87				PVG56 HSB8A	142-178	1233-1287	2118-2156	3478-3513
PVGLC H9VMH	63-87				PVG56 HSB8A	10-44		3388-3424	3617-3556
PVGLC H9VMH	63-87				PVG56 HSB8A				

PVGLF MEAGY	228-262						PVGL2 CVBLY	642-676	850-885	893-1108	1283-1305		
PVGLF MUNPM	20-84	447-486					PVGL2 CVBIM	642-676	850-885	893-1108	1283-1305		
PVGLF MUNPR	20-84	447-486					PVGL2 CVBQ	642-676	850-885	893-1108	1283-1305		
PVGLF MUNP8	181-178	428-511					PVGL2 CVBV	642-676	850-885	893-1108	1283-1305		
PVGLF NDVA	181-178	428-512					PVGL2 CVH22	770-816	1055-1112				
PVGLF NDVB	181-178	428-512					PVGL2 CVM4	843-884	1001-1117	1270-1315			
PVGLF NDVI	181-178	428-512					PVGL2 CVMA6	591-632	849-1079	1218-1263			
PVGLF NDVM	181-178	428-512					PVGL2 CVMAH	592-543	882-976	1128-1174			
PVGLF NDVT	181-178	428-512					PVGL2 CVPF8	98-110	448-482	692-733	889-923	1040-1186	1352-1398
PVGLF NDVTG	181-178	428-512					PVGL2 CVPRU	98-110	448-480	690-731	887-921	1038-1184	1351-1387
PVGLF NDVU	181-178	428-512					PVGL2 CVPR8	224-258	488-509	695-889	818-902	1128-1185	
PVGLF PHOOU	38-63	221-262					PVGL2 CVPRM	224-258	488-509	695-889	818-902	1128-1185	
PVGLF PHHC	147-174	210-268					PVGL2 EBV	68-102					
PVGLF PIZH	80-117	141-175					PVGL2 FIPV	189-245	451-485	695-738	892-926	1043-1189	1355-1392
PVGLF PIZHG	80-117	141-175					PVGL2 IBV8	701-805	1057-1091				
PVGLF PIZHT	80-117	141-175					PVGL2 IBV8	437-478	772-804	1056-1080			
PVGLF PIZ8	115-182	207-241					PVGL2 IBVD2	773-805	1057-1091				
PVGLF PIZ44	115-182	207-241					PVGL2 IBVK	437-478	772-804	1056-1080			
PVGLF RINDK	224-265	458-506					PVGL2 IBVM	437-478	772-804	1056-1080			
PVGLF RINDL	224-265	458-506					PVGLB HCMVA	43-88	128-162	438-484	844-878		
PVGLF REND8	122-149	211-248					PVGLB HCMVT	22-88	128-162	437-485	845-879		
PVGLF RENDF	122-149	211-245					PVGLB H8V11	828-880					
PVGLF RENDH	122-149	211-245					PVGLB H8V1F	827-889					
PVGLF RENDJ	122-149	211-245					PVGLB H8V1K	827-889					
PVGLF RENDZ	122-149	211-245					PVGLB H8V1P	828-890					
PVGLF BV41	144-185	241-289					PVGLB H8V23	828-890					
PVGLF BV6	137-171	417-444					PVGLB H8V2H	828-890					
PVGLF TRTV	124-161	193-200					PVGLB H8V28	817-871					
PVGLB BEFY	823-857						PVGLB H8V8U	37-71	185-223				
PVGLB BR8VC	92-123						PVGLB H8V81	889-913					
PVGLB H8BV1	63-93						PVGLB H8V82	440-474	848-902				
PVGLB H8BV4	68-107						PVGLB H8V8C	863-900					
PVGLB H8BV6	243-273						PVGLB H8VE1	642-676	811-961				
PVGLB H8VB8	68-93						PVGLB H8VE4	474-516	847-900				
PVGLB H8VE4	271-288						PVGLB H8VEA	642-676	811-961				
PVGLB H8VEB	383-410						PVGLB H8VEB	642-676	811-961				
PVGLB RABVT	488-519						PVGLB H8VEL	642-676	810-960				
PVGLB V8VIG	472-499						PVGLB H8VMD	380-435	648-683				
PVGLH EBV	549-578	618-648					PVGLB H8V8A	240-288	409-447				
PVGLH HCMVA	107-136	270-297					PVGLB HCMVB	208-280	427-476				
PVGLH HCMVT	106-136						PVGLB PRVIF	847-881					
PVGLH H8V8G	62-89	360-403					PVGLB VZVD	92-133	588-630	808-867			
PVGLH H8V8A	388-416						PVGLC H8V11	488-510					
PVGLH HCMVA	47-111						PVGLC H8V1K	488-510					
PVGLM BUNGE	512-546	614-841					PVGLC H8V2	442-476					
PVGLM BUNL7	813-850						PVGLC H8V23	443-477					
PVGLM BUNYW	340-374	804-836					PVGLC H8V8C	208-269					

PVGLM DUGBV	845-872						PVGLC HSEVB	182-218				
PVGLM HANTB	73-100	883-720					PVGLC HSNMB	53-97				
PVGLM HANTH	75-102						PVGLC HSNMG	82-86				
PVGLM HANTL	75-102						PVGLC HSNMM	53-97				
PVGLM HANTV	75-102						PVGLC PRVIF	183-236				
PVGLM PHV	88-88						PVGLC VZVD	280-321				
PVGLM PUUMH	72-110						PVGLC VZVB	280-321				
PVGLM PUUMB	72-110						PVGLD HSEVA	88-123				
PVGLM SEOUR	73-100	513-640	694-721				PVGLD HSEVB	139-173				
PVGLM SEOUT	73-100	513-640	694-721				PVGLD HSEVK	139-173				
PVGLN BEV	523-684						PVGLD HSEV1	111-146				
PVGLP BEV	48-82	1145-1178	1184-1211	1506-1532			PVGLD HSEV2	111-158				
PVGLX HSEVB	17-44	413-444					PVGLF BRBVA	148-202	504-548			
PVGLX PRVRI	427-481						PVGLF BRBVC	148-202	287-302	506-547		
PVGLY JUNIN	14-41						PVGLF BRBVR	148-202	287-302	506-554		
PVGLY LABGG	88-113						PVGLF CDVO	228-297	340-381	588-602		
PVGLY MOPEI	88-113	316-346					PVGLF HRSV1	116-203	287-302	506-549		
PVGLY PIARI	334-375						PVGLF HRSVA	116-202	287-302	506-549		
PVGLY TACV	108-136	316-360					PVGLF HRSVL	116-202	287-302	506-547		
PVGLY TACV6	303-338						PVGLF HRSVR	116-202	287-302	506-549		
PVGLY TACV7	302-337						PVGLF MEASE	116-184	228-269	482-500		
PVGLY TACV8	303-338						PVGLF MEAB1	116-187	231-272	485-503		
PVGLZ HSEK	17-44						PVGLF MEAS2	116-184	228-269	482-500		
PVGLM BPMV	403-430						PVGLF MUMPM	20-54	103-178	235-272	447-502	
PVGLM CP8NV	182-221						PVGLF MUMPR	20-54	103-178	235-272	447-502	
PVGLP BEV	106-148						PVGLF MUMPS	20-54	103-178	235-272	447-502	
PVGLM REOVL	280-317						PVGLF NDVA	117-182	231-272	428-512		
PVGLM REOVD	825-882						PVGLF NDVB	122-182	231-272	428-517		
PVGLM REOVD	824-881						PVGLF NDVI	133-182	238-272	428-517		
PVGLM REOVD	824-881						PVGLF NDVM	117-182	231-272	428-517		
PVGLM REOVD	188-186						PVGLF NDVT	117-182	231-272	428-517		
PVGLM REOVD	124-162						PVGLF NDVT8	122-182	231-272	428-517		
PVGLM REOVD	124-161	343-370	458-483	631-680			PVGLF NDVU	122-182	231-272	428-512		
PVGLM REOVD	216-246						PVGLF PHODV	28-63	187-268	308-350	533-581	
PVGLM REOVD	216-246						PVGLF P11HC	123-174	207-267	489-503		
PVGLM REOVD	161-186						PVGLF P12H	83-183	477-528			
PVGLM REOVD	247-274						PVGLF P12H2	83-183	477-528			
PVGLM REOVD	88-123						PVGLF P12HT	83-186	477-528			
PVGLM REOVD	201-231						PVGLF P13B	117-182	207-241	480-518		
PVGLM REOVD	201-231						PVGLF P13H4	117-182	207-241	482-532		
PVGLM REOVD	323-353						PVGLF RINDK	112-180	224-266	448-483		
PVGLM REOVD	176-209						PVGLF RINDL	112-180	224-266	448-508		
PVGLM REOVD	176-209						PVGLF SEND6	127-188	211-271	483-533		
PVGLM REOVD	21-48	184-218					PVGLF SENDF	127-188	211-271	483-533		
PVGLM REOVD	21-48	184-218					PVGLF SENDH	127-188	218-271	483-533		
PVGLM REOVD	21-48	184-218					PVGLF SENDJ	127-188	211-271	483-533		
PVGLM REOVD	184-216						PVGLF SENDZ	127-188	211-271	483-533		

PVMP CAMVC		220-264	273-324			PVGLF 9V41	88-188	484-508			
PVMP CAMVO	28-68	220-264	273-324			PVGLF 9V5	103-171	241-276	451-487		
PVMP CAMVE		227-284	273-324			PVGLF TRTV	105-161	180-224	457-488		
PVMP CAMVN		220-264	273-324			PVGLG BEFV	808-812				
PVMP CAMVS		220-264	273-324			PVGLG BRVC	30-70	104-138			
PVMP CAMVW		220-264	273-324			PVGLG HRSV1	30-81				
PVMP CERV	28-53	100-127				PVGLG HRSV2	30-86				
PVMP SOCMV	4-31	78-118				PVGLG HRSV3	30-95				
PVMSA HPBHE	284-328					PVGLG HRSV4	30-107				
PVMT1 DHV11	38-65	237-284				PVGLG HRSV5	30-85				
PVMT8 MYXVL	163-180					PVGLG HRSV6	30-86				
PVMT9 MYXVL	485-492					PVGLG HRSV7	30-85				
						PVGLG HRSV8	30-81				
						PVGLG HRSVA	30-87				
						PVGLG HRSVL	25-86				
						PVGLG HSE4	271-305				
						PVGLG SIGMA	344-381	464-488			
						PVGLG BYNV	488-523				
						PVGLG VHSV0	383-387				
						PVGLG VSVIG	476-510				
						PVGLH ERV	53-87	160-201	338-380	853-884	
						PVGLH HCMVA	103-137	270-311	693-741		
						PVGLH HCMVT	103-138	682-740			
						PVGLH HSB11	447-481				
						PVGLH HSB1E	447-481				
						PVGLH HSBVG	367-408				
						PVGLH HSBVC	384-418				
						PVGLH HSBVE4	334-378	414-455			
						PVGLH HSBVE8	327-372	407-448			
						PVGLH HSBVA	32-86	374-453	684-712		
						PVGLH MCHV8	440-474				
						PVGLH PRVKA	226-260				
						PVGLH PRVN3	226-260				
						PVGLH PRVRI	226-260				
						PVGLH VZVD	455-508				
						PVGLI HCMVA	47-111	323-359			
						PVGLM BUNGE	612-667	686-737	1228-1262		
						PVGLM BUNL7	643-677	918-950			
						PVGLM BUNSH	643-677				
						PVGLM BUNYW	340-374	504-563	805-838		
						PVGLM DUGBV	937-989	1238-1300			
						PVGLM HANTB	682-727				
						PVGLM HANTH	72-108				
						PVGLM HANTL	72-108				
						PVGLM HANTY	72-109				
						PVGLM PHV	73-111				
						PVGLM PTPV	149-251				



				PVGLM GEOUR	894-728				
				PVGLM GEJUS	683-730				
				PVGLN BEFV	377-414	613-669			
				PVGLP BEV	43-82	80-124	622-656	1128-1238	
				PVGLX HSEB	177-282				
				PVGLX PRVR	420-481				
				PVGLY JUNIN	301-349				
				PVGLY LASSG	317-380	388-422			
				PVGLY LABSJ	316-361	388-423			
				PVGLY LYCVA	333-367	388-432			
				PVGLY LYGVW	124-168	333-367	395-432		
				PVGLY MOPEI	316-369				
				PVGLY PIARV	334-376				
				PVGLY TACV	316-383				
				PVGLY TACVB	303-351	382-416			
				PVGLY TACV7	302-350	381-416			
				PVGLY TACVT	303-351	382-416			
				PVGNB CPMV	835-889				
				PVGNN BPMV	143-177	403-437			
				PVGNN CPMV	160-201				
				PVGNN CP8MV	182-228	758-792	874-916		
				PVGNN RCNV	837-871	912-946			
				PVGP8 EBV	84-149				
				PVM01 VACC	5-56				
				PVM1 REOVL	287-321				
				PVM21 REOVD	416-450	618-683			
				PVM22 REOVD	416-450	618-682			
				PVM2 REOVJ	416-450	618-682			
				PVM2 REOVL	416-450	618-682			
				PVM3 REOVD	135-180	337-371	623-558	618-690	
				PVMA2 BR8VA	42-90				
				PVMA2 HR8VA	42-90				
				PVMAT CDVO	193-234				
				PVMAT INCJJ	73-114	161-208			
				PVMAT NDVA	310-358				
				PVMAT NDVB	324-358				
				PVMAT PISB	88-133	204-262			
				PVMAT PISH4	88-133	204-262			
				PVMAT RABVA	68-103				
				PVMAT RABVC	68-103				
				PVMAT RABVE	68-103				
				PVMAT RABVN	68-103				
				PVMAT RABVP	68-103				
				PVMAT RABVB	68-103				
				PVMAT SYN	246-280				
				PVMAT VEVIQ	189-232				
				PVME1 CVBM	176-209				

[illegible]



**TABLE VI**

Search Results Summary for PCTLZIP,  
P1CTLZIP, and P2CTLZIP Motifs

PCTLZIP	P1CTLZIP	P2CTLZIP
LIBRARY FILE	LIBRARY FILE	LIBRARY FILE
PENV BN06	434-460	525-542
PENV BN27	463-478	564-571
PENV FOAMV	481-496	630-647
PENV HV1MF	183-188	781-788
PENV HV1RH	445-460	776-786
PENV HV18C	186-201	780-787
PENV HV1T2	123-138	824-841
PENV HV1ZH	438-463	605-622
PENV HV2BE	760-785	625-642
PENV HV2D1	741-766	602-619
PENV HV2G1	742-767	710-727
PENV HV2H2	761-786	625-642
PENV HV2S8	743-768	605-622
PENV HV28T	745-760	608-626
PENV J8RV	104-119	123-140
PENV MMTVB	618-633	410-427
PENV MMTVG	618-633	154-171
PENV SIVMK	138-164	760-787
PENV SIVML	138-164	600-617
PHEMA CVBLY	391-406	601-618
PHEMA CVBM	391-406	630-647
PHEMA CVBQ	391-406	625-642
PHEMA CVHOC	391-406	639-656
PHEMA CVMAS	402-417	639-656
PHEMA CVM5	403-418	626-643
PHEMA INBAA	286-310	167-184
PHEMA INBBE	303-318	629-646
PHEMA INBRO	283-308	624-641
PHEMA INBEN	301-316	624-641
PHEMA INBFU	286-301	170-187
PHEMA INBGL	286-311	603-620
PHEMA INBHK	283-308	603-620
PHEMA INBIB	289-303	887-874
PHEMA INBID	289-314	854-871
PHEMA INBLE	302-317	707-724
PHEMA INBMD	282-307	768-783
PHEMA INBME	286-311	786-782
PHEMA INBNA	288-303	764-781
PHEMA INBOR	301-316	788-786
PHEMA INBSI	301-316	773-780
PHEMA INBSJ	286-313	638-653
PHEMA INBUS	284-309	638-653
PHEMA INBVI	286-311	42-59
PHEMA INBVK	303-318	200-217
PHEMA INBVB	286-301	38-53
PHEMA IAAIC	237-253	391-408
PHEMA IABIC	237-253	391-408
PHEMA IACIC	237-253	391-408
PHEMA IADIC	237-253	391-408
PHEMA IAEIC	237-253	391-408
PHEMA IAFIC	237-253	391-408
PHEMA IAGIC	237-253	391-408
PHEMA IAHIC	237-253	391-408
PHEMA IAIIC	237-253	391-408
PHEMA IAJIC	237-253	391-408
PHEMA IAKIC	237-253	391-408
PHEMA IALIC	237-253	391-408
PHEMA IAMIC	237-253	391-408
PHEMA IANIC	237-253	391-408
PHEMA IAOIC	237-253	391-408
PHEMA IAPIC	237-253	391-408
PHEMA IAAIC	237-253	391-408
PHEMA IABIC	237-253	391-408
PHEMA IACIC	237-253	391-408
PHEMA IADIC	237-253	391-408
PHEMA IAEIC	237-253	391-408
PHEMA IAFIC	237-253	391-408
PHEMA IAGIC	237-253	391-408
PHEMA IAHIC	237-253	391-408
PHEMA IAIIC	237-253	391-408
PHEMA IAJIC	237-253	391-408
PHEMA IAKIC	237-253	391-408
PHEMA IALIC	237-253	391-408
PHEMA IAMIC	237-253	391-408
PHEMA IANIC	237-253	391-408
PHEMA IAOIC	237-253	391-408
PHEMA IAPIC	237-253	391-408
PHEMA IAAIC	237-253	391-408
PHEMA IABIC	237-253	391-408
PHEMA IACIC	237-253	391-408
PHEMA IADIC	237-253	391-408
PHEMA IAEIC	237-253	391-408
PHEMA IAFIC	237-253	391-408
PHEMA IAGIC	237-253	391-408
PHEMA IAHIC	237-253	391-408
PHEMA IAIIC	237-253	391-408
PHEMA IAJIC	237-253	391-408
PHEMA IAKIC	237-253	391-408
PHEMA IALIC	237-253	391-408
PHEMA IAMIC	237-253	391-408
PHEMA IANIC	237-253	391-408
PHEMA IAOIC	237-253	391-408
PHEMA IAPIC	237-253	391-408
PHEMA IAAIC	237-253	391-408
PHEMA IABIC	237-253	391-408
PHEMA IACIC	237-253	391-408
PHEMA IADIC	237-253	391-408
PHEMA IAEIC	237-253	391-408
PHEMA IAFIC	237-253	391-408
PHEMA IAGIC	237-253	391-408
PHEMA IAHIC	237-253	391-408
PHEMA IAIIC	237-253	391-408
PHEMA IAJIC	237-253	391-408
PH		

PHEMA MUMPM	133-148		PHEMA IABN	221-237			PHEMA CVHOC	391-408
PHEMA MUMPR	133-148		PHEMA IABUD	234-260			PHEMA IAAIC	322-339
PHEMA MUMPB	133-148		PHEMA IACKA	234-260			PHEMA IABAN	306-323
PHEMA PI1HW	345-360		PHEMA IACKG	231-247			PHEMA IABUD	320-337
PHEMA PI2H	65-80		PHEMA IACKV	230-246			PHEMA IACKA	320-337
PHEMA PI2HT	65-80		PHEMA IADA1	234-260			PHEMA IACKG	316-333
PHEMA RUNDK	368-383		PHEMA IADA3	237-253			PHEMA IACKP	302-319
PHEMA SV5	7-84		PHEMA IADCZ	234-260			PHEMA IACKG	302-318
PHEMA SV5CM	7-84		PHEMA IADH1	221-237			PHEMA IACKV	315-332
PHEMA SV5CP	7-84		PHEMA IADH2	221-237			PHEMA IADH1	320-337
PHEMA SV5LN	7-84		PHEMA IADH3	221-237			PHEMA IADH3	322-339
PVENV DHV11	42-87		PHEMA IADH4	221-237			PHEMA IADH3	320-337
PVRP7 CAPVK	60-104		PHEMA IADH5	221-237			PHEMA IADH1	306-323
PVRJ8 VACC6	72-87		PHEMA IADH6	221-237			PHEMA IADH2	306-323
PVG01 BPP22	242-257		PHEMA IADH7	221-237			PHEMA IADH2	306-323
PVG01 HSBV2	169-184		PHEMA IADH2	237-253			PHEMA IADH3	306-323
PVG01 HSBV1	210-225	317-332	PHEMA IADH2	234-260			PHEMA IADH4	306-323
PVG06 BPT4	184-198		PHEMA IADH6	221-237			PHEMA IADH6	306-323
PVG07 BPT4	686-900		PHEMA IADH7	237-253			PHEMA IADH7	306-323
PVG08 HSBV1	134-149		PHEMA IADH7	230-246			PHEMA IADM2	322-338
PVG10 BPH2	183-198		PHEMA IAHAL	236-252			PHEMA IADN2	320-337
PVG10 BPP2A	183-198		PHEMA IAHAR	235-251			PHEMA IADN2	322-339
PVG10 HSBVA	108-124		PHEMA IAHCB	230-246			PHEMA IADN6	306-323
PVG16 BPP1	81-86		PHEMA IAHCB	230-246			PHEMA IADN7	322-339
PVG19 BPT4	468-483		PHEMA IAHCB	230-246			PHEMA IADN7	315-332
PVG25 BPT4	97-112		PHEMA IAHCB	230-246			PHEMA IADN7	320-337
PVG29 HSBV1	20-36		PHEMA IAHFO	236-252			PHEMA IAGU2	320-337
PVG30 BPPH8	11-84		PHEMA IAHK6	236-252			PHEMA IAGUA	318-336
PVG38 BPOX2	22-37		PHEMA IAHK7	236-252			PHEMA IAHAL	321-338
PVG39 HSBVA	108-123		PHEMA IAHLE	230-246			PHEMA IAHCB	315-332
PVG37 BPT2	1253-1268		PHEMA IAHLO	230-246			PHEMA IAHCB	315-332
PVG37 HSBV1	284-289		PHEMA IAHMI	236-252			PHEMA IAHCB	315-332
PVG55 HSBV1	22-37	143-168	PHEMA IAHNM	236-252			PHEMA IAHDE	321-338
PVG59 HSBV1	288-293		PHEMA IAHRO	236-252			PHEMA IAHFO	321-338
PVG59 HSBV1	102-117		PHEMA IAHBA	236-252			PHEMA IAHK6	321-338
PVG59 HSBV1	207-282		PHEMA IAHBP	230-246			PHEMA IAHK7	321-338
PVG59 HSBV1	618-533		PHEMA IAHBW	230-246			PHEMA IAHLE	318-332
PVG8 BPPH2	234-249		PHEMA IAHTE	236-252			PHEMA IAHLO	316-332
PVG8 BPP2A	234-249		PHEMA IAHTO	236-252			PHEMA IAHMI	321-338
PVG8 BPP1R	67-72		PHEMA IAHUR	236-252			PHEMA IAHNM	321-338
PVG8 BPPHX	234-249		PHEMA IAKIE	236-251			PHEMA IAHNM	316-332
PVG12 CVBF	284-279		PHEMA IALEN	236-251			PHEMA IAHPR	315-332
PVG12 CVBL9	284-279		PHEMA IAMAA	233-249			PHEMA IAHRO	321-338
PVG12 CVBLY	284-279		PHEMA IAMAB	238-254			PHEMA IAHSA	321-338
PVG12 CVBM	284-279		PHEMA IAMAO	237-253			PHEMA IAHSP	316-332
PVG13 CVBQ	284-279		PHEMA IAME1	237-253			PHEMA IAHBW	315-332
PVG13 CVBV	284-279		PHEMA IAME2	237-253			PHEMA IAHTE	321-338

PVGL2 CVPR8	442-467		PHEMA IAME8	221-237			PHEMA IAHTO	321-338
PVGL2 CVPR8	440-455	504-518	PHEMA IANIN	85-101	231-247		PHEMA IAHUR	321-338
PVGL2 CVPR8	218-233		PHEMA IANT8	237-263			PHEMA IAJAP	317-334
PVGL2 CVPRM	218-233		PHEMA IAOU7	221-237			PHEMA IAMAA	318-336
PVGL2 BV8	1068-1071		PHEMA IARUD	234-260			PHEMA IAMAB	324-341
PVGL2 BV8	1065-1070		PHEMA IASE2	234-260			PHEMA IAMAO	322-338
PVGL2 BVD2	1068-1071		PHEMA IASH2	234-260			PHEMA IAME1	322-339
PVGL2 BVK	1065-1070		PHEMA IASTA	230-246			PHEMA IAME2	322-339
PVGL2 BYM	1066-1070		PHEMA IATAI	235-261			PHEMA IAME8	308-323
PVGLB HSYBA	701-716		PHEMA IATKM	234-260			PHEMA IAMIN	316-333
PVGLB PRVF	203-218		PHEMA IATKO	233-248			PHEMA IANT8	322-339
PVGLC HSYBC	475-480		PHEMA IATKR	230-246			PHEMA IAPIL	320-337
PVGLC HSYE4	444-459		PHEMA IATKW	228-245			PHEMA IAOU7	308-323
PVGLC HSYEB	427-442		PHEMA IAUDD	237-263			PHEMA IARUD	320-337
PVGLC PRVF	448-461		PHEMA IAUSS	238-261			PHEMA IABE2	320-337
PVGLD HSY11	78-84		PHEMA IAV7	238-264			PHEMA IABH2	321-338
PVGLD HSY2	78-84		PHEMA IAXIA	238-261			PHEMA IABTA	316-332
PVGLF BRVA	285-280		PHEMA IAZCO	237-263			PHEMA IATKM	320-337
PVGLF BRVC	285-280		PHEMA IAZH2	221-237			PHEMA IAUDD	322-339
PVGLF BRVR	285-280		PHEMA IAZH3	221-237			PHEMA IAV7	323-340
PVGLF HRSV1	285-280		PHEMA IAZUK	237-263			PHEMA IAZCO	322-339
PVGLF HRSVL	285-280		PHEMA INBAA	116-131	285-310		PHEMA IAZH2	305-323
PVGLF HRSVR	285-280		PHEMA INBEE	123-139	303-318		PHEMA IAZH3	305-323
PVGLF HRSVR	285-280		PHEMA INBEO	116-132	289-308		PHEMA IAZUK	322-339
PVGLF HRSVR	285-280		PHEMA INBEN	123-139	301-318		PHEMA MUMPM	101-118
PVGLI VZVO	278-283		PHEMA INBFU	108-124	288-301		PHEMA MUMPR	101-118
PVGLM HANTB	900-915		PHEMA INBGL	118-135	288-311		PHEMA MUMPS	101-118
PVGLM PTFV	743-758		PHEMA INBHK	118-132	283-308		PHEMA NDVA	83-110
PVGLM SEOUR	901-916		PHEMA INBIB	108-124	288-303		PHEMA NDVB	83-110
PVGLM SEOUT	900-915		PHEMA INBID	120-136	288-314		PHEMA NDVO	83-110
PVGLY LA88G	428-441		PHEMA INBLE	123-139	302-317		PHEMA NDVH	83-110
PVGLY LA88J	427-442		PHEMA INBMD	113-128	282-307		PHEMA NDVI	83-110
PVGLY MOPEI	425-440		PHEMA INBME	116-132	286-311		PHEMA NDVM	83-110
PVM3 REOVD	521-536		PHEMA INBNA	108-124	288-303		PHEMA NDVO	83-110
PVMSA HPB08	380-395		PHEMA INBOR	123-139	301-318		PHEMA NDVTG	83-110
PVMSA HPBV8	187-202		PHEMA INBBI	123-139	301-318		PHEMA NDVU	83-110
PVMSA WHV1	378-393		PHEMA INBSJ	118-135	288-313		PHEMA PHODY	38-63
PVMSA WHV59	383-398		PHEMA INBUS	116-132	284-308		PHEMA PI1HW	488-503
PVMSA WHV7	383-398		PHEMA INBVI	116-132	288-311		PHEMA PI3B	111-128
PVMSA WHV8	383-398		PHEMA INBVK	123-139	303-318		PHEMA PI3H4	111-128
PVMSA WHV8I	383-398		PHEMA INBYB	108-124	288-301		PHEMA PI3HA	111-128
PVMSA WHVW6	234-249		PHEMA MUMPM	133-148			PHEMA PI3HT	111-128
PVMT2 IANN	26-40		PHEMA MUMPR	133-148			PHEMA PI3HU	111-128
PVMT2 IABN	26-40		PHEMA MUMPS	133-148			PHEMA PI3HV	111-128
PVMT2 IAFOW	26-40		PHEMA PI1HW	346-380			PHEMA PI3HW	111-128
PVMT2 IAFPR	26-40		PHEMA PI2H	65-81			PHEMA PI3HX	111-128
PVMT2 IAPFW	26-40		PHEMA PI2HT	65-81			PHEMA PI4HA	50-67

PVMT2 IALE1	26-40	PHEMA P13B	324-340			PHEMA 8V41	86-102
PVMT2 IALE2	25-40	PHEMA P13H4	324-340			PHEMA 8V6	84-101
PVMT2 IAMAN	26-40	PHEMA P13HA	324-340			PHEMA 8V6CM	84-101
PVMT2 IAPUE	26-40	PHEMA P13HT	324-340			PHEMA 8V6CP	84-101
PVMT2 IASIN	26-40	PHEMA P13HU	324-340			PHEMA 8V6LN	84-101
PVMT2 IAUDO	26-40	PHEMA P13HV	324-340			PVF05 VACCC	280-287
PVMT2 IAWIL	26-40	PHEMA P13HW	324-340			PVF06 VACCP	280-287
PVMT8 MYXVL	226-241	PHEMA P13HX	324-340			PVF06 VACCY	281-288
		PHEMA RINDK	368-383			PVF08 VACCC	176-183
		PHEMA 8V6	7-94			PVF08 VACCV	176-183
		PHEMA 8V6CM	7-94			PVF08 VACCC	176-183
		PHEMA 8V6CP	7-94			PVG27 HSV8A	208-228
		PHEMA 8V6LN	7-94			PVG28 H6V11	173-180
		PVENV DNV11	42-57			PVG38 H8V11	648-665
		PVENV EAV	26-41			PVG43 H8V11	108-126
		PVFTZ FOWPV	88-104			PVG67 H8V11	171-188
		PVPF7 CAPVK	88-104			PVG72 H8V11	1252-1269
		PVRUS VACC8	72-87			PVGF1 IBVB	3073-3080
		PVG01 H8VEB	168-184			PVGL2 IBV6	1084-1111
		PVG01 H8V11	208-225			PVGL8 H8VE1	736-753
		PVG08 H8V11	134-149		317-332	PVGL8 H8VE4	676-692
		PVG10 H8V8A	108-124			PVGL8 H8VEA	736-753
		PVG11 H8V11	103-119			PVGL8 H8VEB	736-753
		PVG12 H8V11	270-286			PVGL8 H8VEL	736-753
		PVG1 8PV1R	76-92			PVGL8 ILTV8	597-614
		PVG28 H8V11	20-35			PVGL8 ILTV8	607-624
		PVG36 BPOX2	22-37			PVGLC PRVT	180-197
		PVG36 H8V8A	108-123			PVGLC VZVD	489-488
		PVG37 H8V11	284-289			PVGLF 8V6	401-418
		PVG41 H8V11	244-260			PVGLH HCMVA	385-382
		PVG46 H8V11	1244-1260			PVGLH HCMVT	394-381
		PVG55 H8V11	22-37		143-158	PVGLH H8V11	245-282
		PVG66 H8V11	288-283			PVGLH H8V1E	245-262
		PVG68 H8V11	101-117			PVGLI H8V11	43-60
		PVG68 H8V8A	130-146			PVGLM BUNL7	81-86
		PVG68 H8V11	287-282		330-346	PVGLM BUN8H	81-86
		PVG68 H8V11	362-378		518-533	PVGLM PUUMH	712-729
		PVG71 H8V8A	89-105			PVGLM PUUMS	712-729
		PVG8 BPPH2	234-249			PVGLM RVFV	344-361
		PVG8 BPPZA	234-249			PVGLM RVFVZ	344-381
		PVG8 8PV1R	57-72			PVGLY LA88G	12-94
		PVGF1 IBVB	1210-2226			PVGLY LA88J	12-94
		PVGL2 CVBF	123-139		174-190	PVGLY LYCVA	12-94
		PVGL2 CVBL9	123-139		174-190	PVGLY LYCVW	12-94
		PVGL2 CVBLY	123-139		174-190	PVGLY MOPEI	12-94
		PVGL2 CVBM	123-139		174-190	PVGLY REOVD	280-287
		PVGL2 CVBQ	31-47		123-139	PVM1 REOVL	280-287
					174-190		
					284-279		
					284-278		
					284-279		
					284-279		
					174-190		
					123-139		
					31-47		
					294-279		



	PVGL2 CVBV	123-139	174-180	264-279	PVMAT CDVO	148-166
	PVGL2 CVM4	95-111	1287-1288		PVMAT MEAB1	87-104
	PVGL2 CVM46	95-111	1216-1231		PVMP CAMVC	147-184
	PVGL2 CVMJH	95-111	1126-1142		PVMP CAMVD	147-184
	PVGL2 CVPF9	442-467	600-816	1274-1290	PVMP CAMVE	147-184
	PVGL2 CVPFU	440-455	504-518	788-814	PVMP CAMVN	147-184
	PVGL2 CVPR8	218-233	578-592	1050-1088	PVMP CAMVS	147-184
	PVGL2 CVPRM	218-233	578-592	1050-1088	PVMP CAMVW	147-184
	PVGL2 FIPV	803-819	1277-1283		PVMSA HPSBO	11-94
	PVGL2 IBV6	1058-1071			PVMSA HPSB2	185-202
	PVGL2 IBV8	1058-1070			PVMSA HPSB4	185-202
	PVGL2 IBV02	1058-1071			PVMSA HPSBA	174-181
	PVGL2 IBVK	1058-1070			PVMSA HPSBD	11-94
	PVGL2 IBVM	1058-1070			PVMSA HPSBJ	174-181
	PVGLB HSB8A	701-716			PVMSA HPSBL	11-94
	PVGLB PRVF	203-218			PVMSA HPSBN	11-94
	PVGLB VZVD	622-638			PVMSA HPSBO	174-181
	PVGLC HSBVC	476-480			PVMSA HPSBP	185-202
	PVGLC HSVE4	444-468			PVMSA HPSBR	185-202
	PVGLC HSVEB	427-442			PVMSA HPSBS	11-94
	PVGLC PRVF	446-461			PVMSA HPSBW	174-181
	PVGLC VZVD	150-166			PVMSA HPSBY	174-181
	PVGLG VZVB	150-166			PVMSA HPSBZ	174-181
	PVGLD H8V11	76-84			PVMT2 IANNN	28-42
	PVGLD H8V2	76-84			PVMT2 IABAN	28-42
	PVGLF PRVR1	3-94			PVMT2 IAFOW	28-42
	PVGLF BR8VA	205-221	265-280		PVMT2 IAFPR	26-42
	PVGLF BR8VC	205-221	265-280		PVMT2 IAFPW	26-42
	PVGLF BR8VR	205-221	265-280		PVMT2 IALE1	26-42
	PVGLF COVO	398-414			PVMT2 IALE2	26-42
	PVGLF HR8V1	205-221	265-280		PVMT2 IAMAN	26-42
	PVGLF HR8VA	205-221	265-280		PVMT2 IAPUE	26-42
	PVGLF HR8VL	205-221	265-280		PVMT2 IASIN	26-42
	PVGLF HR8VR	205-221	265-280		PVMT2 IAUDO	26-42
	PVGLF MEABE	266-302			PVMT2 IAWIL	26-42
	PVGLF MEAB1	289-305				
	PVGLF MEASY	286-302				
	PVGLF MUMPM	276-292				
	PVGLF MUMPR	276-292				
	PVGLF MUMPS	6-94	276-292			
	PVGLF NDVA	273-289				
	PVGLF NDVB	273-289				
	PVGLF NDVM	273-289				
	PVGLF NDVT	273-289				
	PVGLF NDTVQ	273-289				
	PVGLF NDVV	273-289				
	PVGLF P4ODV	268-285	367-383			

PVGLF RINDK	282-398
PVGLF RINDL	282-268
PVGLF TRTV	176-181
PVGLI VZVD	278-283
PVGML HANTB	355-371
PVGML HANTR	490-515 900-916
PVGML HANTL	499-515
PVGML HANTV	499-515
PVGML PTPV	743-758
PVGML PUUMH	509-528
PVGML PUJMS	509-525
PVGML SECUR	355-371 901-916
PVGML SEOUS	355-371 900-916
PVGML UIUK	825-842
PVGLP BEV	869-895
PVGLY LASSG	12-84 426-441
PVGLY LASSJ	12-84 427-442
PVGLY LYCVA	12-84
PVGLY LYCVW	12-84
PVGLY MOPEI	12-84
PVGLY PIARV	12-84 425-440
PVGNM CPMV	1021-1037
PVM3 REOVD	621-638
PVMAT MUMP8	181-207
PVMAT NDVA	135-151
PVMAT NOV8	135-151
PVMAT PIZHT	189-205
PVMAT SV41	189-205
PVMAT SVS	99-114 132-148
PVMP CAMYC	118-134
PVMP CAMYD	118-134
PVMP CAMYE	118-134
PVMP CAMVN	118-134
PVMP CAMV8	118-134
PVMP CAMYW	118-134
PVMP FMVD	116-131
PVMSA HPBG3	380-395
PVMSA HF8V9	187-202
PVMSA WHV1	376-393
PVMSA WHV59	383-398
PVMSA WHV7	383-398
PVMSA WHVB	383-398
PVMSA WHV81	383-398
PVMSA WHVV6	234-249
PVMT2 IANN	28-40
PVMT2 JABN	25-40
PVMT2 JAFOW	26-40

[illegible]

**TABLE VII**

**Search Results Summary for P3CTLZIP, P4CTLZIP,  
P5CTLZIP, and P6CTLZIP Motifs**

[illegible]

PVMD1 VACCV	83-101	128-144	PVGL2 CVMA4	888-1018	PVENV THGV	358-378	PHEMA P12H	13-34	
PVM1 REOVD	227-245		PVGL2 CVMA5	847-886	PVG01 VACCC	288-318	PHEMA P12HT	13-34	
PVM1 REOVL	227-245		PVGL2 CVMLH	858-877	PVG01 VACCV	237-267	PHEMA SV6	7-28	378-400
PVMAT HRBVA	44-62		PVGL2 CVPF8	84-83	PVG01 VAR	288-318	PHEMA SV5CM	7-28	378-400
PVMAT NDVA	180-208		PVGL2 CVPPU	84-83	PVG08 VACCC	31-51	PHEMA SV6CP	7-28	378-400
PVMAT NDVB	180-208		PVGL2 CVPRB	814-833	PVG08 VAR	31-51	PHEMA SV6LN	7-28	378-400
PVMP CAMVC	183-201		PVGL2 CVPRM	814-833	PVG08 BPPF1	25-45	PVG01 HSEVB	188-190	
PVMP CAMVD	183-201		PVGL2 FIPV	1041-1080	PVG12 HSNV1	151-171	PVG01 HSNV1	589-610	
PVMP CAMVE	183-201		PVGL2 IBV6	588-607	PVG22 HSNV1	300-320	PVG23 HSNV1	314-335	
PVMP CAMV8	183-201		PVGL2 IBV8	587-606	PVG38 HSNV1	848-868	PVG37 BPOX2	66-88	
PVMP CAMVW	183-201		PVGL2 IBVD2	586-607	PVG51 HSNV1	28-49	PVG43 HSNV1	167-178	
PVMP CAMVD	180-188		PVGL2 IBVK	587-606	PVG83 HSNV1	338-358	PVG55 HSNV1	288-308	
			PVGL2 IBVM	587-608	PVG88 HSNV1	117-137	PVG55 HSNV8A	65-108	
			PVGLB HCMVA	705-725	PVG74 HSNV8A	124-144	PVG58 HSNV1	1155-1176	
			PVGLB HCMVT	707-728	PVGL2 IBV8	328-348	PVG58 HSNV8A	268-287	
			PVGLB HSNV8U	117-138	PVGL2 IBV8	327-347	PVG80 HSNV1	30-51	
			PVGLB ILTV8	268-276	PVGL2 IBVD2	328-348	PVG83 HSNV1	238-259	
			PVGLB ILTV8	268-285	PVGL2 IBVD3	328-348	PVG80 HSNV1	1886-1877	
			PVGLB ILTVT	268-286	PVGL2 IBVK	327-347	PVG83 HSNV1	167-178	
			PVGLC HSNV11	3-84	PVGL2 IBVM	327-347	PVG83 HCMVA	1259-1280	
			PVGLC HSNV1K	3-84	PVGL2 IBVU2	310-330	PVGL2 CVBL9	1259-1280	
			PVGLC HSNVBC	475-484	PVGLB EBV	732-762	PVGL2 CVBL9	1259-1280	
			PVGLC CHAV	436-456	PVGLB HCMVA	750-770	PVGL2 CVBM	1259-1280	
			PVGLG RABVH	373-381	PVGLB HCMVT	751-771	PVGL2 CVBQ	1259-1280	
			PVGLI HSEVB	44-83	PVGLB HSNV23	79-98	PVGL2 CVBV	1259-1280	
			PVGLI VZVO	278-287	PVGLB HSNV2H	79-98	PVGL2 CVMA4	1317-1338	
			PVGLM BUNGE	117-136	PVGLB HSNV28	65-86	PVGL2 CVMA5	1285-1288	
			PVGLM PHV	152-171	PVGLB HSNV6U	72-92	PVGL2 CVMLH	1176-1187	
			PVGLM PTPV	987-1016	PVGLB HSNV82	279-289	PVGLB HSNV11	83-104	
			PVGLM PUJNH	158-174	PVGLB HSNV8A	83-93	PVGLB HSNV1F	82-103	
			PVGLM PUJMS	158-174	PVGLB MCMV8	738-768	PVGLB HSNV1K	82-103	
			PVGLM RVFV	830-849	PVGLF P13H4	283-303	PVGLB HSNV1P	83-104	
			PVGLM RVFVZ	830-849	PVGLG RABVE	484-474	PVGLB MCMV9	138-158	
			PVGLM UUK	658-674	PVGLG RABVH	484-474	PVGLC PRVF	448-467	
			PVGLY LYCVW	89-108	PVGLG RABVP	484-474	PVGLF CDVO	338-357	
			PVGNB CPMV	1185-1194	PVGLG RABV8	484-474	PVGLF MEABE	224-245	
			PVM3 REOVD	521-540	PVGLG RABVT	484-474	PVGLF MEAB1	227-248	
			PVME1 CVBM	171-190	PVGLH MCMV8	870-890	PVGLF MEASY	224-245	
			PVME1 CVH22	136-155	PVGLM BUNL7	1326-1346	PVGLF MUMPM	448-467	
			PVME1 CVPF8	174-183	PVGLM BUNBH	898-1016	PVGLF MUMPR	448-467	
			PVME1 CVPPU	174-183	PVGLM BUNYW	898-1016	PVGLF MUMPS	448-467	
			PVME1 CVPRM	174-183	PVGLM HANTB	898-1018	PVGLF PHODV	305-326	
			PVME1 CVTKE	171-180	PVGLM HANTH	1000-1020	PVGLF PIHC	458-477	
					PVGLM HANTL	1001-1021	PVGLF PIH	450-471	
					PVGLM HANTV	1001-1021	PVGLF PI2HG	450-471	
					PVGLM RVFVZ	1158-1178	PVGLF PI2HT	450-471	
					PVGLM SEOUR	1000-1020	PVGLF PI3B	408-428	453-474

	PVGLM SEQUS	98B-101B	PVGLF PL3H4	463-474
	PVGLM UIK	926-946	PVGLF RINDK	220-241
	PVGLY LYCYA	12-32	PVGLF RINDL	220-241
	PVGLY LYCVW	12-32	PVGLF SENDS	460-481
	PVGLY PIARV	12-32	PVGLF SENDF	460-481
	PVGNB CPMV	141-181	PVGLF SENDD	460-481
	PVMAT MUMPS	310-330	PVGLF SENDJ	460-481
	PVMAT NDVA	308-328	PVGLF SENDZ	460-481
	PVMAT NDVB	308-328	PVGLF SV41	463-474
	PVMAT PIZHT	308-328	PVGLF SVB	448-467
	PVMAT PI4HA	312-332	PVGLH HCMVA	981-712
	PVMAT PI4HB	312-332	PVGLH HCMVT	980-711
	PVMAT SV41	308-328	PVGLH HSVE4	304-325
	PVMAT SV5	308-328	PVGLH HSVEB	287-318
	PVME1 IBV6	74-84	PVGLH HSV8A	658-679
	PVME1 IBV8	74-84	PVGLI HSV2	2-23
	PVME1 IBV82	74-84	PVGLI HSV23	2-23
	PVME1 IBVK	74-84	PVGLM BUNGE	187-218
	PVMSA HPBD8	201-221	PVGLM BUNL7	180-211
	PVMSA HPBG8	208-228	PVGLM BUNSH	180-211
	PVMSA HPBHE	289-313	PVGLM BUNYW	183-214
	PVMSA WHV1	207-227	PVGLY LABEG	237-258
	PVMSA WHV69	212-232	PVGLY LAGSJ	238-259
	PVMSA WHV7	212-232	PVGP8 EBV	87-88
	PVMSA WHV8	212-232	PVM01 VACCC	281-302
	PVMSA WHVB1	212-232	PVM01 VACCV	230-251
	PVMSA WHVWB	63-83	PVMAT HRSA	189-180
			PVMAT RINDK	200-221
			PVMAT TRTV	122-143
			PVME1 CVHOC	84-85
			PVMSA HPBD8	201-222
			PVMSA HPBV0	70-81
			PVMSA HPBV2	244-265
			PVMSA HPBV4	244-265
			PVMSA HPBV9	244-265
			PVMSA HPBVA	233-254
			PVMSA HPBVD	70-81
			PVMSA HPBV1	233-254
			PVMSA HPBVJ	233-254
			PVMSA HPBVL	233-254
			PVMSA HPBVN	70-81
			PVMSA HPBVO	233-254
			PVMSA HPBPV	244-286
			PVMSA HPBVR	244-285
			PVMSA HPBS	70-81
			PVMSA HPBVW	233-254
			PVMSA HPBVY	233-254

[illegible]



**TABLE VIII**

Search Results Summary for P7CTLZIP,  
P8CTLZIP, and P9CTLZIP Motifs

[illegible]

PHEMA IAVI7	38-60		PVGL2 IBVK	196-218	PVGLB HSMVD	588-613		
PHEMA IAX31	37-59		PVGL2 IBVM	195-218	PVGLB ILTV8	597-621		
PHEMA IAZCO	37-59		PVGL2 IBVU1	178-201	PVGLB ILTV8	607-631		
PHEMA IAZH2	21-43		PVGL2 IBVU2	178-201	PVGLB ILTVT	607-631		
PHEMA IAZH3	21-43		PVGL2 IBVU3	178-201	PVGLB HSMV11	413-437		
PHEMA IAZUK	37-59		PVGLB HCMVA	535-558	PVGLB VZVD	489-483		
PHEMA PHODV	38-68		PVGLB HCMVT	535-558	PVGLF SV6	401-425		
PHEMA PIZH	68-87		PVGLB HSMVA	493-508	PVGLB HCMVA	574-588		
PHEMA PIZHT	68-87		PVGLB HCMV8	588-589	PVGLB HCMVT	573-587		
PVPF7 CAPVK	88-111		PVGLC HSMV8	487-480	PVGLB HSMV11	443-487	803-827	
PVFUS VACC8	72-84		PVGLC HSMV1K	487-480	PVGLB HSMV1E	443-487	803-827	
PVG01 HSMV1	317-338		PVGLC HSMV2	435-458	PVGLB BUNL7	31-55		
PVG03 VACC	50-72		PVGLC HSMV23	435-458	PVGLM BUNSH	31-55		
PVG03 VARV	50-72		PVGLM BUNL7	1387-1410	PVGLM HANTH	694-718		
PVG04 VACC	11-33		PVGLM BUNSH	1387-1410	PVGLM RVFV	344-368		
PVG04 VARV	11-33		PVGLM UUK	886-889	PVGLM RVFVZ	344-368		
PVG18 HSMV1	68-110		PVGLY JUNIN	12-35	PVGLM UUK	561-585		
PVG28 HSMV1	173-186		PVGLY LAS8Q	12-35	PVGLM CPNV	311-335		
PVG28 HSMV1	20-42		PVGLY LAS8J	12-35	PVGP2 EBV	667-681		
PVG48 HSMV1	134-156		PVGLY LYCVA	12-35	PVGP3 EBV	654-678		
PVG48 HSMVA	71-93		PVGLY LYCVW	12-35	PVM1 REVD	280-304		
PVG58 HSMVA	266-288		PVGLY MOPEI	12-35	PVM1 REOVL	280-304		
PVG58 HSMV1	267-288		PVGLY TACV	12-35	PVM21 REOVD	168-192		
PVG5 8PV4	42-64		PVGLY TACV6	12-35	PVM22 REOVD	168-192		
PVG60 HSMV1	53-75		PVGLY TACV7	12-35	PVM2 REOVL	168-192		
PVG65 HSMV1	1347-1369		PVGLY TACVT	12-35	PVM2 REOVL	168-192		
PVG8 8PV1R	60-82		PVGLM CPNV	741-764	PVMAT MEAB1	97-111		
PVGL2 IBV6	1068-1078		PVM1 REOVD	324-347	PVMAT 6SPVB	314-338		
PVGL2 IBV8	1068-1077		PVM1 REOVL	464-477	PVME1 CVBM	137-181		
PVGL2 IBVD2	1068-1078		PVMAT MUMP8	227-250	PVME1 CVHOC	137-181		
PVGL2 IBVK	1068-1077		PVMSA HPBDB	268-292	PVME1 CVTKE	137-181		
PVGL2 IBVM	1068-1077		PVMSA HPBDC	268-291	PVME1 IBV8	74-88		
PVGLB HSMV8J	117-138		PVMSA HPBDU	231-254	PVME1 IBV8	74-88		
PVGLB HSMV82	746-767		PVMSA HPBDW	268-292	PVME1 IBV82	74-88		
PVGLC HSMVB	389-421		PVMSA HPBHE	236-259	PVME1 IBVK	74-88		
PVGLC HSMVMQ	389-420				PVMSA HPBGB	271-285		
PVGLC HSMVM	389-421				PVMSA WHV1	269-283		
PVGLF BR8VA	285-287	482-504			PVMSA WHV59	274-288		
PVGLF BR8VC	484-508				PVMSA WHV7	274-288		
PVGLF BR8VR	484-508				PVMSA WHV8	274-288		
PVGLF HR8V1	484-508				PVMSA WHV81	274-288		
PVGLF HR8VA	484-508				PVMSA WHVW8	126-149		
PVGLF HR8VL	484-508							
PVGLF HR8VR	484-508							
PVGLF TRTV	452-474							
PVGL8 HNV	77-99							
PVGL8 VHSVO	406-428							

[illegible]

## TABLE IX

### Search Results Summary for P12CTLZIP Motif

**-64-**









[illegible]

[illegible]



[illegible]

**-72-**

**-73-**

[illegible]



[illegible]

[illegible]

## TABLE X

### Search Results Summary for P23CTLZIP Motif

**-78-**

PENV HV1W1	750-763				
PENV HV1W2	721-784				
PENV HV1Z2	264-286	727-760			
PENV HV1Z3	260-281				
PENV HV1Z8	265-288	728-762			
PENV HV1Z8	265-288				
PENV HV2BE	761-811				
PENV HV2D1	772-802				
PENV HV2G1	772-802				
PENV HV2NZ	777-814				
PENV HV29B	743-776				
PENV JSRV	288-332	484-518			
PENV MMTV8	438-472				
PENV MMTV9	438-472				
PENV R8VP	633-670				
PENV 8FV1	44-76	492-530			
PENV 8FV3L	48-82	550-588			
PENV 8IVC2	746-778				
PENV 8IVG8	247-277	383-388			
PENV 8IVM1	789-800				
PENV 8IVMK	768-789				
PENV 8IVML	611-645	764-798			
PENV 8IV84	468-488				
PENV 8IV8P	482-490	810-840			
PHEMA CDVO	200-234				
PHEMA IABUD	23-55				
PHEMA IACKA	23-55				
PHEMA IACKV	517-547				
PHEMA IADA1	23-55				
PHEMA IADCZ	23-55				
PHEMA IADH8	293-323				
PHEMA IADNZ	23-55				
PHEMA IAFPR	16-61				
PHEMA IAGRE	23-55				
PHEMA IAMAA	22-64				
PHEMA IAMAB	27-59				
PHEMA IARUD	23-55				
PHEMA IASE2	23-55				
PHEMA IASTA	617-647				
PHEMA MUMPM	18-52	101-132			
PHEMA MUMPR	18-52	101-132			
PHEMA MUMPS	18-52	101-132			
PHEMA NDVA	80-88				
PHEMA NDVB	80-88				
PHEMA NDVD	80-88				
PHEMA NDVH	80-88				
PHEMA NDVI	80-88				

PHEMA NDVM	60-88						
PHEMA NDVO	60-88						
PHEMA NDVTG	60-88						
PHEMA NDVU	60-88						
PHEMA PI1HW	29-80	198-233					
PHEMA PI2H	13-48	334-389					
PHEMA PI2HT	13-48	334-389					
PHEMA PI3B	194-231						
PHEMA PI3H4	194-231						
PHEMA PI3HA	194-231						
PHEMA PI3HT	194-231						
PHEMA PI3HU	194-231						
PHEMA PI3HV	194-231						
PHEMA PI3HW	194-231						
PHEMA PI3HX	194-231						
PHEMA PI4HA	245-280	338-376					
PHEMA RACV1	255-283						
PHEMA RINDL	282-313						
PHEMA SEND6	18-54	198-233					
PHEMA SENDF	18-54	198-233					
PHEMA SENDH	18-54	198-233					
PHEMA SENDJ	18-54	198-233					
PHEMA SENDZ	23-54	198-233					
PHEMA 8VA1	55-84	330-365					
PHEMA 8VB	7-35						
PHEMA 8V6CM	7-41						
PHEMA 8V6CP	7-41						
PHEMA 8VELN	7-35						
PHEMA VACCC	258-284						
PHEMA VACCI	258-284						
PHEMA VACCT	258-284						
PHEMA VACCV	258-284						
PVENV BEV	18-51	87-117					
PVENV DHV1	287-335						
PVENV MCV1	203-238						
PVENV MCV2	203-238						
PVENV VACCC	208-241						
PVENV VACCI	208-241						
PVENV VACCP	208-241						
PVENV VACCV	208-241						
PVF03 VACCC	2-40	61-93					
PVF03 VACCV	2-40	61-93					
PVF01 FOWPV	287-330						
PVF04 FOWPV	237-287						
PVF07 CAPVK	69-118						
PVF08 VACCC	28-61						
PVF08 VACCV	28-61						

PVG01 HSV11	317-340				
PVG02 HSV8B	163-186				
PVG02 VACCV	82-120				
PVG02 VARV	92-120				
PVG03 HSV11	108-138				
PVG06 HSV11	64-83				
PVG06 VACCC	88-138				
PVG06 VARV	88-138				
PVG07 VACCC	113-145				
PVG07 VARV	113-145				
PVG08 VACCC	303-338				
PVG08 VACCV	288-301				
PVG08 VARV	303-338				
PVG11 HSV11	150-183				
PVG12 HSV11	208-243				
PVG12 HSV8A	68-106				
PVG1 8PV1R	254-282	303-337	414-462		
PVG22 HSV11	300-337	647-678			
PVG23 HSV11	70-108				
PVG26 HSV11	84-125				
PVG27 HSV8A	38-74				
PVG28 HSV11	481-521				
PVG28 HSV8A	7-40				
PVG2R AMEPV	180-217				
PVG2 8PV4	208-244				
PVG35 HSV11	18-48	180-228			
PVG38 HSV8A	151-185				
PVG39 HSV11	543-577	848-882			
PVG40 HSV8A	187-216				
PVG41 HSV11	11-46	202-233			
PVG42 HSV11	81-125				
PVG43 HSV11	108-140	167-185			
PVG48 HSV11	888-926				
PVG48 HSV8A	328-357				
PVG50 HSV8A	113-141				
PVG81 HSV11	28-64	84-120			
PVG82 HSV11	88-134				
PVG85 HSV11	100-129				
PVG86 HSV11	631-667	1081-1128			
PVG88 HSV11	342-376	480-508			
PVG89 HSV8A	28-80	186-233			
PVG89 HSV11	82-118				
PVG81 HSV11	78-108				
PVG84 HSV11	66-88	363-401	420-452		
PVG85 HSV11	801-838	1280-1328			
PVG87 HSV11	150-188	1160-1185			
PVG8 8PV1R	60-88				

PVQ71 H8V8A	128-168				
PVQ72 H8V1	446-478	720-781	1168-1189	1262-1286	
PVQ76 H8V1	283-291	387-422			
PVQ78 H8V1	187-221				
PVQ7 SPV1R	18-48				
PVQF1 IBVB	1718-1747	1856-1881	2108-2148	3801-3833	
PVQH3 HCMVA	80-116	167-188			
PVQL2 CVBF	1259-1284				
PVQL2 CVBL0	681-681	1259-1284			
PVQL2 CVBLV		1259-1284			
PVQL2 CVBM		1259-1284			
PVQL2 CVBQ		1259-1284			
PVQL2 CVBV		1259-1284			
PVQL2 CVH22	1063-1088				
PVQL2 CVH4	1267-1304				
PVQL2 CVMA5	1216-1262				
PVQL2 CVMAH	1128-1183				
PVQL2 CVPF8	632-666	738-764	1328-1363		
PVQL2 CVPPU	630-663	734-762	1328-1361		
PVQL2 CVPR8	612-640	1104-1138			
PVQL2 CVPRM	408-441	1104-1139			
PVQL2 PIPV	835-868	738-767	1331-1368		
PVQL2 IBVB	153-188				
PVQL8 HCMVA	116-147	708-743			
PVQL8 HCMVT	116-147	707-744			
PVQL8 H8VB1	72-110				
PVQL8 H8VB2	264-288	745-774			
PVQL8 H8VBC	263-287				
PVQL8 LTV8	442-472				
PVQL8 LTV8	482-482				
PVQL8 LTVT	462-482				
PVQL8 MCMV8	136-163	738-778			
PVQLC H8V11	487-500				
PVQLC H8V1K	487-500				
PVQLC H8V2	435-466				
PVQLC H8V23	436-466				
PVQLC H8V8C	476-607				
PVQLC VZVD	351-388	613-648			
PVQLC VZVB	351-388	613-648			
PVQLD H8VEA	340-370				
PVQLD H8VEB	41-70	380-420			
PVQLD H8VEK	41-70	380-420			
PVQLD H8VE4	86-126				
PVQLE H8VEB	63-100	380-420			
PVQLE H8VEL	63-100	382-422			
PVQLE PRVRI	332-369				



PVQLF BR8VA	205-301	482-511		
PVQLF BR8VC	484-513			
PVQLF BR8VR	484-513			
PVQLF CDVO	682-888			
PVQLF HRSV1	484-513			
PVQLF HRSVA	484-513			
PVQLF HRSVL	484-513			
PVQLF HRSVR	484-513			
PVQLF MEASE	224-258	451-484		
PVQLF MEABI	227-258	454-487		
PVQLF MEASY	224-258	451-484		
PVQLF MUMPM	448-474			
PVQLF MUMPR	448-474			
PVQLF MUMPS	5-38	445-474		
PVQLF NDVI	132-185			
PVQLF PHODV	531-555			
PVQLF PI1HC	458-484			
PVQLF PI3B	453-481			
PVQLF PI3H4	453-481			
PVQLF RINDK	220-252	447-480		
PVQLF RINDL	220-252	447-480		
PVQLF SEND5	450-488			
PVQLF SENDF	480-488			
PVQLF SENDH	480-488			
PVQLF SENDJ	480-488			
PVQLF SENDZ	480-488			
PVQLF SV6	448-474			
PVQLF TRTV	452-481			
PVQLG H8VEB	327-384			
PVQLG SYNV	524-553			
PVQLG V8VIQ	450-488			
PVQLG V8VJO	457-492			
PVQLG V8VO	450-488			
PVQLG V8VBJ	450-488			
PVQLH HCMVA	601-718			
PVQLH HCMVT	690-718			
PVQLH H8V6G	640-677			
PVQLH H8VE4	814-850			
PVQLH H8VEB	807-843			
PVQLI HCMVA	158-184			
PVQLM BUNGE	197-227	438-468	882-1020	1048-1084
PVQLM BUNL7	180-220			
PVQLM BUNSH	190-220	344-381		
PVQLM BUNYW	183-228	434-472	823-854	
PVQLM DUGBV	244-273	637-672	888-918	935-985
PVQLM HANTB	610-641	1081-1119		1403-1441
PVQLM HANTH	188-222	612-643	1082-1120	

PVGLM HANTL	188-222	812-843	1083-1121	
PVGLM HANTV	188-222	812-843	1083-1121	
PVGLM PHV	616-848	1088-1121		
PVGLM PTPV	848-882	1276-1308		
PVGLM PUUMH	620-853	1082-1128		
PVGLM PUUMS	620-853	1082-1128		
PVGLM RVFV	620-853	830-883		
PVGLM RVFVZ	620-853	830-883	1168-1186	
PVGLM SEOUR	608-841	1082-1120		
PVGLM SEOUR	610-841	1081-1119		
PVGLM UUK	431-488	988-986		
PVGLP BEV	1491-1528			
PVGLY JUNIN	12-45			
PVGLY LASSQ	237-268			
PVGLY LASSJ	238-268			
PVGLY PIARV	12-60			
PVGLY TACV	12-60			
PVGLY TACV6	12-60	88-124		
PVGLY TACV7	12-60	88-124		
PVGLY TACVT	12-60	88-124		
PVGLB CPMV	1527-1555			
PVGLN BPMV	137-187	280-327	837-868	
PVGLN CPMV	208-242	741-771		
PVGLN CPMV	50-88	478-515		
PVGLN RCMV	788-789			
PVGP2 EBV	78-111			
PVGP3 EBV	78-111			
PVM1 REOVD	280-318	324-381		
PVM1 REOVL	280-318			
PVM21 REOVD	188-188			
PVM22 REOVD	188-188			
PVM2 REOVL	188-188			
PVM2 REOVL	188-188			
PVM3 REOVD	333-384			
PVMAT 8V6	308-342			
PVMAT TRTV	122-180			
PVME1 CVBM	64-102			
PVME1 CVHOC	64-102			
PVME1 CVMA6	66-103			
PVME1 CVMAH	66-103			
PVME1 CVTKE	64-102			
PVME1 EBV	178-213			
PVMP CERV	93-126			
PVMP 6OCMV	68-98	273-303		
PVMSA HPBDB	201-238	288-302		
PVMSA HPBDC	184-227	288-301		
PVMSA HPBDU	157-180	231-264		

[illegible]

### 5.3. SYNTHESIS OF PEPTIDES

The peptides of the invention may be synthesized or prepared by techniques well known in the art. See, for example, Creighton, 1983, *Proteins: Structures and Molecular Principles*, W.H. Freeman and Co., NY, which is incorporated herein by reference in its entirety. Short peptides, for example, can be synthesized on a solid support or in solution. Longer peptides may be made using recombinant DNA techniques. Here, the nucleotide sequences encoding the peptides of the invention may be synthesized, and/or cloned, and expressed according to techniques well known to those of ordinary skill in the art. See, for example, Sambrook, et al., 1989, *Molecular Cloning, A Laboratory Manual*, Vols. 1-3, Cold Spring Harbor Press, NY.

The peptides of the invention may alternatively be synthesized such that one or more of the bonds which link the amino acid residues of the peptides are non-peptide bonds. These alternative non-peptide bonds may be formed by utilizing reactions well known to those in the art, and may include, but are not limited to imino, ester, hydrazide, semicarbazide, and azo bonds, to name but a few. In yet another embodiment of the invention, peptides comprising the sequences described above may be synthesized with additional chemical groups present at their amino and/or carboxy termini, such that, for example, the stability, bioavailability, and/or inhibitory activity of the peptides is enhanced. For example, hydrophobic groups such as carbobenzoxy, dansyl, or t-butyloxycarbonyl groups, may be added to the peptides' amino termini. Likewise, an acetyl group or a 9-fluorenylmethoxy-carbonyl group may be placed at the peptides' amino termini. (See "X" in Tables I to IV, above.) Additionally, the hydrophobic group, t-

butyloxycarbonyl, or an amido group may be added to the peptides' carboxy termini. (See "Z" in Tables I to IV, above.) Further, the peptides of the invention may be synthesized such that their steric configuration is altered. For example, the D-isomer  
5 of one or more of the amino acid residues of the peptide may be used, rather than the usual L-isomer. Still further, at least one of the amino acid residues of the peptides of the invention may be substituted by one of the well known non-naturally occurring amino  
10 acid residues. Alterations such as these may serve to increase the stability, bioavailability and/or inhibitory action of the peptides of the invention.

Any of the peptides described above may, additionally, have a non-peptide macromolecular  
15 carrier group covalently attached to their amino and/or carboxy termini. Such macromolecular carrier groups may include, for example, lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates. "X", in Tables I to IV, above, may therefore  
20 additionally represent any of the above macromolecular carrier groups covalently attached to the amino terminus of a peptide. Likewise, "Z", in Tables I to IV, may additionally represent any of the  
25 macromolecular carrier groups described above.

#### 5.4. ASSAYS FOR ANTIVIRAL ACTIVITY

The antiviral activity exhibited by the peptides of the invention may be measured, for example, by easily performed in vitro assays, such as those  
30 described below, which can test the peptides' ability to inhibit syncytia formation, or their ability to inhibit infection by cell-free virus. Using these assays, such parameters as the relative antiviral activity of the peptides, exhibit against a given  
35 strain of virus and/or the strain specific inhibitory

activity of the peptide can be determined. A cell fusion assay may be utilized to test the peptides' ability to inhibit HIV-induced syncytia formation in vitro. Such an assay may comprise culturing uninfected CD-4<sup>+</sup> cells (such as Molt or CEM cells, for example) in the presence of chronically HIV-infected cells and a peptide to be assayed. For each peptide, a range of peptide concentrations may be tested. This range should include a control culture wherein no peptide has been added. Standard conditions for culturing, well known to those of ordinary skill in the art, are used. After incubation for an appropriate period (24 hours at 37°C, for example) the culture is examined microscopically for the presence of multinucleated giant cells, which are indicative of cell fusion and syncytia formation.

A reverse transcriptase (RT) assay may be utilized to test the peptides' ability to inhibit infection of CD-4<sup>+</sup> cells by cell-free HIV. Such an assay may comprise culturing an appropriate concentration (i.e., TCID<sub>50</sub>) of virus and CD-4<sup>+</sup> cells in the presence of the peptide to be tested. Culture conditions well known to those in the art are used. As above, a range of peptide concentrations may be used, in addition to a control culture wherein no peptide has been added. After incubation for an appropriate period (e.g., 7 days) of culturing, a cell-free supernatant is prepared, using standard procedures, and tested for the presence of RT activity as a measure of successful infection. The RT activity may be tested using standard techniques such as those described by, for example, Goff et al. (Goff, S. et al., 1981, J. Virol. 38:239-248) and/or Willey et al. (Willey, R. et al., 1988, J. Virol. 62:139-147). These references are incorporated herein by reference in their entirety.

Standard methods which are well-known to those of skill in the art may be utilized for assaying non-retroviral activity. See, for example, Pringle et al. (Pringle, C.R. et al., 1985, J. Medical Virology 17:377-386) for a discussion of respiratory syncytial virus and parainfluenza virus activity assay techniques. Further, see, for example, "Zinsser Microbiology", 1988, Joklik, W.K. et al., eds., Appleton & Lange, Norwalk, CT, 19th ed., for a general review of such techniques. These references are incorporated by reference herein in its entirety.

#### 5.5. USES OF THE PEPTIDES OF THE INVENTION

The DP-178 (SEQ ID:1) peptides of the invention, and DP-178 fragments, analogs, and homologs, exhibit potent antiviral activity. The DP-107-like and DP-178-like peptides of the invention preferably exhibit antiviral activity. As such, the peptides may be used as inhibitors of human and non-human viral and retroviral, especially HIV, transmission to uninfected cells.

The human retroviruses whose transmission may be inhibited by the peptides of the invention include, but are not limited to all strains of HIV-1 and HIV-2 and the human T-lymphocyte viruses (HTLV-I and II). The non-human retroviruses whose transmission may be inhibited by the peptides of the invention include, but are not limited to bovine leukosis virus, feline sarcoma and leukemia viruses, simian immunodeficiency, sarcoma and leukemia viruses, and sheep progress pneumonia viruses.

Non retroviral viruses whose transmission may be inhibited by the peptides of the invention include, but are not limited to human respiratory syncytial virus, canine distemper virus, newcastle disease virus, human parainfluenza virus, and influenza

viruses. Further, any virus or retrovirus containing peptides listed in Tables V through X above, may be inhibited by the peptides of the invention.

As discussed more fully, below, in Section 5.5.1 and in the Example presented, below, in Section 8, DP-107 and DP-178, and DP-107-like and DP-178-like peptides form non-covalent protein-protein interactions which are required for normal activity of the virus. Thus, the peptides of the invention may also be utilized as components in assays for the identification of compounds that interfere with such protein-protein interactions and may, therefore, act as antiviral agents. These assays are discussed, below, in Section 5.5.1.

5.5.1. ANTIVIRAL COMPOUND SCREENING ASSAYS FOR COMPOUNDS THAT INTERACT WITH THE PKD1 GENE PRODUCT

As demonstrated in the Example presented in Section 8, below, DP-107 and DP-178 portions of the TM protein gp41 form non-covalent protein-protein interactions. As also demonstrated, the maintenance of such interactions is necessary for normal viral infectivity. Thus, compounds which bind DP-107, bind DP-178, and/or act to disrupt normal DP-107/DP-178 protein-protein interactions may act as potent antiviral agents. Described below are assays for the identification of such compounds. Note that, while, for ease and clarity of discussion, DP-107 and DP-178 peptides will be used as components of the assays described, but it is to be understood that any of the DP-107-like or DP-178-like peptides described, above, in Sections 5.1 and 5.2 may also be utilized as part of these screens for antiviral compounds.

Compounds which may be tested for an ability to bind DP-107, DP-178, and/or disrupt DP-107/DP-178 interactions, and which therefore, potentially



represent antiviral compounds, include, but are not limited to, peptides made of D- and/or L-configuration amino acids (in, for example, the form of random peptide libraries; see Lam, K.S. *et al.*, 1991, *Nature* 354:82-84), phosphopeptides (in, for example, the form of random or partially degenerate, directed phosphopeptide libraries; see, for example, Songyang, Z. *et al.*, 1993, *Cell* 72:767-778), antibodies, and small organic or inorganic molecules. Synthetic compounds, natural products, and other sources of potentially effective materials may be screened in a variety of ways, as described in this Section. The compounds, antibodies, or other molecules identified may be tested for an ability to inhibit viral activity, utilizing, for example, viral assays such as those described, above, in Section 5.4.

Among the peptides which may be tested are soluble peptides comprising DP-107 and/or DP-178 domains, and peptides comprising DP-107 and/or DP-178 domains having one or more mutations within one or both of the domains, such as the M41-P peptide described, below, in the Example presented in Section 8, which contains a isoleucine to proline mutation within the DP-178 sequence.

In one embodiment of such screening methods is a method for identifying a compound to be tested for antiviral ability comprising:

- (a) exposing at least one compound to a peptide comprising a DP-107 peptide for a time sufficient to allow binding of the compound to the DP-107 peptide;
- (b) removing non-bound compounds; and
- (c) determining the presence of the compound bound to the DP-107 peptide, thereby identifying an agent to be tested for antiviral ability.

In a second embodiment of such screening methods is a method for identifying a compound to be tested for antiviral ability comprising:

- (a) exposing at least one compound to a peptide comprising a DP-178 peptide for a time  
5 sufficient to allow binding of the compound to the DP-178 peptide;
- (b) removing non-bound compounds; and
- (c) determining the presence of the  
10 compound bound to the DP-178 peptide,  
thereby identifying an agent to be tested for antiviral ability.

One method utilizing these types of approaches that may be pursued in the isolation of such DP-107-binding or DP-178-binding compounds is an assay which  
15 would include the attachment of either the DP-107 or the DP-178 peptide to a solid matrix, such as, for example, agarose or plastic beads, microtiter plate wells, petri dishes, or membranes composed of, for example, nylon or nitrocellulose. In such an assay  
20 system, either the DP-107 or DP-178 protein may be anchored onto a solid surface, and the compound, or test substance, which is not anchored, is labeled, either directly or indirectly. In practice, microtiter plates are conveniently utilized. The  
25 anchored component may be immobilized by non-covalent or covalent attachments. Non-covalent attachment may be accomplished simply by coating the solid surface with a solution of the protein and drying. Alternatively, an immobilized antibody, preferably a  
30 monoclonal antibody, specific for the protein may be used to anchor the protein to the solid surface. The surfaces may be prepared in advance and stored.

In order to conduct the assay, the labeled  
35 compound is added to the coated surface containing the anchored DP-107 or DP-178 peptide. After the reaction

is complete, unreacted components are removed (e.g., by washing) under conditions such that any complexes formed will remain immobilized on the solid surface. The detection of complexes anchored on the solid surface can be accomplished in a number of ways.

5 Where the compound is pre-labeled, the detection of label immobilized on the surface indicates that complexes were formed. Where the labeled component is not pre-labeled, an indirect label can be used to detect complexes anchored on the surface; e.g., using  
10 a labeled antibody specific for the compound (the antibody, in turn, may be directly labeled or indirectly labeled with a labeled anti-Ig antibody).

Alternatively, such an assay can be conducted in a liquid phase, the reaction products separated from  
15 unreacted components, and complexes detected; e.g., using an immobilized antibody specific for DP-107 or DP-178, whichever is appropriate for the given assay, or an antibody specific for the compound, i.e., the test substance, in order to anchor any complexes  
20 formed in solution, and a labeled antibody specific for the other member of the complex to detect anchored complexes.

By utilizing procedures such as this, large numbers of types of molecules may be simultaneously  
25 screened for DP-107 or DP-178-binding capability, and thus potential antiviral activity.

Further, compounds may be screened for an ability to inhibit the formation of or, alternatively, disrupt DP-107/DP-178 complexes. Such compounds may then be  
30 tested for antiviral capability. For ease of description, DP-107 and DP-178 will be referred to as "binding partners." Compounds that disrupt such interactions may exhibit antiviral activity. Such compounds may include, but are not limited to

35

molecules such as antibodies, peptides, and the like described above.

The basic principle of the assay systems used to identify compounds that interfere with the interaction between the DP-107 and DP-178 peptides involves  
5 preparing a reaction mixture containing peptides under conditions and for a time sufficient to allow the two peptides to interact and bind, thus forming a complex. In order to test a compound for disruptive activity, the reaction is conducted in the presence and absence  
10 of the test compound, i.e., the test compound may be initially included in the reaction mixture, or added at a time subsequent to the addition of one of the binding partners; controls are incubated without the test compound or with a placebo. The formation of any  
15 complexes between the binding partners is then detected. The formation of a complex in the control reaction, but not in the reaction mixture containing the test compound indicates that the compound interferes with the interaction of the DP-107 and  
20 DP-178 peptides.

The assay for compounds that interfere with the interaction of the binding partners can be conducted in a heterogeneous or homogeneous format. Heterogeneous assays involve anchoring one of the  
25 binding partners onto a solid phase and detecting complexes anchored on the solid phase at the end of the reaction. In homogeneous assays, the entire reaction is carried out in a liquid phase. In either approach, the order of addition of reactants can be  
30 varied to obtain different information about the compounds being tested. For example, test compounds that interfere with the interaction between the binding partners, e.g., by competition, can be identified by conducting the reaction in the presence  
35 of the test substance; i.e., by adding the test

substance to the reaction mixture prior to r  
simultaneously with the binding partners. On the  
ther hand, test compounds that disrupt preformed  
complexes, e.g. compounds with higher binding  
constants that displace one of the binding partners  
5 from the complex, can be tested by adding the test  
compound to the reaction mixture after complexes have  
been formed. The various formats are described  
briefly below.

10 In a heterogeneous assay system, one binding  
partner, e.g., either the DP-107 or DP-178 peptide, is  
anchored onto a solid surface, and its binding  
partner, which is not anchored, is labeled, either  
directly or indirectly. In practice, microtiter  
15 plates are conveniently utilized. The anchored  
species may be immobilized by non-covalent or covalent  
attachments. Non-covalent attachment may be  
accomplished simply by coating the solid surface with  
a solution of the protein and drying. Alternatively,  
an immobilized antibody specific for the protein may  
20 be used to anchor the protein to the solid surface.  
The surfaces may be prepared in advance and stored.

In order to conduct the assay, the binding  
partner of the immobilized species is added to the  
coated surface with or without the test compound.  
25 After the reaction is complete, unreacted components  
are removed (e.g., by washing) and any complexes  
formed will remain immobilized on the solid surface.  
The detection of complexes anchored on the solid  
surface can be accomplished in a number of ways.  
30 Where the binding partner was pre-labeled, the  
detection of label immobilized on the surface  
indicates that complexes were formed. Where the  
binding partner is not pre-labeled, an indirect label  
can be used to detect complexes anchored on the  
35 surface; e.g., using a labeled antibody specific for

the binding partner (the antibody, in turn, may be directly labeled or indirectly labeled with a labeled anti-Ig antibody). Depending upon the order of addition of reaction components, test compounds which inhibit complex formation or which disrupt preformed  
5 complexes can be detected.

Alternatively, the reaction can be conducted in a liquid phase in the presence or absence of the test compound, the reaction products separated from unreacted components, and complexes detected; e.g.,  
10 using an immobilized antibody specific for one binding partner to anchor any complexes formed in solution, and a labeled antibody specific for the other binding partner to detect anchored complexes. Again, depending upon the order of addition of reactants to  
15 the liquid phase, test compounds which inhibit complex or which disrupt preformed complexes can be identified.

In an alternate embodiment of the invention, a homogeneous assay can be used. In this approach, a  
20 preformed complex of the DP-107 and DP-178 peptides is prepared in which one of the binding partners is labeled, but the signal generated by the label is quenched due to complex formation (see, e.g., U.S. Patent No. 4,109,496 by Rubenstein which utilizes this  
25 approach for immunoassays). The addition of a test substance that competes with and displaces one of the binding partners from the preformed complex will result in the generation of a signal above background. In this way, test substances which disrupt DP-107/  
30 DP-178 protein-protein interaction can be identified.

#### 5.5 PHARMACEUTICAL FORMULATIONS, DOSAGES AND MODES OF ADMINISTRATION

With respect to HIV, the peptides of the  
35 invention may be used as a therapeutic in the

treatment of AIDS. The peptides of the invention may be administered using techniques well known to those in the art. Preferably, agents are formulated and administered systemically. Techniques for formulation and administration may be found in "Remington's  
5 Pharmaceutical Sciences", 18th ed., 1990, Mack Publishing Co., Easton, PA. Suitable routes may include oral, rectal, transmucosal, or intestinal administration; parenteral delivery, including  
10 intramuscular, subcutaneous, intramedullary injections, as well as intrathecal, direct intraventricular, intravenous, intraperitoneal, intranasal, or intraocular injections, just to name a few. Most preferably, administration is intravenous. For injection, the agents of the invention may be  
15 formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hanks' solution, Ringer's solution, or physiological saline buffer. For such transmucosal administration, penetrants appropriate to the barrier to be permeated  
20 are used in the formulation. Such penetrants are generally known in the art.

In addition, the peptides may be used as a prophylactic measure in previously uninfected individuals after acute exposure to an HIV virus.  
25 Examples of such prophylactic use of the peptides may include, but are not limited to, prevention of virus transmission from mother to infant and other settings where the likelihood of HIV transmission exists, such as, for example, accidents in health care settings  
30 wherein workers are exposed to HIV-containing blood products. The peptides of the invention in such cases may serve the role of a prophylactic vaccine, wherein the host raises antibodies against the peptides of the invention, which then serve to neutralize HIV viruses  
35 by, for example, inhibiting further HIV infection.

Administration of the peptides of the invention as a prophylactic vaccine, therefore, would comprise administering to a host a concentration of peptides effective in raising an immune response which is sufficient to neutralize HIV, by, for example, inhibiting HIV ability to infect cells. The exact concentration will depend upon the specific peptide to be administered, but may be determined by using standard techniques for assaying the development of an immune response which are well known to those of ordinary skill in the art. The peptides to be used as vaccines are usually administered intramuscularly.

The peptides may be formulated with a suitable adjuvant in order to enhance the immunological response. Such adjuvants may include, but are not limited to mineral gels such as aluminum hydroxide; surface active substances such as lysolecithin, pluronic polyols, polyanions; other peptides; oil emulsions; and potentially useful human adjuvants such as BCG and Corynebacterium parvum. Many methods may be used to introduce the vaccine formulations described here. These methods include but are not limited to oral, intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, and intranasal routes.

Alternatively, an effective concentration of polyclonal or monoclonal antibodies raised against the peptides of the invention may be administered to a host so that no uninfected cells become infected by HIV. The exact concentration of such antibodies will vary according to each specific antibody preparation, but may be determined using standard techniques well known to those of ordinary skill in the art. Administration of the antibodies may be accomplished using a variety of techniques, including, but not limited to those described in this section.



Effective dosages of the peptides of the invention to be administered may be determined through procedures well known to those in the art which address such parameters as biological half-life, bioavailability, and toxicity. Given the data  
5 presented below in Section 6, DP-178, for example, may prove efficacious in vivo at doses required achieve circulating levels of 10ng per ml of peptide.

A therapeutically effective dose refers to that amount of the compound sufficient to result in  
10 amelioration of symptoms or a prolongation of survival in a patient. Toxicity and therapeutic efficacy of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., for determining the LD50  
15 (the dose lethal to 50% of the population) and the ED50 (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD50/ED50. Compounds  
20 which exhibit large therapeutic indices are preferred. The data obtained from these cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of  
25 circulating concentrations that include the ED50 with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any compound used in the method of the invention, the  
30 therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC50 (i.e., the concentration of the test compound which  
35 achieves a half-maximal disruption of the PTK/adaptor

prot in complex, or a half-maximal inhibition of the cellular level and/or activity of a complex component) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for  
5 example, by high performance liquid chromatography (HPLC).

The exact formulation, route of administration and dosage can be chosen by the individual physician in view of the patient's condition. (See e.g. Fingl  
10 et al., 1975, in "The Pharmacological Basis of Therapeutics", Ch. 1 p1).

It should be noted that the attending physician would know how to and when to terminate, interrupt, or adjust administration due to toxicity, or to organ  
15 dysfunctions. Conversely, the attending physician would also know to adjust treatment to higher levels if the clinical response were not adequate (precluding toxicity). The magnitude of an administered dose in the management of the oncogenic disorder of interest  
20 will vary with the severity of the condition to be treated and to the route of administration. The dose and perhaps dose frequency, will also vary according to the age, body weight, and response of the individual patient. A program comparable to that  
25 discussed above may be used in veterinary medicine.

As demonstrated in the Example presented below in Section 6, the antiviral activity of the peptides of the invention may show a pronounced type and subtype specificity, i.e., specific peptides may be effective  
30 in inhibiting the activity of only specific viruses. This feature of the invention presents many advantages. One such advantage, for example, lies in the field of diagnostics, wherein one can use the antiviral specificity of the peptide of the invention  
35 to ascertain the identity of a viral isolate. With

respect to HIV, one may easily determine whether a viral isolate consists of an HIV-1 or HIV-2 strain. For example, uninfected CD-4<sup>+</sup> cells may be co-infected with an isolate which has been identified as containing HIV the DP-178 (SEQ ID:1) peptide, after  
5 which the retroviral activity of cell supernatants may be assayed, using, for example, the techniques described above in Section 5.2. Those isolates whose retroviral activity is completely or nearly completely inhibited contain HIV-1. Those isolates whose viral  
10 activity is unchanged or only reduced by a small amount, may be considered to not contain HIV-1. Such an isolate may then be treated with one or more of the other DP-178 peptides of the invention, and subsequently be tested for its viral activity in order  
15 to determine the identity of the viral isolate.

Use of pharmaceutically acceptable carriers to formulate the compounds herein disclosed for the practice of the invention into dosages suitable for systemic administration is within the scope of the  
20 invention. With proper choice of carrier and suitable manufacturing practice, the compositions of the present invention, in particular, those formulated as solutions, may be administered parenterally, such as by intravenous injection. The compounds can be  
25 formulated readily using pharmaceutically acceptable carriers well known in the art into dosages suitable for oral administration. Such carriers enable the compounds of the invention to be formulated as tablets, pills, capsules, liquids, gels, syrups,  
30 slurries, suspensions and the like, for oral ingestion by a patient to be treated.

Pharmaceutical compositions suitable for use in the present invention include compositions wherein the active ingredients are contained in an effective  
35 amount to achieve its intended purpose. Determination

of the effective amounts is well within the capability of those skilled in the art, especially in light of the detailed disclosure provided herein.

In addition to the active ingredients, these pharmaceutical compositions may contain suitable  
5 pharmaceutically acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. The preparations formulated  
10 for oral administration may be in the form of tablets, dragees, capsules, or solutions.

The pharmaceutical compositions of the present invention may be manufactured in a manner that is itself known, e.g., by means of conventional mixing,  
15 dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping or lyophilizing processes.

Pharmaceutical formulations for parenteral administration include aqueous solutions of the active  
20 compounds in water-soluble form. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes.  
25 Aqueous injection suspensions may contain substances which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Optionally, the suspension may also contain suitable stabilizers or agents which increase the  
30 solubility of the compounds to allow for the preparation of highly concentrated solutions.

Pharmaceutical preparations for oral use can be obtained by combining the active compounds with solid  
35 excipient, optionally grinding a resulting mixture, and processing the mixture of granules, after adding

suitable auxiliaries, if desired, to obtain tablets or dragee cores. Suitable excipients are, in particular, fillers such as sugars, including lactose, sucrose, mannitol, or sorbitol; cellulose preparations such as, for example, maize starch, wheat starch, rice starch, potato starch, gelatin, gum tragacanth, methyl cellulose, hydroxypropylmethyl-cellulose, sodium carboxymethylcellulose, and/or polyvinylpyrrolidone (PVP). If desired, disintegrating agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, or alginic acid or a salt thereof such as sodium alginate.

Dragee cores are provided with suitable coatings. For this purpose, concentrated sugar solutions may be used, which may optionally contain gum arabic, talc, polyvinyl pyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for identification or to characterize different combinations of active compound doses.

Pharmaceutical preparations which can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a plasticizer, such as glycerol or sorbitol. The push-fit capsules can contain the active ingredients in admixture with filler such as lactose, binders such as starches, and/or lubricants such as talc or magnesium stearate and, optionally, stabilizers. In soft capsules, the active compounds may be dissolved or suspended in suitable liquids, such as fatty oils, liquid paraffin, or liquid polyethylene glycols. In addition, stabilizers may be added.

35

6. EXAMPLE: DP-178 (SEQ ID:1) IS A POTENT  
INHIBITOR OF HIV-1 INFECTION

In this example, DP-178 (SEQ ID:1) is shown to be a potent inhibitor of HIV-1 mediated CD-4<sup>+</sup> cell-cell fusion and infection by cell free virus. In the fusion assay, this peptide completely blocks virus induced syncytia formation at concentrations of from 1-10 ng/ml. In the infectivity assay the inhibitory concentration is somewhat higher, blocking infection at 90ng/ml. It is further shown that DP-178 (SEQ ID:1) shows that the antiviral activity of DP-178 (SEQ ID:1) is highly specific for HIV-1. Additionally, a synthetic peptide, DP-185 (SEQ ID:3), representing a HIV-1-derived DP-178 homolog is also found to block HIV-1-mediated syncytia formation.

15

6.1. MATERIALS AND METHODS

6.1.1. PEPTIDE SYNTHESIS

Peptides were synthesized using Fast Moc chemistry on an Applied Biosystems Model 431A peptide synthesizer. Amidated peptides were prepared using Rink resin (Advanced Chemtech) while peptides containing free carboxy termini were synthesized on Wang (p-alkoxy-benzyl-alcohol) resin (Bachem). First residues were double coupled to the appropriate resin and subsequent residues were single coupled. Each coupling step was followed by acetic anhydride capping. Peptides were cleaved from the resin by treatment with trifluoroacetic acid (TFA) (10ml), H<sub>2</sub>O (0.5ml), thioanisole (0.5ml), ethanedithiol (0.25ml), and crystalline phenol (0.75g). Purification was carried out by reverse phase HPLC. Approximately 50mg samples of crude peptide were chromatographed on a Waters Delta Pak C18 column (19mm x 30cm, 15μ spherical) with a linear gradient; H<sub>2</sub>O/acetonitrile

0.1% TFA. Lyophilized peptides were stored desiccated and peptide solutions were made in water at about 1mg/ml. Electrospray mass spectrometry yielded the following results: DP-178 (SEQ ID:1):4491.87 (calculated 4491.94); DP-180 (SEQ ID:2):4491.45 (calculated 4491.94); DP-185 (SEQ ID:3):not done (calculated 4546.97).

#### 6.1.2. VIRUS

The HIV-1<sub>LA1</sub> virus was obtained from R. Gallo (Popovic, M. et al., 1984, Science 224:497-508) and propagated in CEM cells cultured in RPMI 1640 containing 10% fetal calf serum. Supernatant from the infected CEM cells was passed through a 0.2µm filter and the infectious titer estimated in a microinfectivity assay using the AA5 cell line to support virus replication. For this purpose, 25µl of serial diluted virus was added to 75µl AA5 cells at a concentration of  $2 \times 10^5$ /ml in a 96-well microtitre plate. Each virus dilution was tested in triplicate. Cells were cultured for eight days by addition of fresh medium every other day. On day 8 post infection, supernatant samples were tested for virus replication as evidenced by reverse transcriptase activity released to the supernatant. The TCID<sub>50</sub> was calculated according to the Reed and Muench formula (Reed, L.J. et al., 1938, Am. J. Hyg. 27:493-497). The titer of the HIV-1<sub>LA1</sub> and HIV-1<sub>MAN</sub> stocks used for these studies, as measured on the AA5 cell line, was approximately  $1.4 \times 10^6$  and  $3.8 \times 10^4$  TCID<sub>50</sub>/ml, respectively.

#### 6.1.3. CELL FUSION ASSAY

Approximately  $7 \times 10^4$  Molt cells were incubated with  $1 \times 10^4$  CEM cells chronically infected with the HIV-1<sub>LA1</sub> virus in 96-well plates (one-half area cluster plates; Costar, Cambridge, MA) in a final volume of

100 $\mu$ l culture medium as previously described (Matthews, T.J. et al., 1987, Proc. Natl. Acad. Sci. USA 84: 5424-5428). Peptide inhibitors were added in a volume of 10 $\mu$ l and the cell mixtures were incubated for 24 hr. at 37°C. At that time, multinucleated  
5 giant cells were estimated by microscopic examination at a 40x magnification which allowed visualization of the entire well in a single field.

#### 6.1.4. CELL FREE VIRUS INFECTION ASSAY

10 Synthetic peptides were incubated at 37°C with either 247 TCID<sub>50</sub> (for experiment depicted in FIG. 2), or 62 TCID<sub>50</sub> (for experiment depicted in FIG.3) units of HIV-1<sub>LAI</sub> virus or 25 TCID<sub>50</sub> units of HIV-2<sub>NH2</sub> and CEM CD4<sup>+</sup> cells at peptide concentrations of 0, 0.04, 0.4,  
15 4.0, and 40 $\mu$ g/ml for 7 days. The resulting reverse transcriptase (RT) activity in counts per minute was determined using the assay described, below, in Section 6.1.5. See, Reed, L.J. et al., 1938, Am. J. Hyg. 27: 493-497 for an explanation of TCID<sub>50</sub>  
20 calculations.

#### 6.1.5. REVERSE TRANSCRIPTASE ASSAY

The micro-reverse transcriptase (RT) assay was adapted from Goff et al. (Goff, S. et al., 1981, J. Virol. 38:239-248) and Willey et al. (Willey, R. et al., 1988, J. Virol. 62:139-147). Supernatants from virus/cell cultures are adjusted to 1% Triton-X100. A  
25 10 $\mu$ l sample of supernatant was added to 50 $\mu$ l of RT cocktail in a 96-well U-bottom microtitre plate and the samples incubated at 37°C for 90 min. The RT  
30 cocktail contained 75mM KCl, 2mM dithiothreitol, 5mM MgCl<sub>2</sub>, 5 $\mu$ g/ml poly A (Pharmacia, cat. No. 27-4110-01), 0.25 units/ml oligo dT (Pharmacia, cat. No. 27-7858-01), 0.05% NP40, 50mM Tris-HCl, pH 7.8, 0.5 $\mu$ M non-  
35



radioactive dTTP, and 10 $\mu$ Ci/ml <sup>32</sup>P-dTTP (Amersham, cat. No. PB.10167).

After the incubation period, 40 $\mu$ l of reaction mixture was applied to a Schleicher and Schuell (S+S) NA45 membrane (or DE81 paper) saturated in 2 x SSC buffer (0.3M NaCl and 0.003M sodium citrate) held in a S+S Minifold over one sheet of GB003 (S+S) filter paper, with partial vacuum applied. Each well of the minifold was washed four times with 200 $\mu$ l 2xSSC, under full vacuum. The membrane was removed from the minifold and washed 2 more times in a pyrex dish with an excess of 2xSSC. Finally, the membrane was drained on absorbent paper, placed on Whatman #3 paper, covered with Saran wrap, and exposed to film overnight at -70°C.

## 6.2. RESULTS

### 6.2.1. PEPTIDE INHIBITION OF INFECTED CELL-INDUCED SYNCYTIA FORMATION

The initial screen for antiviral activity assayed peptides' ability to block syncytium formation induced by overnight co-cultivation of uninfected Molt4 cells with chronically HIV-1 infected CEM cells. The results of several such experiments are presented herein. In the first of these experiments, serial DP-178 (SEQ ID:1) peptide concentrations between 10 $\mu$ g/ml and 12.5ng/ml were tested for blockade of the cell fusion process. For these experiments, CEM cells chronically infected with either HIV-1<sub>LAI</sub>, HIV-1<sub>MN</sub>, HIV-1<sub>RF</sub>, or HIV-1<sub>SF2</sub> virus were cocultivated overnight with uninfected Molt 4 cells. The results (FIG. 4) show that DP-178 (SEQ ID:1) afforded complete protection against each of the HIV-1 isolates down to the lowest concentration of DP-178 (SEQ ID:1) used. For HIV<sub>LAI</sub> inhibition, the lowest concentration tested was

12.5ng/ml; for all other HIV-1 viruses, the lowest concentration of DP-178 (SEQ ID:1) used in this study was 100ng/ml. A second peptide, DP-180 (SEQ ID:2), containing the same amino acid residues as DP-178 (SEQ ID:1) but arranged in a random order exhibited no  
5 evidence of anti-fusogenic activity even at the high concentration of 40µg/ml (FIG. 4). These observations indicate that the inhibitory effect of DP-178 (SEQ ID:1) is primary sequence-specific and not related to non-specific peptide/protein interactions. The actual  
10 endpoint (i.e., the lowest effective inhibitory concentration) of DP-178 inhibitory action is within the range of 1-10 ng/ml.

The next series of experiments involved the preparation and testing of a DP-178 (SEQ ID:1) homolog  
15 for its ability to inhibit HIV-1-induced syncytia formation. As shown in FIG. 1, the sequence of DP-185 (SEQ ID:3) is slightly different from DP-178 (SEQ ID:1) in that its primary sequence is taken from the HIV-1<sub>SP2</sub> isolate and contains several amino acid  
20 differences relative to DP-178 (SEQ ID:1) near the N terminus. As shown in FIG. 4, DP-185 (SEQ ID:3), exhibits inhibitory activity even at 312.5ng/ml, the lowest concentration tested.

The next series of experiments involved a  
25 comparison of DP-178 (SEQ ID:1) HIV-1 and HIV-2 inhibitory activity. As shown in FIG. 5, DP-178 (SEQ ID:1) blocked HIV-1-mediated syncytia formation at peptide concentrations below 1ng/ml. DP-178 (SEQ ID:1) failed, however, to block HIV-2 mediated  
30 syncytia formation at concentrations as high as 10µg/ml. This striking 4 log selectivity of DP-178 (SEQ ID:1) as an inhibitor of HIV-1-mediated cell fusion demonstrates an unexpected HIV-1 specificity in the action of DP-178 (SEQ ID:1). DP-178 (SEQ ID:1)  
35 inhibition of HIV-1-mediated cell fusion, but the

peptide's inability to inhibit HIV-2 medicated cell fusion in the same cell type at the concentrations tested provides further evidence for the high degree of selectivity associated with the antiviral action of DP-178 (SEQ ID:1).

5

#### 6.2.2. PEPTIDE INHIBITION OF INFECTION BY CELL-FREE VIRUS

DP-178 (SEQ ID:1) was next tested for its ability to block CD-4<sup>+</sup> CEM cell infection by cell free HIV-1 virus. The results, shown in FIG. 2, are from an experiment in which DP-178 (SEQ ID:1) was assayed for its ability to block infection of CEM cells by an HIV-1<sub>LAI</sub> isolate. Included in the experiment were three control peptides, DP-116 (SEQ ID:9), DP-125 (SEQ ID:8), and DP-118 (SEQ ID:10). DP-116 (SEQ ID:9) represents a peptide previously shown to be inactive using this assay, and DP-125 (SEQ ID:8; Wild, C. *et al.*, 1992, Proc. Natl. Acad. Sci. USA **89**:10,537) and DP-118 (SEQ ID:10) are peptides which have previously been shown to be active in this assay. Each concentration (0, 0.04, 0.4, 4, and 40 µg/ml) of peptide was incubated with 247 TCID<sub>50</sub> units of HIV-1<sub>LAI</sub> virus and CEM cells. After 7 days of culture, cell-free supernatant was tested for the presence of RT activity as a measure of successful infection. The results, shown in FIG. 2, demonstrate that DP-178 (SEQ ID:1) inhibited the de novo infection process mediated by the HIV-1 viral isolate at concentrations as low as 90ng/ml (IC<sub>50</sub>=90ng/ml). In contrast, the two positive control peptides, DP-125 (SEQ ID:8) and DP-118 (SEQ ID:10), had over 60-fold higher IC<sub>50</sub> concentrations of approximately 5 µg/ml.

In a separate experiment, the HIV-1 and HIV-2 inhibitory action of DP-178 (SEQ ID:1) was tested with CEM cells and either HIV-1<sub>LAI</sub> or HIV-2<sub>NIH2</sub>. 62 TCID<sub>50</sub>

HIV-1<sub>LAI</sub> or 25 GCID<sub>50</sub> HIV-2<sub>NIH</sub> were used in these experiments, and were incubated for 7 days. As may be seen in FIG. 3, DP-178 (SEQ ID:1) inhibited HIV-1 infection with an IC<sub>50</sub> of about 31ng/ml. In contrast, DP-178 (SEQ ID:1) exhibited a much higher IC<sub>50</sub> for HIV-2<sub>NIH</sub>, thus making DP-178 (SEQ ID:1) two logs more potent as a HIV-1 inhibitor than a HIV-2 inhibitor. This finding is consistent with the results of the fusion inhibition assays described, above, in Section 6.2.1, and further supports a significant level of selectivity (i.e., for HIV-1 over HIV-2).

7. EXAMPLE: THE HIV-1 INHIBITOR, DP-178 (SEQ ID:1) IS NON-CYTOXIC

In this Example, the 36 amino acid synthetic peptide inhibitor DP-178 (SEQ ID:1) is shown to be non-cytotoxic to cells in culture, even at the highest peptide concentrations (40μg/ml) tested.

7.1. MATERIALS AND METHODS

Cell proliferation and toxicity assay:  
Approximately 3.8x10<sup>5</sup> CEM cells for each peptide concentration were incubated for 3 days at 37°C in T25 flasks. Peptides tested were DP-178 (SEQ ID:1) and DP-116 (SEQ ID:9), as described in FIG. 1. The concentrations of each peptide used were 0, 2.5, 10, and 40μg/ml. Cell counts were taken at incubation times of 0, 24, 48, and 72 hours.

7.2. RESULTS

Whether the potent HIV-1 inhibitor DP-178 (SEQ ID:1) exhibited any cytotoxic effects was assessed by assaying the peptide's effects on the proliferation and viability of cells in culture. CEM cells were incubated in the presence of varying concentrations of DP-178 (SEQ ID:1), and DP-116 (SEQ ID:9), a peptide

previously shown to be ineffective as a HIV inhibitor (Wild, C. et al., 1992, Proc. Natl. Acad. Sci. USA 89:10,537-10,541). Additionally, cells were incubated in the absence of either peptide.

5 The results of the cytotoxicity study demonstrate that DP-178 (SEQ ID:1) exhibits no cytotoxic effects on cells in culture. As can be seen, below, in Table XI, even the proliferation and viability characteristics of cells cultured for 3 days in the presence of the highest concentration of DP-178 (SEQ  
10 ID:1) tested (40 µg/ml) do not significantly differ from the DP-116 (SEQ ID:9) or the no-peptide controls. The cell proliferation data is also represented in graphic form in FIG. 6. As was demonstrated in the Working Example presented above in Section 6, DP-178  
15 (SEQ ID:1) completely inhibits HIV-1 mediated syncytia formation at peptide concentrations between 1 and 10 ng/ml, and completely inhibits cell-free viral infection at concentrations of at least 90 ng/ml. Thus, this study demonstrates that even at peptide  
20 concentrations greater than 3 log higher than the HIV inhibitory dose, DP-178 (SEQ ID:1) exhibits no cytotoxic effects.

25 TABLE XI

Peptide	Peptide Concentration µg/ml	% Viability at time (hours)			
		0	24	48	72
30 DP178 (SEQ ID:1)	40	98	97	95	97
	10	98	97	98	98
	2.5	98	93	96	96

35

5	DP116 (SEQ ID:9)	40	98	95	98	97
		10	98	95	93	98
		2.5	98	96	98	99
	No Peptide	0	98	97	99	98

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10        8. EXAMPLE: THE INTERACTION OF DP178 AND DP107  
 Soluble recombinant forms of gp41 used in the  
 example described below provide evidence that the  
 DP178 peptide associates with a distal site on gp41  
 whose interactive structure is influenced by the DP107  
 15 leucine zipper motif. A single mutation disrupting  
 the coiled-coil structure of the leucine zipper domain  
 transformed the soluble recombinant gp41 protein from  
 an inactive to an active inhibitor of HIV-1 fusion.  
 This transformation may result from liberation of the  
 20 potent DP178 domain from a molecular clasp with the  
 leucine zipper, DP107, determinant. The results also  
 indicate that the anti-HIV activity of various gp41  
 derivatives (peptides and recombinant proteins) may be  
 due to their ability to form complexes with viral gp41  
 25 and interfere with its fusogenic process.

#### 8.1. MATERIALS AND METHODS

##### 8.1.1. CONSTRUCTION OF FUSION PROTEINS AND GP41 MUTANTS

30        Construction of fusion proteins and mutants shown  
 in FIG. 7 was accomplished as follows: the DNA  
 sequence corresponding to the extracellular domain of  
 gp41 (540-686) was cloned into the Xmn I site of the  
 expression vector pMal-p2 (New England Biolab) to give  
 35 M41. The gp41 sequence was amplified from pgtat

(Malim et al., 1988, Nature 355: 181-183) by using polymerase chain reaction (PCR) with upstream primer 5'-ATGACGCTGACGGTACAGGCC-3' (primer A) and downstream primer 5'-TGAATAAGCTTAATACCACAGCCAATTTGTTAT-3' (primer B). M41-P was constructed by using the T7-Gen  
5 in vitro mutagenesis kit from United States Biochemicals (USB) following the supplier's instructions. The mutagenic primer (5'-GGAGCTGCTTGGGGCCCCAGAC-3') introduces an Ile to Pro mutation in M41 at position 578. M41Δ107 was made  
10 using a deletion mutagenic primer 5'-CCAAATCCCCAGGAGCTGCTCGAGCTGCACTATACCAGAC-3' (primer C) following the USB T7-Gen mutagenesis protocol. M41Δ178 was made by cloning the DNA fragment corresponding to gp41 amino acids 540-642 into the Xmn  
15 I site of pMal-p2. Primer A and 5'-ATAGCTTCTAGATTAAATTGTTAATTTCTCTGTCCC-3' (primer D) were used in the PCR with the template pgtat to generate the inserted DNA fragments. M41-P was used as the template with primer A and D in PCR to generate M41-  
20 PA178. All inserted sequences and mutated residues were checked by restriction enzyme analysis and confirmed by DNA sequencing.

#### 25 8.1.2. PURIFICATION AND CHARACTERIZATION OF FUSION PROTEINS

The fusion proteins were purified according to the protocol described in the manufacturer's brochure of protein fusion and purification systems from New England Biolabs (NEB). Fusion proteins (10 ng) were  
30 analyzed by electrophoresis on 8% SDS polyacrylamide gels. Western blotting analysis was performed as described by Sambrook et al, 1989, Molecular Cloning: A Laboratory Manual, 2d Ed, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, Ch. 18,  
35 pp. 64-75. An HIV-1 positive serum diluted 1000-fold,

or a human Fab derived from repertoire cloning was used to react with the fusion proteins. The second antibody was HRP-conjugated goat antihuman Fab. An ECL Western blotting detection system (Amersham) was used to detect the bound antibody. A detailed  
5 protocol for this detection system was provided by the manufacturer. Rainbow molecular weight marker (Amersham) were used to estimate the size of fusion proteins.

10 8.1.3. CELL FUSION ASSAYS FOR ANTI-HIV ACTIVITY

Cell fusion assays were performed as previously described (Matthews et al., 1987, Proc. Natl. Acad. Sci. USA 84: 5424-5481). CEM cells ( $7 \times 10^4$ ) were  
15 incubated with HIV-1<sub>ms</sub> chronically infected CEM cells ( $10^4$ ) in 96-well flat-bottomed half-area plates (Costar) in 100  $\mu$ l culture medium. Peptide and fusion proteins at various concentrations in 10  $\mu$ l culture medium were incubated with the cell mixtures at 37°C for 24 hours. Multinucleated syncytia were estimated  
20 with microscopic examination. Both M41 and M41-P did not show cytotoxicity at the concentrations tested and shown in FIG. 8.

Inhibition of HIV-1 induced cell-cell fusion activity was carried out in the presence of 10 nM  
25 DP178 and various concentrations of M41 $\Delta$ 178 or M41-PA178 as indicated in FIG. 9. There was no observable syncytia in the presence of 10 nM DP178. No peptide or fusion protein was added in the control samples.

30 8.1.4. ELISA ANALYSIS OF DP178 BINDING TO THE LEUCINE ZIPPER MOTIF OF GP41

The amino acid sequence of DP178 used is:  
YTSLIHSLEESQNQQEKNEQELLELDKWASLWNWF. For enzyme  
linked immunoassay (ELISA), M41 $\Delta$ 178 or M41-PA178 (5  
35  $\mu$ g/ml) in 0.1M NaHCO<sub>3</sub>, pH 8.6, were coated on 96 wells



Linbro ELISA plates (Flow Lab, Inc.) overnight. Each well was washed three times with distilled water then blocked with 3% bovine serum albumin (BSA) for 2 hours. After blocking, peptides with 0.5% BSA in TBST (40 mM Tris-HCl pH7.5, 150 mM NaCl, 0.05% Tween 20) were added to the ELISA plates and incubated at room temperature for 1 hour. After washing three times with TBST, Fab-d was added at a concentration of 10 ng/ml with 0.5% BSA in TBST. The plates were washed three times with TBST after incubation at room temperature for 1 hour. Horse radish peroxidase (HRP) conjugated goat antihuman Fab antiserum at a 2000 fold dilution in TBST with 0.5% BSA was added to each well and incubated at room temperature for 45 minutes. The plates were then washed four times with TBST. The peroxidase substrate o-phenylene diamine (2.5 mg/ml) and 0.15% H<sub>2</sub>O<sub>2</sub> were added to develop the color. The reaction was stopped with an equal volume of 4.5 N H<sub>2</sub>SO<sub>4</sub> after incubation at room temperature for 10 minutes. The optical density of the stopped reaction mixture was measured with a micro plate reader (Molecular Design) at 490 nm. Results are shown in FIG. 10.

## 8.2. RESULTS

### 8.2.1. THE EXPRESSION AND CHARACTERIZATION OF THE ECTODOMAIN OF GP41

As a step toward understanding the roles of the two helical regions in gp41 structure and function, the ectodomain of gp41 was expressed as a maltose binding fusion protein (M41) (Fig. 7). The fusogenic peptide sequence at the N-terminal of gp41 was omitted from this recombinant protein and its derivatives to improve solubility. The maltose binding protein facilitated purification of the fusion proteins under relatively mild, non-denaturing conditions. Because

the M41 soluble r combinant gp41 was not glycosylated, lacked several regions of the transmembrane protein (i.e., the fusion peptid , the membrane spanning, and the cytoplasmic domains), and was expressed in the absence of gp120, it was not expected to precisely  
5 reflect the structure of native gp41 on HIV-1 virions. Nevertheless, purified M41 folded in a manner that preserved certain discontinuous epitopes as evidenced by reactivity with human monoclonal antibodies, 98-6, 126-6, and 50-69, previously shown to bind  
10 conformational epitopes on native gp41 expressed in eukaryotic cells (Xu et al., 1991, J. Virol. 65: 4832-4838; Chen, 1994, J. Virol. 68:2002-2010). Thus, at least certain regions of native gp41 defined by these antibodies appear to be reproduced in the recombinant  
15 fusion protein M41. Furthermore, M41 reacted with a human recombinant Fab (Fab-d) that recognizes a conformational epitope on gp41 and binds HIV-1 virions as well as HIV-1 infected cells but not uninfected cells as analyzed by FACS. Deletion of either helix  
20 motif, i.e., DP107 or DP178, of the M41 fusion protein eliminated reactivity with Fab-d. These results indicate that both helical regions, separated by 60 amino acids in the primary sequence, are required to maintain the Fab-d epitope.  
25

#### 8.2.2. ANTI-HIV ACTIVITY OF THE RECOMBINANT ECTODOMAIN OF GP41

The wild type M41 fusion protein was tested for  
30 anti-HIV-1 activity. As explained, supra, synthetic peptides corresponding to the leucine zipper (DP107) and the C-terminal putative helix (DP178) show potent anti-HIV activity. Despite inclusion of both these regions, the recombinant M41 protein did not affect  
35

HIV-1 induced membrane fusion at concentrations as high as 50  $\mu$ M (Table XII, below).

**TABLE XII**  
**DISRUPTION OF THE LEUCINE ZIPPER OF**  
**GP41 FREES THE ANTI-HIV MOTIF**

	<u>DP107</u>	<u>DP178</u>	<u>M41</u>	<u>M41-P</u>	<u>M41-PA178</u>
Cell fusion (IC <sub>50</sub> )	1 $\mu$ M	1 nM	> 50 $\mu$ M	83 nM	> 50 $\mu$ M
Fab-D binding (K <sub>D</sub> )	-	-	3.5x10 <sup>-9</sup>	2.5x10 <sup>-8</sup>	-
HIV infectivity (IC <sub>50</sub> )	1 $\mu$ M	80 nM	> 16 $\mu$ M	66 nM	> 8 $\mu$ M

The affinity constants of Fab-d binding to the fusion proteins were determined using a protocol described by B. Friguet et al., 1985, J. Immunol. Method. 77:305-319.

- = No detectable binding of Fab-d to the fusion proteins.

*Antiviral Infectivity Assays.* 20  $\mu$ l of serially diluted virus stock was incubated for 60 minutes at ambient temperature with 20  $\mu$ l of the indicated concentration of purified recombinant fusion protein in RPMI 1640 containing 10% fetal bovine serum and antibiotics in a 96-well microtiter plate. 20  $\mu$ l of CEM4 cells at 6 x 10<sup>5</sup> cells/ml were added to each well, and cultures were incubated at 37°C in a humidified CO<sub>2</sub> incubator. Cells were cultured for 9 days by the addition of fresh medium every 2 to 30 days. On days 5, 7, and 9 postinfection, supernatant samples were assayed for reverse transcriptase (RT) activity, as described below, to monitor viral replication. The 50% tissue culture infectious dose (TCID<sub>50</sub>) was calculated for each condition according to the formula of Reed & Muench, 1937, Am. J. Hyg. 27:493-497. RT activity was determined by a modification of the published methods of Goff et al., 1981, J. Virol. 38:239-248 and Willey et al., 1988, J. Virol. 62:139-147 as described in Chen et al., 1993, AIDS Res. Human Retroviruses 9:1079-1086.

Surprisingly, a single amino acid substitution, proline in place of isoleucine in the middle of the leucine zipper motif, yielded a fusion protein (M41-P)

which did exhibit antiviral activity (Table XII and Fig. 8). As seen in Table XII, M41-P blocked syncytia formation by 90% at approximately 85 nM and neutralized HIV-1<sub>MB</sub> infection by 90% at approximately 70 nM concentrations. The anti-HIV-1 activity of M41-P appeared to be mediated by the C-terminal helical sequence since deletion of that region from M41-P yielded an inactive fusion protein, M41-PA178 (Table XII). That interpretation was reinforced by experiments demonstrating that a truncated fusion protein lacking the DP178 sequence, M41Δ178, abrogated the potent anti-fusion activity of the DP178 peptide in a concentration-dependent manner (FIG. 9). The same truncated fusion protein containing the proline mutation disrupting the leucine zipper, M41-PA178, was not active in similar competition experiments (FIG. 9). The results indicate that the DP178 peptide associates with a second site on gp41 whose interactive structure is dependent on a wild type leucine zipper sequence. A similar interaction may occur within the wild type fusion protein, M41, and act to form an intramolecular clasp which sequesters the DP178 region, making it unavailable for anti-viral activity.

A specific association between these two domains is also indicated by other human monoclonal Fab-d studies. For example, Fab-d failed to bind either the DP178 peptide or the fusion protein M41Δ178, but its epitope was reconstituted by simply mixing these two reagents together (FIG. 10). Again, the proline mutation in the leucine zipper domain of the fusion protein, M41-PA178, failed to reconstitute the epitope in similar mixing experiments.

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9. EXAMPLE: METHOD FOR COMPUTER-ASSISTED  
IDENTIFICATION OF DP-107-LIKE  
AND DP-178-LIKE SEQUENCES

A number of known coiled-coil sequences have been well described in the literature and contain heptad repeat positioning for each amino acid. Coiled-coil nomenclature labels each of seven amino acids of a heptad repeat A through G, with amino acids A and D tending to be hydrophobic positions. Amino acids E and G tend to be charged. These four positions (A, D, E, and G) form the amphipathic backbone structure of a monomeric alpha-helix. The backbones of two or more amphipathic helices interact with each other to form di-, tri-, tetrameric, etc., coiled-coil structures. In order to begin to design computer search motifs, a series of well characterized coiled coils were chosen including yeast transcription factor GCN4, Influenza Virus hemagglutinin loop 36, and human proto-oncogenes c-Myc, c-Fos, and c-Jun. For each peptide sequence, a strict homology for the A and D positions, and a list of the amino acids which could be excluded for the B, C, E, F, and G positions (because they are not observed in these positions) was determined. Motifs were tailored to the DP-107 and DP-178 sequences by deducing the most likely possibilities for heptad positioning of the amino acids of HIV-1 Bru DP-107, which is known to have coiled-coil structure, and HIV-1 Bru DP-178, which is still structurally undefined. The analysis of each of the sequences is contained in FIG. 12. For example, the motif for GCN4 was designed as follows:

1. The only amino acids (using standard single letter amino acid codes) found in the A or D positions of GCN4 were [LMNV].
2. All amino acids were found at B, C, E, F, and G positions except {CFGIMPTW}.

3. The PESEARCH motif would, therefore, be written as follows:

[LMNV]-{CFGIMPTW}(2)-[LMNV]-{CFGIMPTW}(3)-  
 [LMNV]-{CFGIMPTW}(2)-[LMNV]-{CFGIMPTW}(3)-  
 [LMNV]-{CFGIMPTW}(2)-[LMNV]-{CFGIMPTW}(3)-  
 5 [LMNV]-{CFGIMPTW}(2)-[LMNV]-{CFGIMPTW}(3)

Translating or reading the motif: "at the first A position either L, M, N, or V must occur; at positions B and C (the next two positions) accept everything  
 10 except C, F, G, I, M, P, T, or W; at the D position either L, M, N, or V must occur; at positions E, F, and G (the next 3 positions) accept everything except C, F, G, I, M, P, T, or W." This statement is  
 15 contained four times in a 28-mer motif and five times in a 35-mer motif. The basic motif key then would be: [LMNV]-{CFGIMPTW}. The motif keys for the remaining well described coiled-coil sequences are summarized in FIG. 12.

The motif design for DP-107 and DP-178 was  
 20 slightly different than the 28-mer model sequences described above due to the fact that heptad repeat positions are not defined and the peptides are both longer than 28 residues. FIG. 13 illustrates several possible sequence alignments for both DP-107 and DP-  
 25 178 and also includes motif designs based on 28<sup>-mer</sup>, 35<sup>-mer</sup>, and full-length peptides. Notice that only slight differences occur in the motifs as the peptides are lengthened. Generally, lengthening the base  
 30 peptide results in a less stringent motif. This is very useful in broadening the possibilities for identifying DP-107-or DP-178-like primary amino acid sequences referred to in this document as "hits".

In addition to making highly specific motifs for  
 35 each type peptide sequence to be searched, it is also possible to make "hybrid" motifs. These motifs are

made by "crossing" two or more very stringent motifs to make a new search algorithm which will find not only both "parent" motif sequences but also any peptide sequences which have similarities to one, the other, or both "parents". For example, in Table 3 the  
5 "parent" sequence of GCN4 is crossed with each of the possible "parent" motifs of DP-107. Now the hybrid motif must contain all of the amino acids found in the A and D positions of both parents, and exclude all of the amino acids not found in either parent at the  
10 other positions. The resulting hybrid from crossing GCN4 or [LMNV]{CFGIMPTW} and DP-107 (28-mer with the first L in the D position) or [ILQT]{CDFIMPST}, is [ILMNQTV]{CFIMPT}. Notice that now only two basic  
15 hybrid motifs exist which cover both framing possibilities, as well as all peptide lengths of the parent DP-107 molecule. FIG. 15 represents the hybridizations of GCN4 with DP-178. FIG. 16 represents the hybridizations of DP-107 and DP-178. It is important to keep in mind that the represented  
20 motifs, both parent and hybrid, are motif keys and not the depiction of the full-length motif needed to actually do the computer search.

Hybridizations can be performed on any combination of two or more motifs. Table 5  
25 summarizes several three-motif hybridizations including GCN4, DP-107 (both frames), and DP-178 (also both frames). Notice that the resulting motifs are now becoming much more similar to each other. In fact, the first and third hybrid motifs are actually  
30 subsets of the second and fourth hybrid motifs respectively. This means that the first and third hybrid motifs are slightly more stringent than the second and fourth. It should also be noted that with only minor changes in these four motifs, or by  
35 hybridizing them, a single motif could be obtained

which would find all of the sequences. However, it should be remembered that stringency is also reduced. Finally, the most broad-spectra and least-stringent hybrid motif is described in FIG. 18 which summarizes the hybridization of GCN4, DP-107 (both frames), DP-178 (both frames), c-Fos, c-Jun, c-Myc, and Flu loop 36.

A special set of motifs was designed based on the fact that DP-178 is located only approximately ten amino acids upstream of the transmembrane spanning region of gp41 and just C-terminal to a proline which separates DP-107 and DP-178. It has postulated that DP-178 may be an amphipathic helix when membrane associated, and that the proline might aid in the initiation of the helix formation. The same arrangement was observed in Respiratory Syncytial Virus; however, the DP-178-like region in this virus also had a leucine zipper just C-terminal to the proline. Therefore, designed N-terminal proline-leucine zipper motifs were designed to analyze whether any other viruses might contain this same pattern. The motifs are summarized in FIG. 19.

The PC/Gene protein database contains 5879 viral amino acid sequences (library file PVIRUSES; CD-ROM release 11.0). Of these, 1092 are viral envelope or glycoprotein sequences (library file PVIRUSE1). Tables V through X contain lists of protein sequence names and motif hit locations for all the motifs searched.

10. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION  
OF DP-107 AND DP-178-LIKE SEQUENCES  
IN HUMAN IMMUNODEFICIENCY VIRUS

FIG. 20 represents search results for HIV-1 BRU isolate gp41 (PC/Gene protein sequence PENV\_HV1BR). Notice that the hybrid motif which crosses DP-107 and



DP-178 (named 107x178x4; the same motif as found in FIG. 16 found three hits including amino acids 550-599, 636-688, and 796-823. These areas include DP-107 plus eight N-terminal and four C-terminal amino acids; DP-178 plus seven N-terminal and ten C-terminal amino acids; and an area inside the transmembrane region (cytoplasmic). FIG. 20 also contains the results obtained from searching with the motif named ALLMOTI5, for which the key is found in FIG. 17 ({CDGHP}{CFP}x5). This motif also found three hits including DP-107 (amino acids 510-599), DP-178 (615-717), and a cytoplasmic region (772-841). These hits overlap the hits found by the motif 107x178x4 with considerable additional sequences on both the amino and carboxy termini. This is not surprising in that 107x178x4 is a subset of the ALLMOTI5 hybrid motif. Importantly, even though the stringency of ALLMOTI5 is considerably less than 107x178x4, it still selectively identifies the DP-107 and DP-178 regions of gp41 shown to contain sequences for inhibitory peptides of HIV-1. The results of these two motif searches are summarized in Table V under the PC/Gene protein sequence name PENV HV1BR. The proline-leucine zipper motifs also gave several hits in HIV-1 BRU including 503-525 which is at the very C-terminus of gp120, just upstream of the cleavage site (P7LZIPC and P12LZIPC); and 735-768 in the cytoplasmic domain of gp41 (P23LZIPC). These results are found in Tables VIII, IX, and X under the same sequence name as mentioned above. Notice that the only area of HIV-1 BRU which is predicted by the Lupas algorithm to contain a coiled-coil region, is from amino acids 635-670. This begins eight amino acids N-terminal to the start and ends eight amino acids N-terminal to the end of DP-178. DP-107, despite the fact that it is a known coiled coil, is

not predicted to contain a coiled-coil region using the Lupas method.

11. **EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION OF DP-107-LIKE AND DP-178-LIKE SEQUENCES IN HUMAN RESPIRATORY SYNCYTIAL VIRUS**

FIG. 21 represents search results for Human Respiratory Syncytial Virus (RSV; Strain A2) fusion glycoprotein F1 (PC/Gene protein sequence name PVGLF\_HRSVA). Motif 107x178x4 finds three hits including amino acids 152-202, 213-243, and 488-515. The arrangement of these hits is similar to what is found in HIV-1 except that the motif finds two regions with similarities to DP-178, one just downstream of what would be called the DP-107 region or amino acids 213-243, and one just upstream of the transmembrane region (also similar to DP-178) or amino acids 488-515. Motif ALLMOTI5 also finds three areas including amino acids 116-202, 267-302, and 506-549. The proline-leucine zipper motifs also gave several hits including amino acids 205-221 and 265-287 (P1LZIPC 265-280, P12LZIPC), and 484-513 (P7LZIPC and P12LZIPC 484-506, P23LZIPC). Notice that the PLZIP motifs also identify regions which share location similarities with DP-178 of HIV-1.

12. **EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION OF DP-107-LIKE AND DP-178-LIKE SEQUENCES IN SIMIAN IMMUNODEFICIENCY VIRUS**

Motif hits for Simian immunodeficiency Virus gp41 (AGM3 isolate; PC/Gene protein sequence name PENV\_SIVAG) are shown in FIG. 22. Motif 107x178x4 finds three hits including amino acids 566-593, 597-624, and 703-730. The first two hits only have three amino acids between them and could probably be combined into one hit from 566-624 which would

represent a DP-107-like hit. Amino acids 703 to 730 would then represent a DP-178-like hit. ALLMOTI5 also finds three hits including amino acids 556-628 (DP-107-like), 651-699 (DP-178-like), and 808-852 which represents the transmembrane spanning region. SIV  
5 also has one region from 655-692 with a high propensity to form a coiled coil as predicted by the Lupas algorithm. Both 107x178x4 and ALLMOTI5 motifs find the same region. SIV does not have any PLZIP motif hits in gp41.

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13. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION OF  
DP-107-LIKE AND DP-178 LIKE SEQUENCES  
IN CANINE DISTEMPER VIRUS

Canine Distemper Virus (strain Onderstepoort)

15 fusion glycoprotein F1 (PC/Gene Protein sequence name PVGLF\_CDVO) has regions similar to Human RSV which are predicted to be DP-107-like and DP-178-like (FIG. 23). Motif 107x178x4 highlights one area just C-terminal to the fusion peptide at amino acids 252-293. Amino  
20 acids 252-286 are also predicted to be coiled coil using the Lupas algorithm. Almost 100 amino acids C-terminal to the first region is a DP-178-like area at residues 340-367. ALLMOTI5 highlights three areas of interest including: amino acids 228-297, which  
25 completely overlaps both the Lupas prediction and the DP-107-like 107x178x4 hit; residues 340-381, which overlaps the second 107x178x4 hit; and amino acids 568-602, which is DP178-like in that it is located just N-terminal to the transmembrane region. It also  
30 overlaps another region (residues 570-602) predicted by the Lupas method to have a high propensity to form a coiled coil. Several PLZIP motifs successfully identified areas of interest including P6 and P12LZIPC which highlight residues 336-357 and 336-361  
35 respectively; P1 and P12LZIPC which find residues 398-

414; and P12 and P23LZIPC which find residues 562-589 and 562-592 respectively.

14. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION OF  
DP-107-LIKE AND DP-178-LIKE SEQUENCES  
IN NEWCASTLE DISEASE VIRUS

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FIG. 24 shows the motif hits found in Newcastle Disease Virus (strain Australia-Victoria/32; PC Gene protein sequence name PVGLF\_NDVA). Motif 107x178x4 finds two areas including a DP-107-like hit at amino acids 151-178 and a DP-178-like hit at residues 426-512. ALLMOTI5 finds three areas including residues 117-182, 231-272, and 426-512. The hits from 426-512 include a region which is predicted by the Lupas method to have a high coiled-coil propensity (460-503). The PLZIP motifs identify only one region of interest at amino acids 273-289 (P1 and 12LZIPC).

15. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION  
OF DP-107-LIKE AND DP-178-LIKE  
SEQUENCES IN HUMAN PARAINFLUENZA VIRUS

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Both motifs 107x178x4 and ALLMOTI5 exhibit DP-107-like hits in the same region, 115-182 and 117-182 respectively, of Human Parainfluenza Virus (strain NIH 47885; PC/Gene protein sequence name PVGLF\_p13H4; (FIG. 25). In addition, the two motifs have a DP-178-like hit just slightly C-terminal at amino acids 207-241. Both motifs also have DP-178-like hits nearer the transmembrane region including amino acids 457-497 and 462-512 respectively. Several PLZIP motif hits are also observed including 283-303 (P5LZIPC), 283-310 (P12LZIPC), 453-474 (P6LZIPC), and 453-481 (P23LZIPC). The Lupas algorithm predicts that amino acids 122-176 have a propensity to form a coiled-coil.

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16. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION OF  
DP-107-LIKE AND DP-178-LIKE SEQUENCES OF  
INFLUENZA A VIRUS

FIG. 26 illustrates the Lupas prediction for a coiled coil in Influenza A Virus (strain A/Aichi/2/68) at residues 379-436, as well as the motif hits for 107x178x4 at amino acids 387-453, and for ALLMOTI5 at residues 380-456. Residues 383-471 (38-125 of HA2) were shown by Carr and Kim to be an extended coiled coil when under acidic pH (Carr and Kim, 1993, Cell 73: 823-832). The Lupas algorithm predicts a coiled-coil at residues 379-436. All three methods successfully predicted the region shown to actually have coiled-coil structure; however, ALLMOTI5 predicted the greatest portion of the 88 residue stretch.

17. EXAMPLE: RSV ANTIVIRAL COMPOUNDS

In the Example presented herein, respiratory syncytial virus (RSV) peptide sequences identified by utilizing the computer-assisted coiled-coil peptide sequence searches described in Example 9, above, are shown to encode peptide domains that exhibit structural similarity to actual, known coiled-coil peptides, and are, additionally found to exhibit antiviral activity.

17.1 MATERIALS AND METHODS

Structural analyses consisted of circular dichroism (CD) studies, which were conducted according to the methods described in the Applicants' co-pending U.S. Patent Application Ser. No 08/073,028.

Anti-RSV antiviral activity was assayed as described in Pringle, C.R. et al., 1985, J. Medical Vir. 17:377-386.

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A 48 amino acid RSV F2 peptide and a 53 amino acid RSV T67 peptide are utilized which span sequences that were identified via the computer assisted peptide sequence search strategies described in Example 9, above. See FIG. 21 for the exact position of these sequences and for the motifs utilized.

## 17.2 RESULTS

35-mer oligopeptides were synthesized which constituted portions of the 48 amino acid RSV F2 peptide sequence (FIG. 27) and portions of the 53 amino acid RSV T67 peptide sequence (FIG. 28). The oligopeptides were assayed, via CD analysis, for structural similarity to known coiled-coil structures, and for anti-RSV activity. As shown in FIGS. 27 and 28, a number of these oligopeptides exhibited substantial coiled-coil structural similarity and/or antiviral activity.

Thus, the computer assisted searches described, herein, in Example 9, for example, successfully identified viral peptide domains that represent highly promising anti-RSV antiviral compounds.

## 18. EXAMPLE: HPF3 ANTIVIRAL COMPOUNDS

In the Example presented herein, human parainfluenza virus 3 (HPF3) peptide sequences identified by utilizing the computer-assisted coiled-coil peptide sequence searches described in Example 9, above, are shown to encode peptide domains that exhibit structural similarity to actual, known coiled-coil peptides, and are, additionally found to exhibit antiviral activity.

### 18.1 MATERIALS AND METHODS

Structural analyses consisted of circular dichroism (CD) studies, which were conducted according

to the methods described in the Applicants' co-pending U.S. Patent Application Ser. No 08/073,028.

Anti-HPF3 antiviral activity was assayed as described in Pringle, C.R. et al., 1985, J. Medical Vir. 17:377-386.

5       A 56 amino acid and 70 amino acid HPF3 peptide are utilized which span sequences that were identified via the computer assisted peptide sequence search strategies described in Example 9, above. See FIG. 25 for the exact positions of these sequences and for the  
10 motifs utilized.

## 18.2 RESULTS

35-mer oligopeptides were synthesized which constituted portions of the 56 amino acid HPF3 peptide  
15 sequence (FIG. 29) and portions of the 70 amino acid HPF3 peptide sequence (FIG. 30). The oligopeptides were assayed, via CD analysis, for structural similarity to known coiled-coil structures, and for anti-HPF3 activity. As shown in FIGS. 29 and 30, a  
20 number of these oligopeptides exhibited substantial coiled-coil structural similarity and/or antiviral activity.

Thus, the computer assisted searches described, herein, in Example 9, for example, successfully  
25 identified viral peptide domains that represent highly promising anti-HPF3 antiviral compounds.

The present invention is not to be limited in scope by the specific embodiments described which are  
30 intended as single illustrations of individual aspects of the invention, and functionally equivalent methods and components are within the scope of the invention. Indeed, various modifications of the invention, in addition to those shown and described herein will  
35 become apparent to those skilled in the art from the

foregoing description and accompanying drawings. Such modifications are intended to fall within the scope of the appended claims.

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WHAT IS CLAIMED IS:

1. A peptide having an amino acid sequence corresponding to an  $\alpha$ -helix region of an extracellular domain of a viral envelope protein, which interacts  
5 with and binds to a second  $\alpha$ -helix region of the viral envelope protein containing a leucine-zipper domain having a coiled-coil structure.

2. The peptide of Claim 1 wherein the peptide  
10 is recognized by a computer-assisted peptide sequence search utilizing an ALLMOTI5, 107x178x4 motif, or a PLZIP motif.

3. The peptide of Claim 1 in which the  
15 enveloped virus is a retrovirus.

4. The peptide of Claim 3 in which the retrovirus is a human retrovirus.

20 5. The peptide of Claim 4 in which the human retrovirus is HIV-1 or HIV-2.

6. The peptide of Claim 4 in which the human  
25 retrovirus is HTLV-I or HTLV-II

7. The peptide of Claim 1 in which the enveloped virus is a non-human retrovirus.

8. The peptide of Claim 6 in which the non-  
30 human retrovirus is bovine leukosis virus, feline sarcoma virus, feline leukemia virus, simian immunodeficiency virus, simian sarcoma virus, and sheep progress pneumonia virus.

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9. The peptide of Claim 1 in which the enveloped virus is a non-retroviral virus.

10. The peptide of Claim 9 in which the virus is respiratory syncytial virus, influenza virus,  
 5 parainfluenza virus, canine distemper virus, or newcastle disease virus.

11. A peptide having a formula selected from the group consisting of:

10 X-YTS-Z  
 X-YTSL-Z  
 X-YTSLI-Z  
 X-YTSLIH-Z  
 X-YTSLIHS-Z  
 X-YTSLIHSL-Z  
 X-YTSLIHSLI-Z  
 15 X-YTSLIHSLIE-Z  
 X-YTSLIHSLIEE-Z  
 X-YTSLIHSLIEES-Z  
 X-YTSLIHSLIEESQ-Z  
 X-YTSLIHSLIEESQN-Z  
 X-YTSLIHSLIEESQNQ-Z  
 X-YTSLIHSLIEESQNQQ-Z  
 X-YTSLIHSLIEESQNQQE-Z  
 20 X-YTSLIHSLIEESQNQQEK-Z  
 X-YTSLIHSLIEESQNQQEKN-Z  
 X-YTSLIHSLIEESQNQQEKNE-Z  
 X-YTSLIHSLIEESQNQQEKNEQ-Z  
 X-YTSLIHSLIEESQNQQEKNEQE-Z  
 X-YTSLIHSLIEESQNQQEKNEQEL-Z  
 X-YTSLIHSLIEESQNQQEKNEQELL-Z  
 25 X-YTSLIHSLIEESQNQQEKNEQELLE-Z  
 X-YTSLIHSLIEESQNQQEKNEQELLEL-Z  
 X-YTSLIHSLIEESQNQQEKNEQELLELD-Z  
 X-YTSLIHSLIEESQNQQEKNEQELLELDK-Z  
 X-YTSLIHSLIEESQNQQEKNEQELLELDKW-Z  
 X-YTSLIHSLIEESQNQQEKNEQELLELDKWA-Z  
 X-YTSLIHSLIEESQNQQEKNEQELLELDKWAS-Z  
 X-YTSLIHSLIEESQNQQEKNEQELLELDKWASLW-Z  
 30 X-YTSLIHSLIEESQNQQEKNEQELLELDKWASLWN-Z  
 X-YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNW-Z and  
 X-YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-Z (SEQ ID:1), or

35

5 X-NWF-Z  
 X-WNWF-Z  
 X-LWNWF-Z  
 X-SLWNWF-Z  
 X-ASLWNWF-Z  
 X-WASLWNWF-Z  
 X-KWASLWNWF-Z  
 X-DKWASLWNWF-Z  
 X-LDKWASLWNWF-Z  
 X-ELDKWASLWNWF-Z  
 X-LELDKWASLWNWF-Z  
 X-LLELDKWASLWNWF-Z  
 X-ELLELDKWASLWNWF-Z  
 X-QELLELDKWASLWNWF-Z  
 X-EQELLELDKWASLWNWF-Z  
 10 X-NEQELLELDKWASLWNWF-Z  
 X-KNEQELLELDKWASLWNWF-Z  
 X-EKNEQELLELDKWASLWNWF-Z  
 X-QEKNEQELLELDKWASLWNWF-Z  
 X-QQEKNEQELLELDKWASLWNWF-Z  
 X-NQEKNEQELLELDKWASLWNWF-Z  
 X-QNQEKNEQELLELDKWASLWNWF-Z  
 15 X-SQNQEKNEQELLELDKWASLWNWF-Z  
 X-ESQNQEKNEQELLELDKWASLWNWF-Z  
 X-EESQNQEKNEQELLELDKWASLWNWF-Z  
 X-IEESQNQEKNEQELLELDKWASLWNWF-Z  
 X-SLIEESQNQEKNEQELLELDKWASLWNWF-Z  
 X-HSLIEESQNQEKNEQELLELDKWASLWNWF-Z  
 X-IHSLIEESQNQEKNEQELLELDKWASLWNWF-Z  
 20 X-LIHSLIEESQNQEKNEQELLELDKWASLWNWF-Z  
 X-SLIHSLIEESQNQEKNEQELLELDKWASLWNWF-Z  
 and X-TSLIHSLIEESQNQEKNEQELLELDKWASLWNWF-Z

in which:

25 amino acid residues are presented by the single-  
 letter code;  
 X comprises an amino group, an acetyl group, a 9-  
 fluorenylmethoxy-carbonyl group, a  
 hydrophobic group, or a macromolecule  
 30 carrier group;  
 Z comprises a carboxyl group, an amido group, a  
 hydrophobic group, or a macromolecular  
 carrier group.

35 12. A peptide having a formula selected from the  
 group consisting of:

X-LEA-Z  
 X-LEAN-Z  
 X-LEANI-Z  
 X-LEANIS-Z  
 X-LEANISQ-Z  
 X-LEANISQS-Z  
 X-LEANISQSL-Z  
 5 X-LEANISQSLE-Z  
 X-LEANISQSLEQ-Z  
 X-LEANISQSLEQA-Z  
 X-LEANISQSLEQAQ-Z  
 X-LEANISQSLEQAQI-Z  
 X-LEANISQSLEQAQIQ-Z  
 X-LEANISQSLEQAQIQQ-Z  
 10 X-LEANISQSLEQAQIQQE-Z  
 X-LEANISQSLEQAQIQQEK-Z  
 X-LEANISQSLEQAQIQQEKN-Z  
 X-LEANISQSLEQAQIQQEKNM-Z  
 X-LEANISQSLEQAQIQQEKNMY-Z  
 X-LEANISQSLEQAQIQQEKNMYE-Z  
 X-LEANISQSLEQAQIQQEKNMYEL-Z  
 X-LEANISQSLEQAQIQQEKNMYELQ-Z  
 15 X-LEANISQSLEQAQIQQEKNMYELQK-Z  
 X-LEANISQSLEQAQIQQEKNMYELQKL-Z  
 X-LEANISQSLEQAQIQQEKNMYELQKLN-Z  
 X-LEANISQSLEQAQIQQEKNMYELQKLNS-Z  
 X-LEANISQSLEQAQIQQEKNMYELQKLNSW-Z  
 X-LEANISQSLEQAQIQQEKNMYELQKLNSWD-Z  
 X-LEANISQSLEQAQIQQEKNMYELQKLNSWDV-Z  
 20 X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVF-Z  
 X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTN-Z  
 X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNW-Z and  
 X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z (SEQ ID:7), or

X-NWL-Z  
 X-TNWL-Z  
 25 X-FTNWL-Z  
 X-VFTNWL-Z  
 X-DVFTNWL-Z  
 X-WDVFTNWL-Z  
 X-SWDVFTNWL-Z  
 X-NSWDVFTNWL-Z  
 X-LNSWDVFTNWL-Z  
 30 X-KLNSWDVFTNWL-Z  
 X-QKLNSWDVFTNWL-Z  
 X-LQKLNSWDVFTNWL-Z  
 X-ELQKLNSWDVFTNWL-Z  
 X-YELQKLNSWDVFTNWL-Z  
 X-MYELQKLNSWDVFTNWL-Z  
 X-NMYELQKLNSWDVFTNWL-Z  
 X-KNMYELQKLNSWDVFTNWL-Z  
 35 X-EKNMYELQKLNSWDVFTNWL-Z  
 X-QEKNMYELQKLNSWDVFTNWL-Z

X-QQEKMYELQKLNSWDVFTNWL-Z  
 X-IQQEKMYELQKLNSWDVFTNWL-Z  
 X-QIQQEKMYELQKLNSWDVFTNWL-Z  
 X-AQIQQEKMYELQKLNSWDVFTNWL-Z  
 X-QAQIQQEKMYELQKLNSWDVFTNWL-Z  
 X-EAQIQQEKMYELQKLNSWDVFTNWL-Z  
 X-LEAQIQQEKMYELQKLNSWDVFTNWL-Z  
 X-SLEAQIQQEKMYELQKLNSWDVFTNWL-Z  
 X-QKSLEAQIQQEKMYELQKLNSWDVFTNWL-Z  
 X-SQSLEAQIQQEKMYELQKLNSWDVFTNWL-Z  
 X-ISQSLEAQIQQEKMYELQKLNSWDVFTNWL-Z  
 X-NISQSLEAQIQQEKMYELQKLNSWDVFTNWL-Z  
 X-ANISQSLEAQIQQEKMYELQKLNSWDVFTNWL-Z  
 and X-EANISQSLEAQIQQEKMYELQKLNSWDVFTNWL-Z

5

10

in which:

amino acid residues are presented by the single-letter code;

15

X comprises an amino group, an acetyl group, a 9-fluoromethoxymethyl-carbonyl group, a hydrophobic group, or a macromolecule carrier group;

20

Z comprises a carboxyl group, an amido group, a hydrophobic group, or a macromolecular carrier group.

13. A peptide having a formula selected from the group consisting of:

X-YTS-Z  
 X-YTSV-Z  
 25 X-YTSVI-Z  
 X-YTSVIT-Z  
 X-YTSVITI-Z  
 X-YTSVITIE-Z  
 X-YTSVITIEL-Z  
 X-YTSVITIELS-Z  
 X-YTSVITIELSN-Z  
 X-YTSVITIELSNI-Z  
 30 X-YTSVITIELSNIK-Z  
 X-YTSVITIELSNIKE-Z  
 X-YTSVITIELSNIKEN-Z  
 X-YTSVITIELSNIKENK-Z  
 X-YTSVITIELSNIKENKC-Z  
 X-YTSVITIELSNIKENKCN-Z  
 X-YTSVITIELSNIKENKCNG-Z  
 35 X-YTSVITIELSNIKENKCNGT-Z  
 X-YTSVITIELSNIKENKCNGTD-Z

X-YTSVITIELSNIKENKCNGTDA-Z  
 X-YTSVITIELSNIKENKCNGTDAK-Z  
 X-YTSVITIELSNIKENKCNGTDAKV-Z  
 X-YTSVITIELSNIKENKCNGTDAKVK-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKL-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLI-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIK-Z  
 5 X-YTSVITIELSNIKENKCNGTDAKVKLIQ-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQE-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQEL-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELD-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDK-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKY-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYK-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKN-Z  
 10 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNA-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAV-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTE-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTEL-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELQ-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELQLL-Z  
 15 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELQLLM-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELQLLMQ-Z  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELQLLMQS-Z and  
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELQLLMQST-Z, or  
  
 X-QST-Z  
 X-MQST-Z  
 20 X-LMQST-Z  
 X-LLMQST-Z  
 X-QLLMQST-Z  
 X-LQLLMQST-Z  
 X-ELQLLMQST-Z  
 X-TELQLLMQST-Z  
 X-VTELQLLMQST-Z  
 X-AVTELQLLMQST-Z  
 25 X-NAVTELQLLMQST-Z  
 X-KNAVTELQLLMQST-Z  
 X-YKNAVTELQLLMQST-Z  
 X-KYKNAVTELQLLMQST-Z  
 X-DKYKNAVTELQLLMQST-Z  
 X-LDKYKNAVTELQLLMQST-Z  
 X-ELDKYKNAVTELQLLMQST-Z  
 30 X-QELDKYKNAVTELQLLMQST-Z  
 X-KQELDKYKNAVTELQLLMQST-Z  
 X-IKQELDKYKNAVTELQLLMQST-Z  
 X-LIKQELDKYKNAVTELQLLMQST-Z  
 X-KLIKQELDKYKNAVTELQLLMQST-Z  
 X-VKLIKQELDKYKNAVTELQLLMQST-Z  
 X-KVKLIKQELDKYKNAVTELQLLMQST-Z  
 35 X-AKVLIKQELDKYKNAVTELQLLMQST-Z  
 X-DAKVLIKQELDKYKNAVTELQLLMQST-Z

X-TDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-GTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-NGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-CNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-KCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-NKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-ENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 5 X-KENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-IKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-NIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-SNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-LSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-ELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-IELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 10 X-TIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-ITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-VITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-SVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z  
 X-TSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST-Z

in which:

15 amino acid residues are presented by the single-  
 letter code;  
 X comprises an amino group, an acetyl group, a 9-  
 fluoromethoxymethyl-carbonyl group, a  
 hydrophobic group, or a macromolecule  
 carrier group;  
 20 Z comprises a carboxyl group, an amido group, a  
 hydrophobic group, or a macromolecular  
 carrier group.

25 14. A peptide having a formula selected from the  
 group consisting of:

X-FYD-Z  
 X-FYDP-Z  
 X-FYDPL-Z  
 X-FYDPLV-Z  
 X-FYDPLVF-Z  
 30 X-FYDPLVFP-Z  
 X-FYDPLVFPS-Z  
 X-FYDPLVFPSD-Z  
 X-FYDPLVFPSDE-Z  
 X-FYDPLVFPSDEF-Z  
 X-FYDPLVFPSDEFD-Z  
 X-FYDPLVFPSDEFDA-Z  
 35 X-FYDPLVFPSDEFDAS-Z  
 X-FYDPLVFPSDEFDASI-Z

X-FYDPLVFPSEFDASIS-Z  
 X-FYDPLVFPSEFDASISQ-Z  
 X-FYDPLVFPSEFDASISQV-Z  
 X-FYDPLVFPSEFDASISQVN-Z  
 X-FYDPLVFPSEFDASISQVNE-Z  
 X-FYDPLVFPSEFDASISQVNEK-Z  
 X-FYDPLVFPSEFDASISQVNEKI-Z  
 5 X-FYDPLVFPSEFDASISQVNEKIN-Z  
 X-FYDPLVFPSEFDASISQVNEKINQ-Z  
 X-FYDPLVFPSEFDASISQVNEKINQS-Z  
 X-FYDPLVFPSEFDASISQVNEKINQSL-Z  
 X-FYDPLVFPSEFDASISQVNEKINQSLA-Z  
 X-FYDPLVFPSEFDASISQVNEKINQSLAF-Z  
 X-FYDPLVFPSEFDASISQVNEKINQSLAFI-Z  
 10 X-FYDPLVFPSEFDASISQVNEKINQSLAFIR-Z  
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRK-Z  
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRKS-Z  
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRKSD-Z  
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRKSDE-Z  
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRKSDEL-Z  
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRKSDELL-Z, or  
  
 15 X-DELL-Z  
 X-SDELL-Z  
 X-KSDELL-Z  
 X-RKSDELL-Z  
 X-IRKSDELL-Z  
 X-FIRKSDELL-Z  
 X-AFIRKSDELL-Z  
 X-LAFIRKSDELL-Z  
 20 X-SLAFIRKSDELL-Z  
 X-QSLAFIRKSDELL-Z  
 X-NQSLAFIRKSDELL-Z  
 X-INQSLAFIRKSDELL-Z  
 X-KINQSLAFIRKSDELL-Z  
 X-EKINQSLAFIRKSDELL-Z  
 X-NEKINQSLAFIRKSDELL-Z  
 25 X-VNEKINQSLAFIRKSDELL-Z  
 X-QVNEKINQSLAFIRKSDELL-Z  
 X-SQVNEKINQSLAFIRKSDELL-Z  
 X-ISQVNEKINQSLAFIRKSDELL-Z  
 X-SISQVNEKINQSLAFIRKSDELL-Z  
 X-ASISQVNEKINQSLAFIRKSDELL-Z  
 X-DASISQVNEKINQSLAFIRKSDELL-Z  
 X-FDASISQVNEKINQSLAFIRKSDELL-Z  
 30 X-EFDASISQVNEKINQSLAFIRKSDELL-Z  
 X-DEFDASISQVNEKINQSLAFIRKSDELL-Z  
 X-SDEFDASISQVNEKINQSLAFIRKSDELL-Z  
 X-PSDEFDASISQVNEKINQSLAFIRKSDELL-Z  
 X-FPSDEFDASISQVNEKINQSLAFIRKSDELL-Z  
 X-VFPSDEFDASISQVNEKINQSLAFIRKSDELL-Z  
 X-LVFPSEFDASISQVNEKINQSLAFIRKSDELL-Z  
 35 X-PLVFPSEFDASISQVNEKINQSLAFIRKSDELL-Z  
 X-DPLVFPSEFDASISQVNEKINQSLAFIRKSDELL-Z



X-YDPLVFPSEFDASISQVNEKINQSLAFIRKSDELL-Z

in which:

amino acid residues are presented by the single-letter code;

5

X comprises an amino group, an acetyl group, a 9-fluoromethoxymethyl-carbonyl group, a hydrophobic group, or a macromolecule carrier group;

10

Z comprises a carboxyl group, an amido group, a hydrophobic group, or a macromolecular carrier group.

15 15. A peptide having a formula selected from the group consisting of:

X-ITL-Z  
 X-ITLN-Z  
 X-ITLNN-Z  
 X-ITLNNS-Z  
 X-ITLNNSV-Z  
 X-ITLNNSVA-Z  
 20 X-ITLNNSVAL-Z  
 X-ITLNNSVALD-Z  
 X-ITLNNSVALDP-Z  
 X-ITLNNSVALDPI-Z  
 X-ITLNNSVALDPID-Z  
 X-ITLNNSVALDPIDI-Z  
 X-ITLNNSVALDPIDIS-Z  
 X-ITLNNSVALDPIDISI-Z  
 25 X-ITLNNSVALDPIDISIE-Z  
 X-ITLNNSVALDPIDISIEL-Z  
 X-ITLNNSVALDPIDISIELN-Z  
 X-ITLNNSVALDPIDISIELNK-Z  
 X-ITLNNSVALDPIDISIELNKA-Z  
 X-ITLNNSVALDPIDISIELNKAK-Z  
 X-ITLNNSVALDPIDISIELNKAKS-Z  
 X-ITLNNSVALDPIDISIELNKAKSD-Z  
 30 X-ITLNNSVALDPIDISIELNKAKSDL-Z  
 X-ITLNNSVALDPIDISIELNKAKSDLE-Z  
 X-ITLNNSVALDPIDISIELNKAKSDLEE-Z  
 X-ITLNNSVALDPIDISIELNKAKSDLEES-Z  
 X-ITLNNSVALDPIDISIELNKAKSDLEESK-Z  
 X-ITLNNSVALDPIDISIELNKAKSDLEESKE-Z  
 X-ITLNNSVALDPIDISIELNKAKSDLEESKEW-Z  
 35 X-ITLNNSVALDPIDISIELNKAKSDLEESKEWI-Z  
 X-ITLNNSVALDPIDISIELNKAKSDLEESKEWIR-Z

X-ITLNNVALDPIDISIELNKAUSDLEESKEWIRR-Z  
 X-ITLNNVALDPIDISIELNKAUSDLEESKEWIRRS-Z, or

5 X-RRS-Z  
 X-IRRS-Z  
 X-WIRRS-Z  
 X-EWIRRS-Z  
 X-KEWIRRS-Z  
 X-SKEWIRRS-Z  
 X-ESKEWIRRS-Z  
 X-EESKEWIRRS-Z  
 X-LEESKEWIRRS-Z  
 X-DLEESKEWIRRS-Z  
 X-SDLEESKEWIRRS-Z  
 10 X-KSDLEESKEWIRRS-Z  
 X-AKSDLEESKEWIRRS-Z  
 X-KAKSDLEESKEWIRRS-Z  
 X-NKAKSDLEESKEWIRRS-Z  
 X-LNKAKSDLEESKEWIRRS-Z  
 X-ELNKAKSDLEESKEWIRRS-Z  
 X-IELNKAKSDLEESKEWIRRS-Z  
 X-SIELNKAKSDLEESKEWIRRS-Z  
 15 X-ISIELNKAKSDLEESKEWIRRS-Z  
 X-DISIELNKAKSDLEESKEWIRRS-Z  
 X-IDISIELNKAKSDLEESKEWIRRS-Z  
 X-PIDISIELNKAKSDLEESKEWIRRS-Z  
 X-DPIDISIELNKAKSDLEESKEWIRRS-Z  
 X-LDPIDISIELNKAKSDLEESKEWIRRS-Z  
 X-ALDPIDISIELNKAKSDLEESKEWIRRS-Z  
 20 X-VALDPIDISIELNKAKSDLEESKEWIRRS-Z  
 X-SVALDPIDISIELNKAKSDLEESKEWIRRS-Z  
 X-NSVALDPIDISIELNKAKSDLEESKEWIRRS-Z  
 X-NNSVALDPIDISIELNKAKSDLEESKEWIRRS-Z  
 X-LNNSVALDPIDISIELNKAKSDLEESKEWIRRS-Z  
 X-TLNNVALDPIDISIELNKAKSDLEESKEWIRRS-Z

in which:

25 amino acid residues are presented by the single-  
 letter code;  
 X comprises an amino group, an acetyl group, a 9-  
 fluoromethoxymethyl-carbonyl group, a  
 hydrophobic group, or a macromolecule  
 30 carrier group;  
 Z comprises a carboxyl group, an amido group, a  
 hydrophobic group, or a macromolecular  
 carrier group.

35

16. A peptide having a formula selected from the group consisting of:

- X-ALG-Z  
 X-ALGV-Z  
 X-ALGVA-Z  
 X-ALGVAT-Z  
 5 X-ALGVATS-Z  
 X-ALGVATSA-Z  
 X-ALGVATSAQ-Z  
 X-ALGVATSAQI-Z  
 X-ALGVATSAQIT-Z  
 X-ALGVATSAQITA-Z  
 X-ALGVATSAQITAA-Z  
 10 X-ALGVATSAQITA-AV-Z  
 X-ALGVATSAQITA-AVA-Z  
 X-ALGVATSAQITA-AVAL-Z  
 X-ALGVATSAQITA-AVALV-Z  
 X-ALGVATSAQITA-AVALVE-Z  
 X-ALGVATSAQITA-AVALVEA-Z  
 X-ALGVATSAQITA-AVALVEAK-Z  
 X-ALGVATSAQITA-AVALVEAKQ-Z  
 15 X-ALGVATSAQITA-AVALVEAKQA-Z  
 X-ALGVATSAQITA-AVALVEAKQAR-Z  
 X-ALGVATSAQITA-AVALVEAKQARS-Z  
 X-ALGVATSAQITA-AVALVEAKQARSD-Z  
 X-ALGVATSAQITA-AVALVEAKQARSDI-Z  
 X-ALGVATSAQITA-AVALVEAKQARSDIE-Z  
 X-ALGVATSAQITA-AVALVEAKQARSDIEK-Z  
 20 X-ALGVATSAQITA-AVALVEAKQARSDIEKL-Z  
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLK-Z  
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLKE-Z  
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLKEA-Z  
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLKEAI-Z  
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLKEAIR-Z  
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLKEAIRD-Z, or  
 25 X-IRD-Z  
 X-AIRD-Z  
 X-EAIRD-Z  
 X-KEAIRD-Z  
 X-LKEAIRD-Z  
 X-KLKEAIRD-Z  
 X-EKLKEAIRD-Z  
 X-IEKLKEAIRD-Z  
 30 X-DIEKLKEAIRD-Z  
 X-SDIEKLKEAIRD-Z  
 X-RSDIEKLKEAIRD-Z  
 X-ARSDIEKLKEAIRD-Z  
 X-QARSDIEKLKEAIRD-Z  
 X-KQARSDIEKLKEAIRD-Z  
 X-AKQARSDIEKLKEAIRD-Z  
 35 X-EAKQARSDIEKLKEAIRD-Z  
 X-VEAKQARSDIEKLKEAIRD-Z

X-LVEAKQARSDIEKLKEAIRD-Z  
 X-ALVEAKQARSDIEKLKEAIRD-Z  
 X-VALVEAKQARSDIEKLKEAIRD-Z  
 X-AVALVEAKQARSDIEKLKEAIRD-Z  
 X-AAVALVEAKQARSDIEKLKEAIRD-Z  
 X-TAAVALVEAKQARSDIEKLKEAIRD-Z  
 X-ITAAVALVEAKQARSDIEKLKEAIRD-Z  
 5 X-QITAVALVEAKQARSDIEKLKEAIRD-Z  
 X-AQITAVALVEAKQARSDIEKLKEAIRD-Z  
 X-SAQITAVALVEAKQARSDIEKLKEAIRD-Z  
 X-TSAQITAVALVEAKQARSDIEKLKEAIRD-Z  
 X-ATSAQITAVALVEAKQARSDIEKLKEAIRD-Z  
 X-VATSAQITAVALVEAKQARSDIEKLKEAIRD-Z  
 X-GVATSAQITAVALVEAKQARSDIEKLKEAIRD-Z  
 10 X-LGVATSAQITAVALVEAKQARSDIEKLKEAIRD-Z

in which:

amino acid residues are presented by the single-letter code;

X comprises an amino group, an acetyl group, a 9-fluoromethoxymethyl-carbonyl group, a hydrophobic group, or a macromolecule carrier group;

Z comprises a carboxyl group, an amido group, a hydrophobic group, or a macromolecular carrier group.

17. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein X is a hydrophobic group.

18. The peptide of Claim 17 wherein the hydrophobic group X is carbobenzoxyl, dansyl, or t-butyloxycarbonyl.

19. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein Z is a hydrophobic group.

20. The peptide of Claim 19 wherein the hydrophobic group Z is t-butyloxycarbonyl.

35

21. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein X is a macromolecular carrier group.

22. The peptide of Claim 21 wherein the macromolecular carrier group is a lipid-fatty acid conjugate, a polyethylene glycol, or a carbohydrate moiety.

23. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein Z is a macromolecular carrier group.

24. The peptide of Claim 23 wherein the macromolecular carrier group Z is a lipid-fatty acid conjugate, a polyethylene glycol, or a carbohydrate moiety.

25. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein at least one bond linking adjacent amino acid residues is a non-peptide bond.

26. The peptide of Claim 25 wherein the non-peptide bond is an imino, ester, hydrazine, semicarbazide, or azo bond.

27. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein at least one amino acid residue is in a D-isomer configuration.

28. The peptide of Claim 11, 12, 13, 14, 15 or 16 further comprising at least one amino acid insertion.

29. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein the amino acid insertion is between 1 and 15 amino acid residues.

30. The peptide of Claim 11, 12, 13, 14, 15 or 16 having at least one less amino acid residue, wherein the amino acid residue(s) represents an amino acid deletion, and wherein the peptide comprises at least three amino acid residues.

5

31. The peptide of Claim 11, 12, 13, 14, 15 or 16 further comprising at least one amino acid substitution wherein a first amino acid residue is substituted for a second, different amino acid residue.

10

32. The peptide of Claim 31 wherein the amino acid substitution is a conserved substitution.

15

33. The peptide of Claim 31 wherein the amino acid substitution is a non-conserved substitution.

20

34. A method for the inhibition of transmission of an enveloped virus to a cell, comprising contacting the cell with an effective concentration of the peptide of Claim 1 for an effective period of time so that no infection of the cell by the virus occurs.

25

35. A method for neutralizing an enveloped virus in a host, comprising administering to the host an effective concentration of the peptide of Claim 1 so that the host raises an immune response sufficient to neutralize the virus, and viral infection of uninfected cells in the host is inhibited.

30

36. A method for neutralizing an enveloped virus in a host, comprising administering to the host an effective concentration of an antibody raised against the peptide of Claim 1 so that viral infection of uninfected cells in the host is inhibited.

35

37. A method for the detection of an enveloped virus comprising:

contacting a viral isolate with an effective concentration of the peptide of Claim 1 for an effective amount of time so that viral infectivity is inhibited; and

assaying the viral isolate for viral enzyme activity.

38. A method for the inhibition of transmission of an HIV retrovirus to a cell, comprising contacting the cell with an effective concentration of the peptide of Claim 11 or 12 for an effective period of time so that no infection of the cell by the retrovirus occurs.

39. A method for neutralizing an HIV retrovirus in a host, comprising administering to the host an effective concentration of the peptide of Claim 11 or 12 so that the host raises an immune response sufficient to neutralize the HIV retrovirus, and HIV infection of uninfected cells in the host is inhibited.

40. A method for neutralizing an HIV retrovirus in a host, comprising administering to the host an effective concentration of an antibody raised against the peptide of Claim 11 or 12 so that HIV infection of uninfected cells in the host is inhibited.

41. A method for the detection of HIV, comprising:

contacting a viral isolate with an effective concentration of the peptide of Claim 11 or 12 for an effective amount of time so that HIV viral infectivity is inhibited; and

assaying the viral isolat for retroviral enzyme activity.

42. A method for the inhibition of transmission of a respiratory syncytial virus to a cell, comprising  
5 contacting the cell with an effective concentration of the peptide of Claim 13 or 14 for an effective period of time so that no infection of the cell by the virus occurs.

10 43. A method for neutralizing a respiratory syncytial virus in a host, comprising administering to the host an effective concentration of the peptide of Claim 13 or 14 so that the host raises an immune  
15 response sufficient to neutralize the virus, and respiratory syncytial virus infection of uninfected cells in the host is inhibited.

44. A method for neutralizing a respiratory syncytial virus in a host comprising administering to  
20 the host an effective concentration of an antibody raised against the peptide of Claim 13 or 14 so that respiratory syncytial virus infection of uninfected cells in the host is inhibited.

25 45. A method for the detection of respiratory syncytial virus comprising:  
contacting a viral isolate with an effective concentration of the peptide of Claim 13 or 14 for an effective amount of time so that respiratory syncytial  
30 viral infectivity is inhibited; and  
assaying the viral isolate for respiratory syncytial virus enzyme activity.

46. A method for the inhibition of transmission  
35 of a parainfluenza virus to a cell comprising,



contacting the cell with an effective concentration of the peptide of Claim 15 or 16 for an effective period of time so that no infection of the cell by the virus occurs.

5           47. A method for neutralizing a parainfluenza virus in a host, comprising administering to the host an effective concentration of the peptide of Claim 15 or 16 so that the host raises an immune response  
10           sufficient to neutralize the virus, and parainfluenza infection of uninfected cells in the host is inhibited.

          48. A method for neutralizing a parainfluenza virus in a host comprising administering to the host  
15           an effective concentration of an antibody raised against the peptide of Claim 15 or 16 so that parainfluenza infection of uninfected cells in the host is inhibited.

20           49. A method for the detection of parainfluenza virus comprising:

          contacting a viral isolate with an effective concentration of the peptide of Claim 15 or 16 for an effective amount of time so that parainfluenza viral  
25           infectivity is inhibited; and

          assaying the viral isolate for parainfluenza virus enzyme activity.

30

35

HIV1LAI (DP-178; SEQ ID:1)	YTSLIHSLIEESNQQEKNEQELLELDKWASLWNWF
HIV1SF2 (DP-185; SEQ ID:3)	YTNTIYNLLEESNQQEKNEQELLELDKWASLWNWF
HIV1RF (SEQ ID:4)	YTGIIYNLLEESNQQEKNEQELLELDKWANLWNWF
HIV1MN (SEQ ID:5)	YTSLIYSLLEKSTQQEKNEQELLELDKWASLWNWF
HIV2ROD (SEQ ID:6)	LEANISKSLEQAQIQQEKNMYELOKLSWDIFGNWF
HIV2NIHZ (SEQ ID:7)	LEANISQSLEQAQIQQEKNMYELOKLSWDVFTNWL
DP180 (SEQ ID:2)	SSSFLLLEQNNMKLQAEQMLEQINEKHYLEDIS
DP118 (SEQ ID:10)	QQLLDVVKRQQEMLRLTVHGTKNLQARVTAIEKYLKDQ
DP125 (SEQ ID:8)	CCGNLLRAIEAQCHLLQLTVHGIKQLQARILAVERYLKDQ
DP116 (SEQ ID:9)	LQARILAVERYLKDQQQ

FIG.1

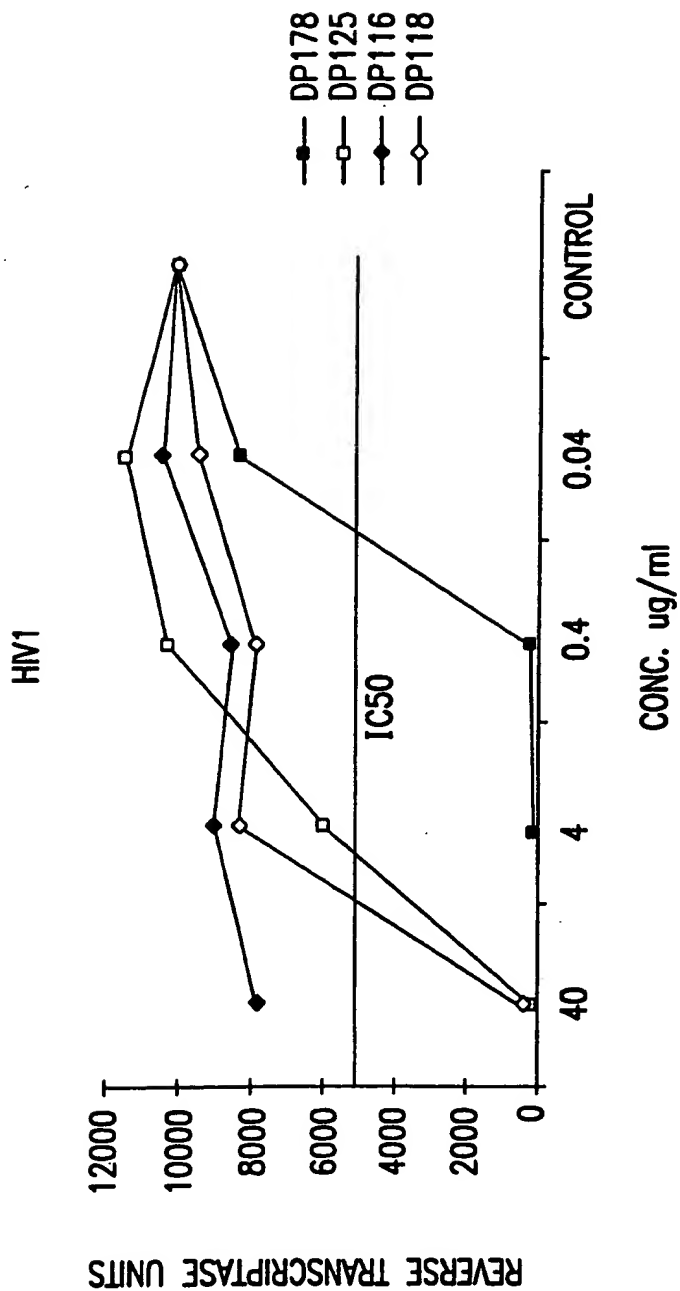


FIG.2

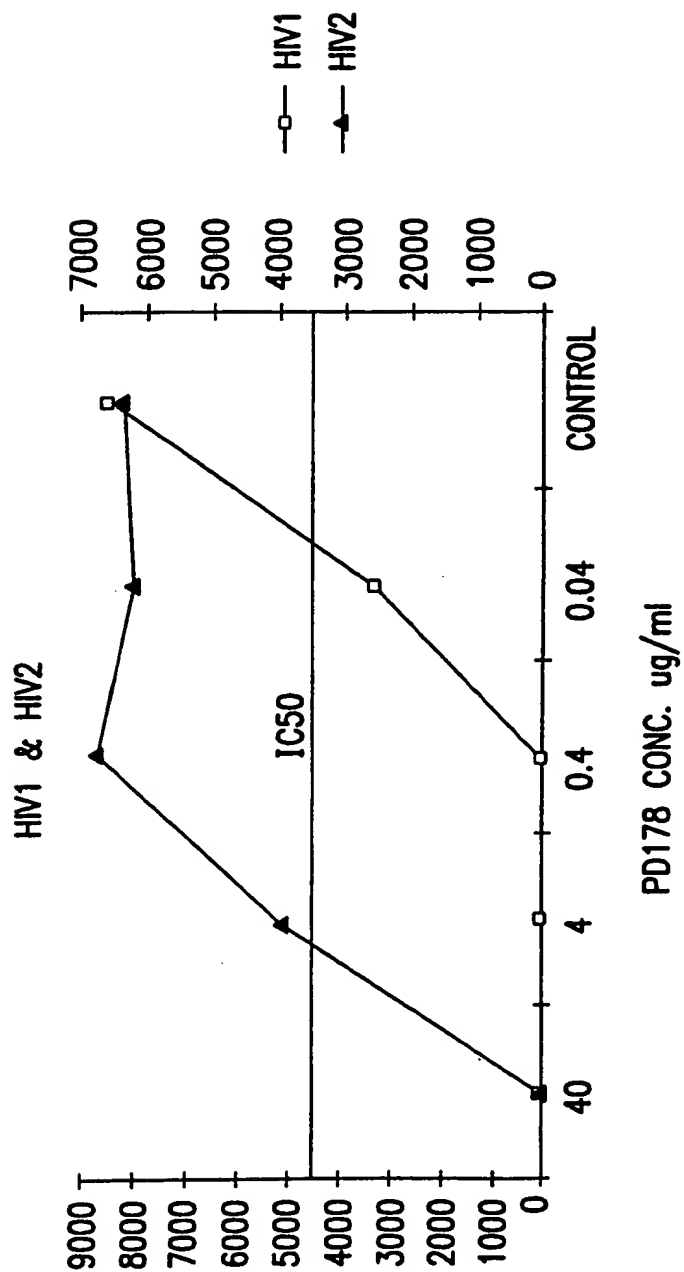


FIG.3

Number of Syncytia/well: concentration in $\mu\text{g/ml}$ (micrograms/ml)									
DP178	10	5	1	0.2	0.1	0.05	0.025	0.0125	Control
<i>Syncytia</i>									
HIV1LAT	0	0	0	0	0	0	0	0	67
HIV1MN	0	0	0	0	0	ND	ND	ND	34
HIV1RF	0	0	0	0	0	ND	ND	ND	65
HIV1SF2	0	0	0	0	0	ND	ND	ND	58
DP125	10	5	1	0.2	0.1	0.05	0.025	0.0125	Control
<i>Syncytia</i>									
HIV1LAT	0	0	54	69	80	75	79	82	67
HIV1MN	0	0	30	36	ND	ND	ND	ND	34
HIV1RF	0	0	67	63	ND	ND	ND	ND	65
HIV1SF2	0	0	9	66	ND	ND	ND	ND	58
DP116	10	5	1	0.2	0.1	0.05	0.025	0.0125	Control
<i>Syncytia</i>									
HIV1LAT	75	ND	ND	ND	ND	ND	ND	ND	67
HIV1MN	35	ND	ND	ND	ND	ND	ND	ND	34
HIV1RF	81	ND	ND	ND	ND	ND	ND	ND	65
HIV1SF2	81	ND	ND	ND	ND	ND	ND	ND	58

FIG.4A

DP180	40	20	10	5	2.5	1.25	0.625	0.3125	Control
<i>Syncytia</i>									
HIV1LAT	50	>45	>45	>45	>45	>45	>45	>45	58
DP185	40	20	10	5	2.5	1.25	0.625	0.3125	Control
<i>Syncytia</i>									
HIV1LAT	0	0	0	0	0	0	0	ND	60

FIG.4B

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<u>HIV1</u>								
Number of Syncytia/well: concentration in ng/ml (nanograms/ml)								
DP178	20	10	5	2.5	1.25	0.625	0.3125	Control
<u>Syncytia</u>								
HIV1	0	0	0	0	0	14	20	48
DP116	20	10	5	2.5	1.25	0.625	0.3125	Control
<u>Syncytia</u>								
HIV1	ND	48	ND	ND	ND	ND	ND	ND
<u>HIV2</u>								
Number of Syncytia/well: concentration in $\mu$ g/ml (micrograms/ml)								
DP178	20	10	5	2.5	1.25	0.625	0.3125	Control
<u>Syncytia</u>								
HIV2	50	54	55	57	63	77	78	76
DP116	20	10	5	2.5	1.25	0.625	0.3125	Control
<u>Syncytia</u>								
HIV2	ND	58	ND	ND	ND	ND	ND	ND

FIG.5

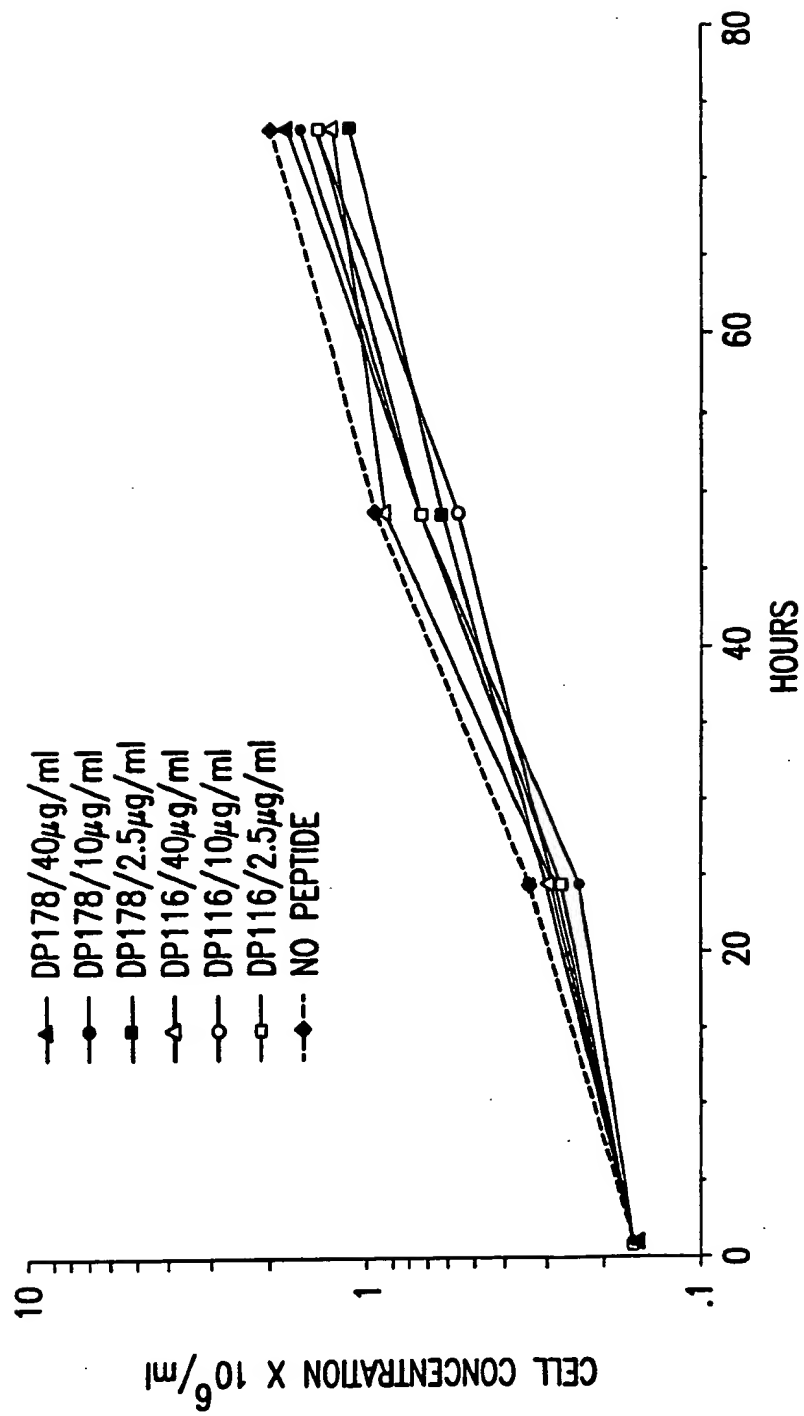


FIG. 6

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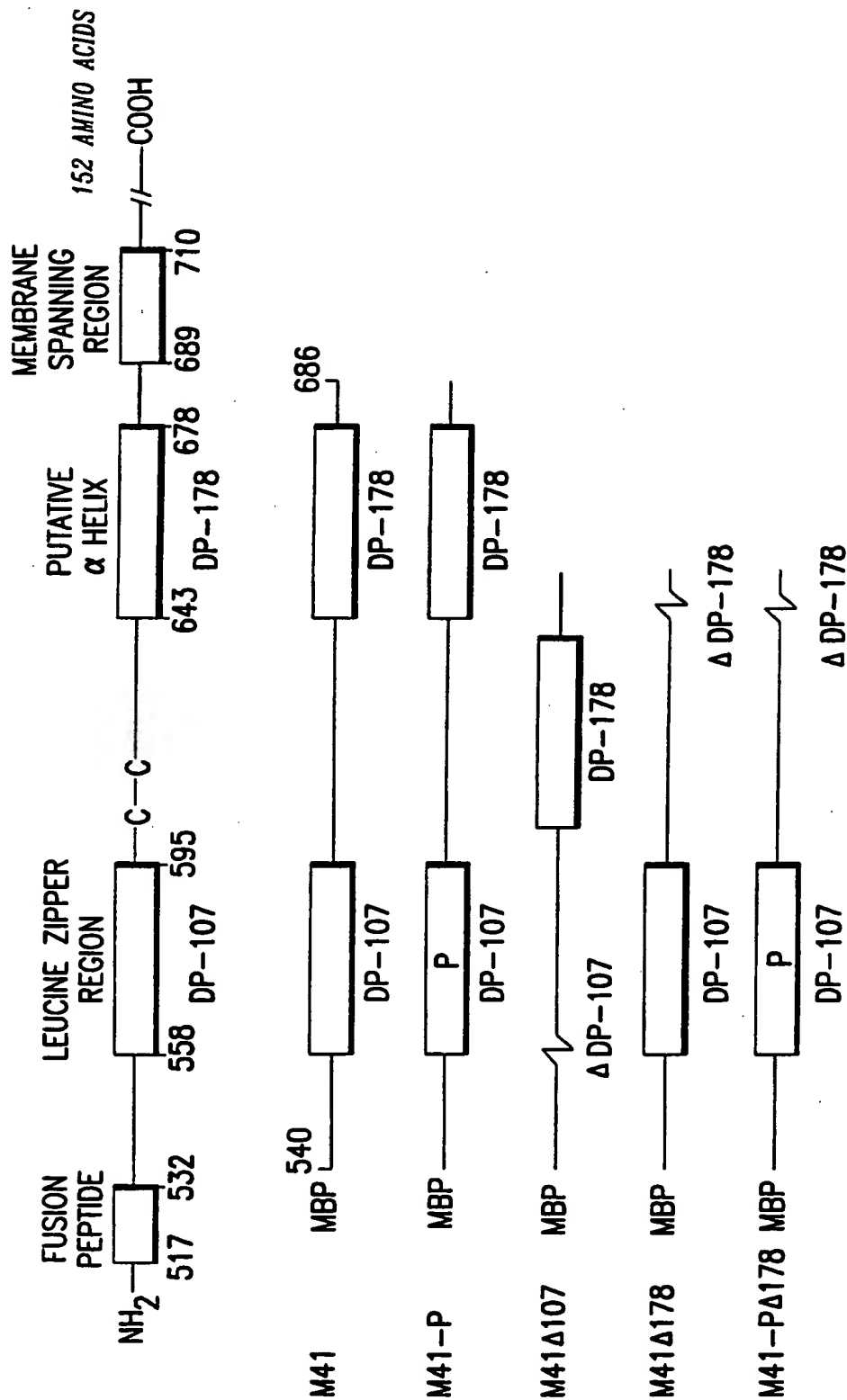


FIG.7



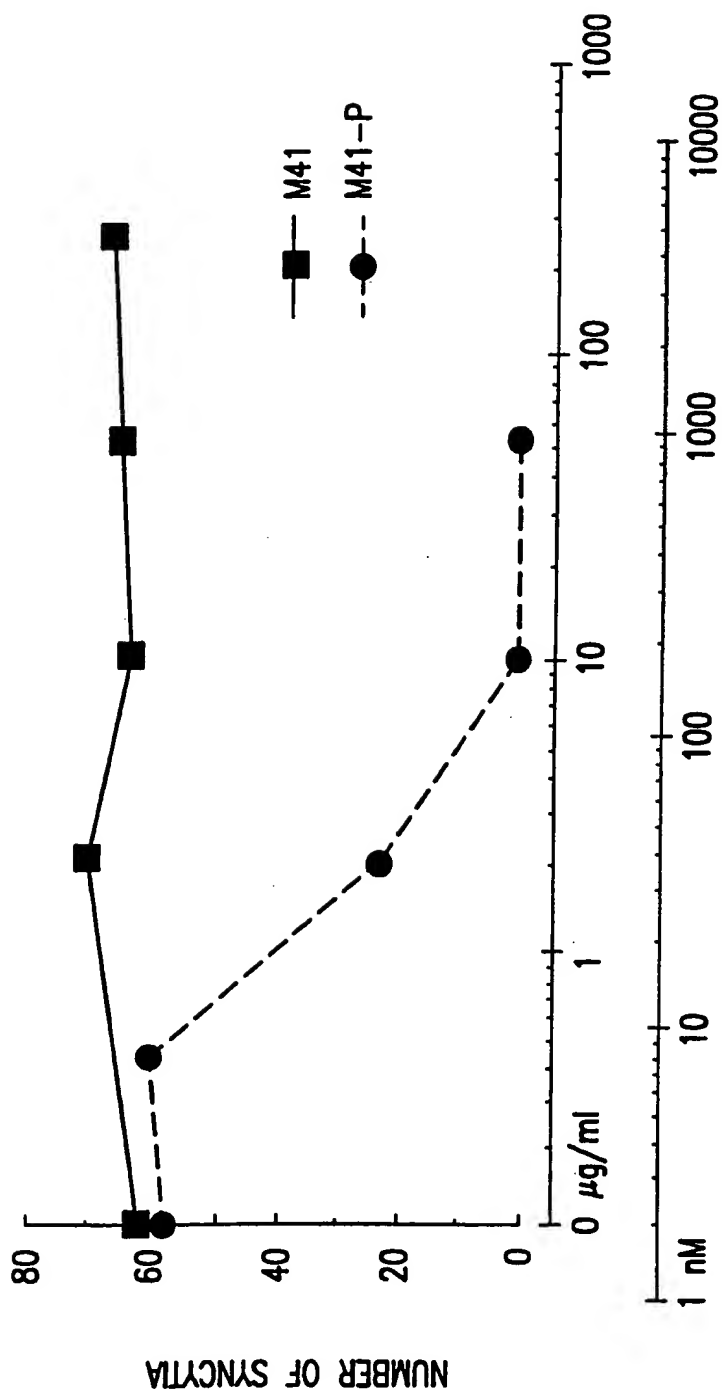


FIG.8

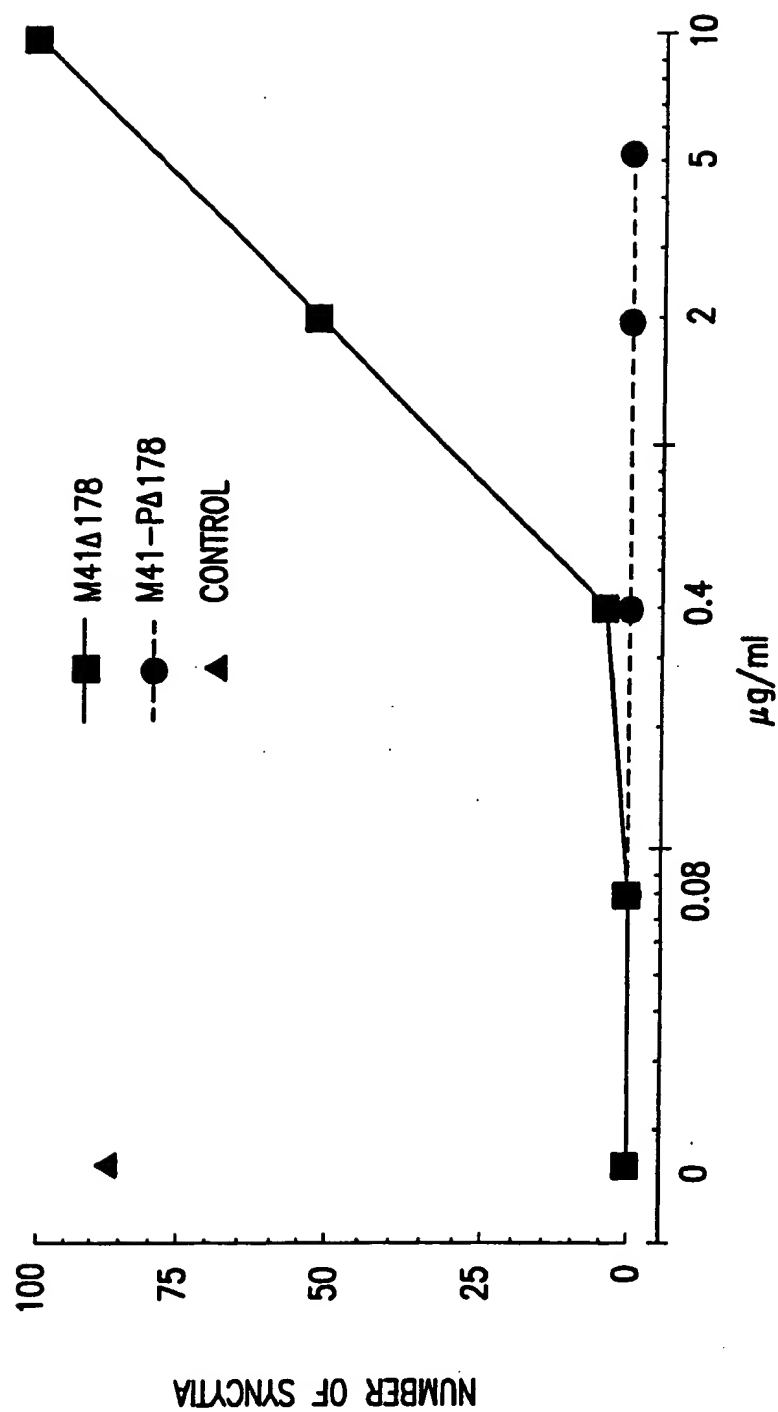


FIG.9

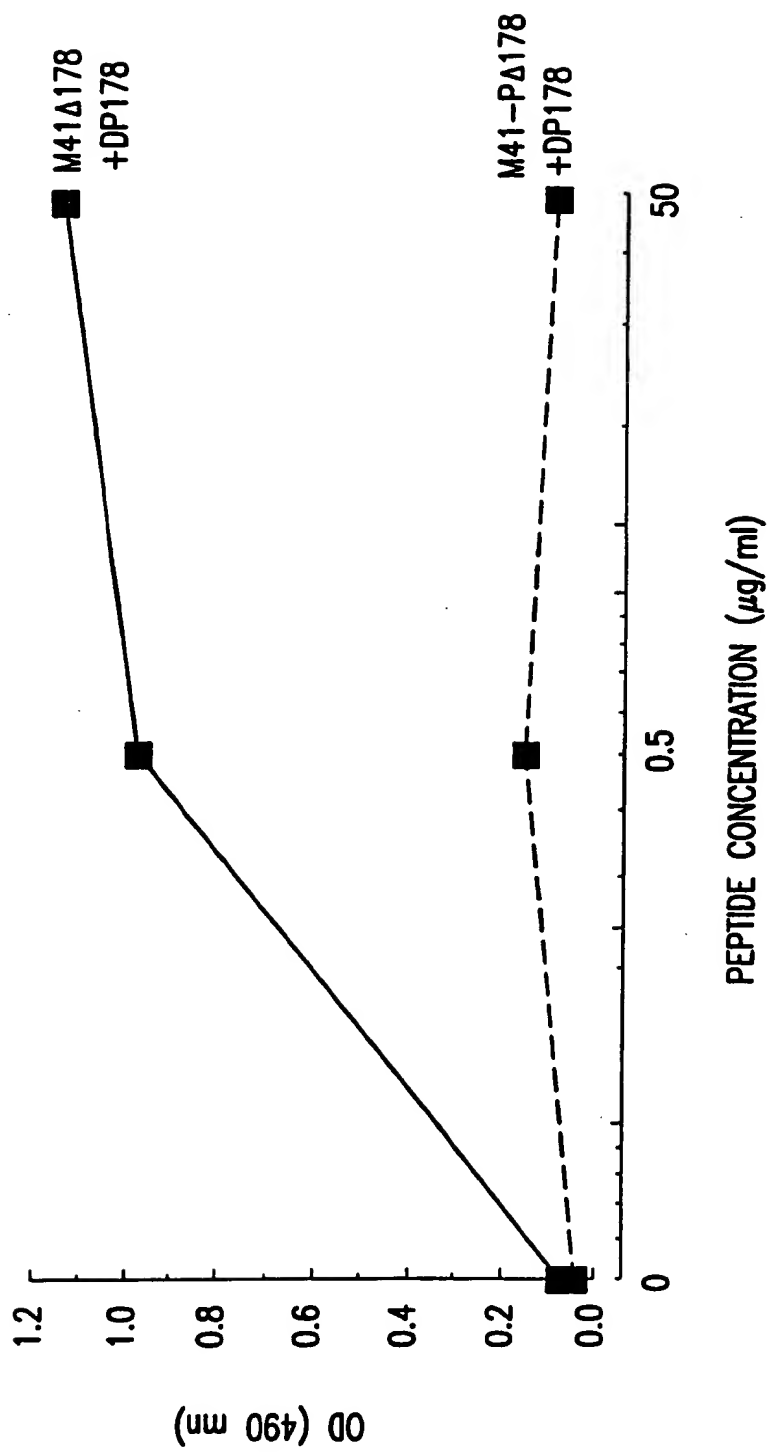


FIG.10

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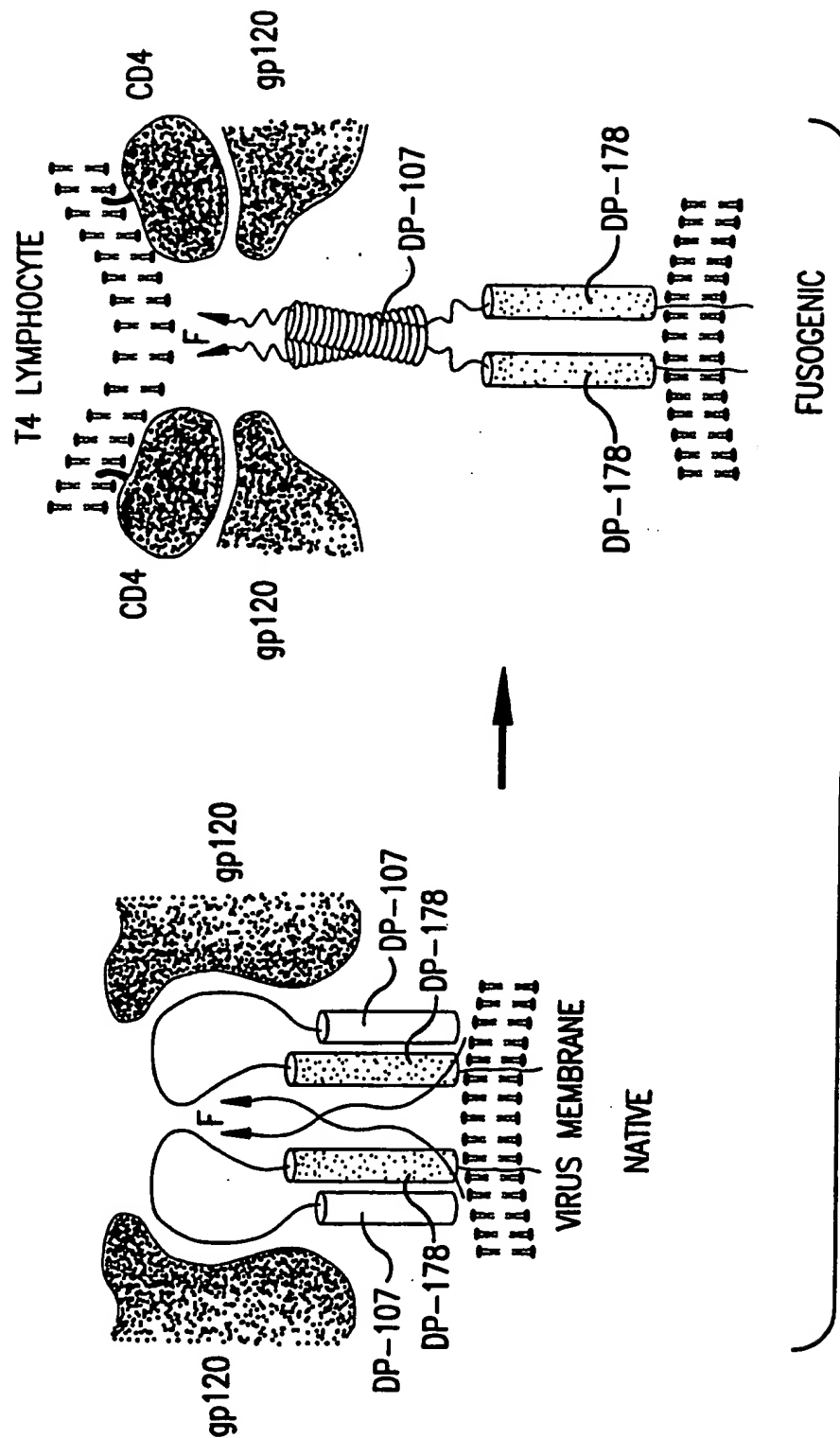
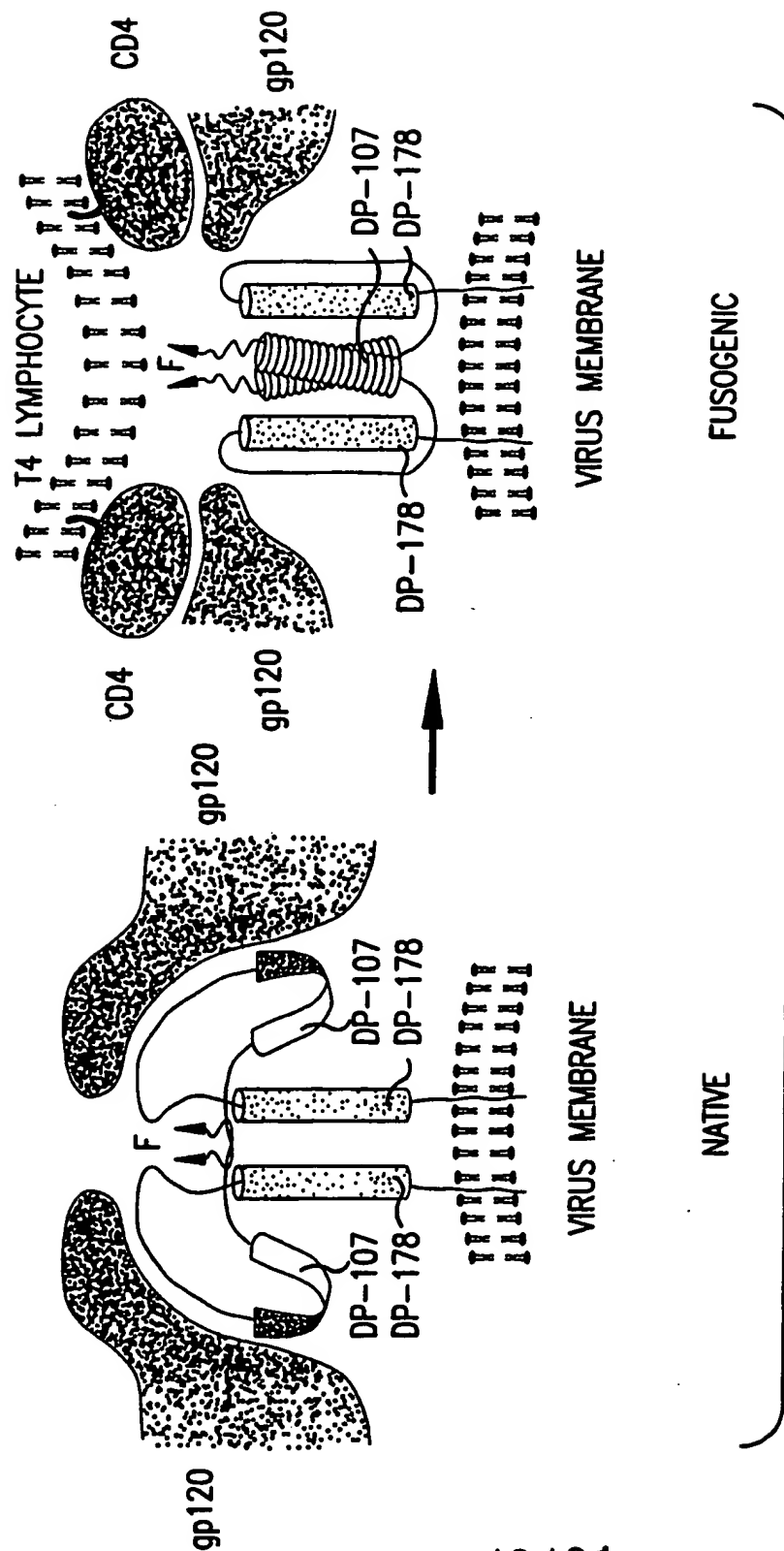


FIG.11A



Sequence	Positions																Motifs			
	A	D	A	D	A	A	D	A	D	A	D	A	D	A	D					
GCN4 (gcn4 yeast)	M	K	Q	L	E	D	K	V	E	E	L	L	S	K	N	[LMNV]	{CEGIMPTW}			
C-FOS (fos_human)	T	D	T	L	Q	A	E	T	D	Q	L	E	D	E	K	[IKLT]	{CFGHIMPRVWY}			
C-JUN (tap1_human)	I	A	R	L	E	E	K	V	K	T	L	K	A	Q	N	[AILNV]	{CDFGHILPVWY}			
C-MYC (myo_human)	E	Q	K	L	I	S	E	E	D	L	L	E	K	R	E	[ELR]	{ACFGMPVWY}			
FLU LOOP 36	I	E	K	T	N	E	K	F	H	Q	I	E	K	E	F	[FILTV]	{ACFLMPTVW}			

FIG.12

Sequence	A	D	A	D	A	D	A	D	A	D	A	D	A	D	Motifs
DP-107 (env_hv1bru)L1=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	[ILQT] {CFIMPSTY}
DP-107 (env_hv1bru)L1=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	[ILQTV] {CDFIMPST}
DP-107 (env_hv1bru)L1=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	[ILQTV] {CDFIMPST}
DP-107 (env_hv1bru)L2=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	[EKLNOV] {CDFKMPSTY}
DP-107 (env_hv1bru)L2=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	[EKLNOV] {CFKMPST}
DP-107 (env_hv1bru)L2=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	[EKLNOV] {CFKMPST}
DP-178 (env_hv1bru)Y1=A	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	[EKLQY] {ACFGMPRVWY}
DP-178 (env_hv1bru)Y1=A	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	[EKLQWY] {CFGMPRVY}
DP-178 (env_hv1bru)Y1=A	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	[EFKLOWY] {CFGMPRVY}
DP-178 (env_hv1bru)Y1=D	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	[EILNOSY] {ACFGMPRVWY}
DP-178 (env_hv1bru)Y1=D	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	[EILNOSWY] {CFGMPRVY}
DP-178 (env_hv1bru)Y1=D	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	[EFILNOSWY] {CFGMPRVY}

FIG.13

Sequence	Positions																Parent Motif	Hybrid Motif																			
	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D																					
GCM4 (gcn4 yeast)	M	K	Q	L	E	D	K	V	E	L	S	K	N	Y	H	L	E	N	E	V	A	R	L	K	K	L		[LMNV] {CFGIMPTW}									
DP-107 (env_hv1bru)L1=0	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	[ILOI] {CFIMPTSY}	[ILMNQTV] {CFIMPT}								
DP-107 (env_hv1bru)L1=0	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	L	A	V	E	R	Y	L			
DP-107 (env_hv1bru)L1=0	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	L	A	V	E	R	Y	L	K	D	Q
DP-107 (env_hv1bru)L2=0	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I		[EKLINQV] {CDFKMPSTV}	[EKLINQV] {CFIMP}							
DP-107 (env_hv1bru)L2=0	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	L	A	V	E	R	Y	L			
DP-107 (env_hv1bru)L2=0	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	L	A	V	E	R	Y	L	K	D	Q

**FIG. 14**



Sequence	A	D	A	D	A	D	A	D	A	D	A	D	Parent Motif	Hybrid Motif
	Positions													
GCM4 (gcn4 yeast)	M K Q L E D K V E E I L S K N Y H L E N E V A R L K K L												[LMNV] {CFGIMPTIW}	
DP-178 (env_hv1bru)YI=A	Y T S L I H S L I E E S O N Q O E K N E Q E L L E L D K												[EKLOY] {ACFGMPRVWY} [EKLINVY] {CFGMPH}	
DP-178 (env_hv1bru)YI=A	Y T S L I H S L I E E S O N Q O E K N E Q E L L E L D K W A S L W N W												[EKLOWY] {CFGMPRVY} [EKLINQWY] {CFGMP}	
DP-178 (env_hv1bru)YI=A	Y T S L I H S L I E E S O N Q O E K N E Q E L L E L D K W A S L W N W F												[EFKLOWY] {CFGMPRVY} [FEKLINQWY] {CFGMP}	
DP-178 (env_hv1bru)YI=0	Y T S L I H S L I E E S O N Q O E K N E Q E L L E L D K												[EILNQSY] {ACFGMPRVWY} [EILINQSVY] {CFGMPH}	
DP-178 (env_hv1bru)YI=0	Y T S L I H S L I E E S O N Q O E K N E Q E L L E L D K W A S L W N W												[EILNQSWY] {CFGMPRVY} [EILINQSVWY] {CFGMP}	
DP-178 (env_hv1bru)YI=0	Y T S L I H S L I E E S O N Q O E K N E Q E L L E L D K W A S L W N W F												[EFLINQSWY] {CFGMPRVY} [EFLINQSVWY] {CFGMP}	

FIG. 15

Sequence	Positions												Parent Motif	Hybrid Motif
	A	D	A	D	A	D	A	D	A	D	A	D		
DP-107 (env_hv1bru) L1=D	N	N	L	R	A	T	E	A	Q	H	L	L	[ILQTV] {CDFIMPST}	
DP-107 (env_hv1bru) L2=D	N	N	L	R	A	T	E	A	Q	H	L	L	[EKLQV] {CFKAPS}	
DP-178 (env_hv1bru) Y1=A	Y	T	S	L	I	H	S	L	I	E	S	Q	[EFKLQWY] {CFGMPRVY}	
DP-178 (env_hv1bru) Y1=D													[EFILQSWY] {CFGMPRVY}	[EFIKLQDSTVWY] {CFMP}
FLU LOOP 36	I	E	K	T	N	E	K	F	H	Q	I	E	[FILTV] {ACFLMPTVW}	

FIG.16

Sequence	Positions												Parent Motif	Hybrid Motif
	A	D	A	D	A	D	A	D	A	D	A	D		
GCN4 (gcn4 yeast)	MKQLLEDKVEE	LLSKN	YHLENE	VARL	KKL								[LMNV] {CFGIMPTH}	
DP-107 (env_hv1bru)L1=D	NNLLRAIEAQ	HLQL	TVWCI	KQL	QARIL	AVERYL	KDQ						[ILOTV] {CDFIMPST}	
DP-178 (env_hv1bru)Y1=A	YTSLSIMSLIE	ESQNOQ	EEKNEQE	ELLE	LDKWA	SLWNWF							[EFKLOWY] {CFGMPRVY}	[EFIKLANQTVWY] {CFMP}
GCN4 (gcn4 yeast)	MKQLLEDKVEE	LLSKN	YHLENE	VARL	KKL								[LMNV] {CFGIMPTH}	
DP-107 (env_hv1bru)L1=D	NNLLRAIEAQ	HLQL	TVWCI	KQL	QARIL	AVERYL	KDQ						[ILOTV] {CDFIMPST}	
DP-178 (env_hv1bru)Y1=D	YTSLSIHSLIE	ESQNOQ	EEKNEQE	ELLE	LDKWA	SLWNWF							[EFILNOSWY] {CFGMPRVY}	[EFILNQRSTWY] {CFMP}
GCN4 (gcn4 yeast)	MKQLLEDKVEE	LLSKN	YHLENE	VARL	KKL								[LMNV] {CFGIMPTH}	
DP-107 (env_hv1bru)L2=D	NNLLRAIEAQ	HLQL	TVWCI	KQL	QARIL	AVERYL	KDQ						[EKLNV] {CFKMP}	
DP-178 (env_hv1bru)Y1=A	YTSLSIHSLIE	ESQNOQ	EEKNEQE	ELLE	LDKWA	SLWNWF							[EFKLOWY] {CFGMPRVY}	[EFKLANQWY] {CFMP}
GCN4 (gcn4 yeast)	MKQLLEDKVEE	LLSKN	YHLENE	VARL	KKL								[LMNV] {CFGIMPTH}	
DP-107 (env_hv1bru)L2=D	NNLLRAIEAQ	HLQL	TVWCI	KQL	QARIL	AVERYL	KDQ						[EKLNV] {CFKMP}	
DP-178 (env_hv1bru)Y1=D	YTSLSIHSLIE	ESQNOQ	EEKNEQE	ELLE	LDKWA	SLWNWF							[EFILNOSWY] {CFGMPRVY}	[EFIKLANQSWY] {CFMP}

FIG.17

Sequence	Positions												Parent Motif	Hybrid Motif	
	A	D	A	D	A	D	A	D	A	D	A	D			
UCM4 (gcn4 yeast)	MKQL	EDKVEE	L	SKN	YH	L	EN	V	A	R	L	K	K	[LMNV] {CFGIMPSTW}	
DP-107 (env_hv1bru) L1=D	NNL	LRA	IEAQ	HL	L	QL	T	V	W	G	I	K	Q	[ILOTV] {CDFIMPST}	
DP-107 (env_hv1bru) L2=D	NNL	LRA	IEAQ	HL	L	QL	T	V	W	G	I	K	Q	[EKLNV] {CFKAPS}	
DP-178 (env_hv1bru) Y1=A	YTS	L	I	H	S	L	I	E	S	Q	N	Q	E	[EFKLOW] {CFGMPRVY}	
DP-178 (env_hv1bru) Y1=D	YTS	L	I	H	S	L	I	E	S	Q	N	Q	E	[EFILNDSWY] {CFGMPRVY}	
C-FOS (fos_human)	TDT	L	Q	A	E	T	D	Q	L	E	D	E	K	[IKLT] {CFGHIMPVWY}	
C-JUN (tap1_human)	IAR	L	E	E	K	V	K	T	L	K	A	Q	N	[AILNV] {CDFGHILPVWY}	
C-MYC (myo_human)	EQL	I	S	E	E	D	L	L	E	K	R	R	E	[ELR] {ACFGMPVWY}	
FLU LOOP 36	IEKT	N	E	K	F	H	Q	I	E	K	E	F	I	[FILTV] {ACFLMPTVW}	
														[AEFIKLMNOSTVWY] {CFP}	
														= {CDGHP} {CFP}	

FIG.18

P-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-{P}(1)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-{P}(2)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-{P}(3)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-{P}(4)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-{P}(5)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-{P}(6)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-{P}(7)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-{P}(8)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-{P}(9)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-{P}(10)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-X(1,12)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]  
 P-X(13,23)-[LIV]-{P}(6)-[LIV]-{P}(6)-[LIV]

FIG.19

Fusion ♥ALLMOTI5♥  
 Peptide ♣107x178x4♣  
 ♥.....ELGELG A AGSTMGARSM TLTVQARQ ♣LLSGIVQQQ DPI07-NNL

LRAIEAQOHL LOLTWGIKO LOARILAYER YLKDO-DPI07 QLLG♥♥ I WGC

♥ALLMOTI5♥ ♣107x178x4♣  
 \*LVS Coiled-Coil\*  
 SGKLICT TAVP ♥WNASWS NKSLEQIWNN MTWM \*E ♣WDREINN DPI78-

YTSLIHSL IEESONOOEK NEOELLELDK\* WASLWNWF-DPI78 NI

♦Transmembrane Region♦  
 TNWLWYIK♣ ♦IEIMIVGGLVGLRIVEAVLSIV NRVROGYS♥ PL

♣P23LZIPC♣  
 SFQTHLPTPR GPDR ♣PEGIEE EGGERDRDRS IRLVNGSLAL IWDDLRLSL♣ CL

♥ALLMOTI5♥ ♣107x178x4♣  
 F ♥SYHRLRDLL LIVTRIVELL GRRGW ♣EALKYWWNLLOYWSQ

ELKNSAVSLLNAT♣ AIAVAEG TDRVIEVVQG A♥ CRAIRHIPR

RIRQGLERIL L

FIG. 20

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SUBSTITUTE SHEET (RULE 26)

Fusion ♡ALLMOTI5♡  
 Peptide ♡107x178x4♡  
 ♡.....ELGEL LGVGSALAS GVA ♡VSKVLHLEGEVNKIKSA

♡P1&12LZIPC♡  
LLSTNKA VVS LSNGVSVLTS KVL DLK NYID KQ ♡ ♡ LL ♡PIVNKQ

♡107x178x4♡  
 SC ♡SISNIETV I ♡ EEOOKNNRLLETREFSVNAG ♡ V TTPVSTMLTNSELLSL

♡P1&12LZIPC♡  
 ♡ALLMOTI5♡  
 INDM ♡PI ♡TNDQ KKLMSNNVQI V ♡ RQSYSI ♡ MS IIKEEVLAYV

VQ ♡ LPLYGVID TPCWKLHTSP LCTTNTKEGS NICLTRTDRG WYCDNAGSVS

FFPQAETCKV QSNRVFCDTM NSLTLPSEIN LCNVDIFNPK

YDCKIMTSKT DVSSSVITSL GAIVSCYGKT KCTASNKNRG

IIKTFSNGCDYVSNKGMDTV SVGNTLYYVN KQEGKSLYVK G

♡P7, 12, & 23LZIPC♡  
 ♡107x178x4♡ ♡ALLMOTI5♡  
 EPIINFYDPLVF ♡PSDE ♡EDASISQVNEKINOSLAF ♡I ♡ RKSDELL ♡

♡Transmembrane Region ♡  
HNVNA ♡ GK STTN ♡IMITTLIIIVILLSLIAVGLLLY ♡ C ♡

KARSTPVTLS KDQLSGINNI AFSN

FIG. 21

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Fusion  
 Peptide ♡ALLMOTI5♡ ♡107x178x4♡  
 .....FLGFLG ♡AAGTA MGAAA ♡TALTVOSOHLLAGILOQQKNLLAAV

♡107x178x4♡  
EAQ ♡ QQM ♡LKLTIWGVKNLNARVTALEKYLEDOARLN ♡ AWG♡ CA

\*LVS Coiled-Coil\*  
 ♡ALLMOTI5♡ ♡107x178x4♡  
 WKQVCHTTVP WQWNNRTPDW ♡NNMT \*WLE ♡WEROISYLEGNTT

♡107x178x4♡  
TOLEEARAQEEKNLD ♡ AYOKLSS\* WSDFWSW♡ FDF ♡SKWLN ♡ILK

♦Transmembrane Region♦  
IGFLDVLGIGLRLLYTV♦ YS ♡ CIARVRQGYSP LSPQIHHP WKGQPDNAEG

PGEGGDKRKN SSEPWQKESG TAEWKS NWCK RL TNWCSISS IWL YNS

♡ALLMOTI5♡  
 ♡CLTL LVHLRSAFQY IQYGLGELKA AAQEAVVALA RLAQNAGYQIWL♡

ACRSAYRA IINSPRRVRQ GLEGILN

FIG. 22



Fusion ♣107x178x4♣  
 Peptide ♡ALLMOTIS♡ \*LVS Coiled-Coil\*  
 .....EAG ♡VYL AGVALGVATA AQITAGIALHQ ♣\*SNLNAQAIQ

SLRTSLEQSNKAIEEIREATOETVIA\* VOGVQDY♣ VNNEL♡ VP

♡ALLMOTIS♡  
 ♣107x178x4♣  
 ♣P6 & 12LZIPC♣

AMQHMSCELVGQRLGLRLLRYYTELLSIFGPSLRD ♣PISA ♣♡EISIQALIVAL

GGEIHKILEKLGYSGSD♣ MIAILES RGIKTKI♡ THVDLP GKF ILSISY

♣P1 & 12LZIPC♣  
 ♣PTLSEVKG VIVHRLEAV♣ SYNIGSQEWYTTVP RYIATNGYLISN FDESSCVFVS

ESAICSQNSL YPMSPLLQQC IRGDTSSCAR TLVSGTMGNK FILSKGNIVA

NCASILCKCY STSTINQSP DKLLTFLASD TCPLVEIDGA TIQVGGRQYP

\*LVS Coiled-Coil\*  
 ♡ALLMOTIS♡  
 ♣P12 & 23LZIPC♣

DMVYEGKVAL G ♣PAISLD ♡RL\*DVGTNLGNALKKLDDAKVLI♣

♦Transmembrane Region♦

DSS♣ NOILETVR RS♡\* SFN ♦EGSLL SVPIL SCTAL ALLLLIYCC♦

K RRYQQTLKQH TKVDPAFKPD LTGTSKSYVR SL

FIG. 23

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SUBSTITUTE SHEET (RULE 26)

Fusion ♥ALLMOTIS♥  
 Peptide ♣107x178x4♣  
 ♥.....FIGAI IGSVALGVA TAAQITAASA LIQANQNAAN ♣ILRLKESITA  
  
TIEAVHEVTDGLSQLAVA♣ VG KM♥ QQFVNDQFNNTAQELDCIKITQQV  
  
 ♥ALLMOTIS♥  
 GVELNLYLTELT TV FGPQITSPAL ♥TQLTIQALYNAGGNMDYLLTKLGVG  
  
 ♣P1 & 12LZIPC♣  
 NNQLSSLIGSGLIT GN♥ ♣PILYDSQT QLLGIQVTLP SVGNLNNMRATYLET  
  
 LSVST TKGFASALVP KVVVTQVGSVI EELDTSYCIE TDLDLYCTRI VTFPMSPGIY  
  
 SCLNGNTSAC MYSKTEGALT TPYMTLKGSV IANCKMTTCR CADPPGIISQ  
  
 ♥ALLMOTIS♥  
 ♣107x178x4♣  
 NYGEAVSLID RHSCN ♣♥VLSLD GITLRLSGEF DATYQKNISI LDSQVIVTG  
  
 \*LVS Coiled-Coil\* ♦Trans-  
 \*NLDISTELGNV NNSISNALDK LEESNSKLDK VNVKLTSTSA ♦LIT\* YIA  
  
membrane Region♦  
LTAISLVCGLSLV♥♣ LACYLMY♦ KQKAQQKTLLWLGNNILGQMRATTKM

FIG. 24

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Fusion ♥ALLMOTIS♥  
 Peptide ♣107x178x4♣ \*LVS Coiled-Coil\*  
 .....EEGGV ♣IG ♥TIALG \*YATSAQITAAYALVEAKQARSDIEKLKE

AIRDTNKAVOSVOSSIGNLIVAIKSVQ\* DYVNKE♥♣ IVPSIARLGCEAAG

♥ALLMOTIS♥  
 ♣107x178x4♣  
 LQLGIALTQH ♣♥YSELTNIEGDNIGSLOEKGIKLOGIASLYRTNITE♥♣

♣P5 & 12LZIPC♣  
 IFTTSTVDKYDIYDLLFTESIKVRVIDVDLNDYSITLQVRL ♣PLLTRLNTQIYR

VDSISYNI♣ QNREWI♣ PLPSHIMTKGAFLGGADVKECIEAFSSYIC

PSDPGFVLNHEMESCLSGNISQCPRTVVKSDIVPRYAFVNGGVVANCITT

TCTCNGIGNRINQPPDQGVKIITHKECNTIGINGMLFNTNKEGTLAFYTP

♥ALLMOTIS♥  
 ♣107x178x4♣  
 ♣P6 & 23LZIPC♣  
 NDITLNNVALD ♣PIDI ♣SIELN ♥KAKSDLEESKEWI♣ RRSNOKL♣

♦Transmembrane Region♦  
DSIGNWHOSSIT ♦IIIV♣ LIMIIILEIINVTII♦ IIHVKY♥ R

IQKRNRVDQN DKPYVLTNK

FIG. 25

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Fusion  
Peptide  
.....GLEGAI AGFIENGWEGMIDGWYGFRHQNSEGTG

♠107x178x4♠

♥ALLMOTI5♥

\*LVS Coiled-Coil\*

\*Q ♥AADLKST ♠QAAIDQINGKLNRVIEKTNEKTHQIEKEESEVEGRIQ

DLEKYYEDTKIDL\* WSYNAELLVALENQHTI♠ DLT♥ DSEMKNLFETR

RQLRENAEEMGNGCFKIYHKCDNACIESIRNGTYDHDVYRDEALNNRFQIKG

VELKSGYKDWILWISFAISCFLLCVLLGFIMWACQQRGNIRCNICI

FIG. 26

AV	CD	RSV F2	YTSVITIELSNIKENKCNCTDAKVKL IKQELDKYKNVTEQLQLMOST
+	+ / ++	T-142	YTSVITIELSNIKENKCNCTDAKVKL IKQELDKYK
++	+ / +++	T-143	TSVITIELSNIKENKCNCTDAKVKL IKQELDKYKN
+	+ / ++	T-144	SVITIELSNIKENKCNCTDAKVKL IKQELDKYKNA
-	+ / +	T-145	VITIELSNIKENKCNCTDAKVKL IKQELDKYKNV
-	+ / -	T-146	ITIELSNIKENKCNCTDAKVKL IKQELDKYKNV
-	-	T-147	TIIELSNIKENKCNCTDAKVKL IKQELDKYKNVTE
-	-	T-148	IELSNIKENKCNCTDAKVKL IKQELDKYKNVTE
-	+ / -	T-149	ELSNIKENKCNCTDAKVKL IKQELDKYKNVTELO
-	-	T-150	LSNIKENKCNCTDAKVKL IKQELDKYKNVTELO
-	+ / +	T-151	SNIKENKCNCTDAKVKL IKQELDKYKNVTELOLL
-	+ / ++	T-152	NIKENKCNCTDAKVKL IKQELDKYKNVTELOLLM
-	+ / +	T-153	IKENKCNCTDAKVKL IKQELDKYKNVTELOLLMQ
-	+ / ++	T-154	KENKCNCTDAKVKL IKQELDKYKNVTELOLLMQS
-	+ / +	T-155	ENKCNCTDAKVKL IKQELDKYKNVTELOLLMOST

FIG.27

AV	CD	RSV	
++	+/-	T-67	DEFDASISQWNEKINQSLAF IRKSDELL
		F1-178	GEPIINFYDPLVFPSEDFDASISQWNEKINQSLAF IRKSDELLHNNWAGKSTT
+/-		T-104	IIINFYDPLVFPSEDFDASISQWNEKINQSLAF IRK
+/-		T-105	INFYDPLVFPSEDFDASISQWNEKINQSLAF IRKS
+/-		T-106	NFYDPLVFPSEDFDASISQWNEKINQSLAF IRKSD
+		T-107	FYDPLVFPSEDFDASISQWNEKINQSLAF IRKSDE
++		T-108	YDPLVFPSEDFDASISQWNEKINQSLAF IRKSDEL
++		T-109	DPLVFPSEDFDASISQWNEKINQSLAF IRKSDELL
+		T-110	PLVFPSEDFDASISQWNEKINQSLAF IRKSDELLH
++		T-111	LVFPSEDFDASISQWNEKINQSLAF IRKSDELLHN
++	+/-	T-112	VFPSEDFDASISQWNEKINQSLAF IRKSDELLHNV
++	+/-	T-113	FPSDEDFDASISQWNEKINQSLAF IRKSDELLHNV
++	+/-	T-114	PSDEDFDASISQWNEKINQSLAF IRKSDELLHNVNA
++	+/-	T-115	SDEDFDASISQWNEKINQSLAF IRKSDELLHNVNAG
++	+/-	T-116	DEFDASISQWNEKINQSLAF IRKSDELLHNVNAGK
++	+/-	T-117	EFDASISQWNEKINQSLAF IRKSDELLHNVNAGKS
++	+/-	T-118	FDASISQWNEKINQSLAF IRKSDELLHNVNAGKST
++	+/-	T-119	DASISQWNEKINQSLAF IRKSDELLHNVNAGKSTT

(T-67 LIKE)

FIG.28

AV	CD	HPF 3	178	YTPNDITLNSVALDPIDISIELNKAQSDLEESKEWIRRSNQKLD SIGNWHQSSTT
-	-		189	YTPNDITLNSVALDPIDISIELNKAQSDLEESKE
-	-		190	TPNDITLNSVALDPIDISIELNKAQSDLEESKEW
-	-		191	PNDITLNSVALDPIDISIELNKAQSDLEESKEWI
-	-		192	NDITLNSVALDPIDISIELNKAQSDLEESKEWIR
-	+/-		193	DITLNSVALDPIDISIELNKAQSDLEESKEWIRR
+/-	+/-		194	ITLNSVALDPIDISIELNKAQSDLEESKEWIRRS
+/-	+/+		195	TLNSVALDPIDISIELNKAQSDLEESKEWIRRSN
+	+/+		196	LNSVALDPIDISIELNKAQSDLEESKEWIRRSNQ
+	+/+		197	NNSVALDPIDISIELNKAQSDLEESKEWIRRSNQK
+++	+/+		198	NSVALDPIDISIELNKAQSDLEESKEWIRRSNQKL
++	+/+		199	SVALDPIDISIELNKAQSDLEESKEWIRRSNQKLD
-			200	VALDPIDISIELNKAQSDLEESKEWIRRSNQKLD S
+++			201	ALDPIDISIELNKAQSDLEESKEWIRRSNQKLD SI
+++			202	LDPIDISIELNKAQSDLEESKEWIRRSNQKLD SIG
+++			203	DPIDISIELNKAQSDLEESKEWIRRSNQKLD SIGN
+++			204	PIDISIELNKAQSDLEESKEWIRRSNQKLD SIGNW
+++			205	IDISIELNKAQSDLEESKEWIRRSNQKLD SIGNWH
+			206	DISIELNKAQSDLEESKEWIRRSNQKLD SIGNWHQ
+			207	ISIELNKAQSDLEESKEWIRRSNQKLD SIGNWHQS
+			208	SIELNKAQSDLEESKEWIRRSNQKLD SIGNWHQSS
++			209	IELNKAQSDLEESKEWIRRSNQKLD SIGNWHQSST
++			210	ELNKAQSDLEESKEWIRRSNQKLD SIGNWHQSSTT

FIG.29

CD	HPF3 107	GTIALGVATSAQITA AVALVEAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNLIVA I KSVQDYVNKE IVP
+/+	157	ALGVATSAQITA AVALVEAKQARSDIEKLKEAIRD
+/+	158	LGVATSAQITA AVALVEAKQARSDIEKLKEAIRDT
+/-	159	GVATSAQITA AVALVEAKQARSDIEKLKEAIRDTN
+/+	160	VATSAQITA AVALVEAKQARSDIEKLKEAIRDTNK
+/+	161	ATSAQITA AVALVEAKQARSDIEKLKEAIRDTNKA
+/-	162	TSAQITA AVALVEAKQARSDIEKLKEAIRDTNKAV
+/+	163	SAQITA AVALVEAKQARSDIEKLKEAIRDTNKAVQ
+ /+++	164	AQITA AVALVEAKQARSDIEKLKEAIRDTNKAVQS
+/+	165	QITA AVALVEAKQARSDIEKLKEAIRDTNKAVQSV
+/-	166	ITA AVALVEAKQARSDIEKLKEAIRDTNKAVQSVQ
+/-	167	TAAVALVEAKQARSDIEKLKEAIRDTNKAVQSVQS
+/-	168	AAVALVEAKQARSDIEKLKEAIRDTNKAVQSVQSS
+/-	169	AVALVEAKQARSDIEKLKEAIRDTNKAVQSVQSSI
+/-	170	VALVEAKQARSDIEKLKEAIRDTNKAVQSVQSSIG
+/-	171	ALVEAKQARSDIEKLKEAIRDTNKAVQSVQSSIGN
+/-	172	LVEAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNL
+/-	173	VEAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNLI
+ /++	174	EAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNLIV
T-40		AKQARSDIEKLKEAIRDTNKAVQSVQSSIGNLIVA
+/+	175	KQARSDIEKLKEAIRDTNKAVQSVQSSIGNLIVAI
+ /+++	176	QARSDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIK
+/-	177	ARSDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIKS
+/-	178	RSDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIKSV
-	179	SDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIKSVQ
-	180	DIEKLKEAIRDTNKAVQSVQSSIGNLIVAIKSVQD
-	181	IEKLKEAIRDTNKAVQSVQSSIGNLIVAIKSVQDY
-	182	EKLKEAIRDTNKAVQSVQSSIGNLIVAIKSVQDYV
+/+	183	KLKEAIRDTNKAVQSVQSSIGNLIVAIKSVQDYVN
+ /+++	184	LKEAIRDTNKAVQSVQSSIGNLIVAIKSVQDYVNK
-	185	KEAIRDTNKAVQSVQSSIGNLIVAIKSVQDYVNKE
-	186	EAIRDTNKAVQSVQSSIGNLIVAIKSVQDYVNKEI
-	187	AIRDTNKAVQSVQSSIGNLIVAIKSVQDYVNKEIV
-	188	IRDTNKAVQSVQSSIGNLIVAIKSVQDYVNKEIVP

FIG.30

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SUBSTITUTE SHEET (RULE 26)



## INTERNATIONAL SEARCH REPORT

International application No.

PCT/US94/05739

<b>A. CLASSIFICATION OF SUBJECT MATTER</b> IPC(5) : A61K 37/02, 39/12; C12Q 1/70; G01N 33/53 US CL : 424/88, 89; 435/5, 7.1, 7.92-7.95, 974; 530/324-331, 333, 334 According to International Patent Classification (IPC) or to both national classification and IPC																								
<b>B. FIELDS SEARCHED</b> Minimum documentation searched (classification system followed by classification symbols) U.S. : 424/88, 89; 435/5, 7.1, 7.92-7.95, 974; 530/324-331, 333, 334 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) APS, Biosis																								
<b>C. DOCUMENTS CONSIDERED TO BE RELEVANT</b>																								
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.																						
NONE	NONE	NONE																						
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.																								
<table border="0"><tr><td>* Special categories of cited documents:</td><td>*T</td><td>later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</td></tr><tr><td>*A</td><td>document defining the general state of the art which is not considered to be of particular relevance</td><td></td></tr><tr><td>*E</td><td>earlier document published on or after the international filing date</td><td>*X</td><td>document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</td></tr><tr><td>*L</td><td>document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</td><td>*Y</td><td>document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</td></tr><tr><td>*O</td><td>document referring to an oral disclosure, use, exhibition or other means</td><td>*G</td><td>document member of the same patent family</td></tr><tr><td>*P</td><td>document published prior to the international filing date but later than the priority date claimed</td><td></td><td></td></tr></table>			* Special categories of cited documents:	*T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention	*A	document defining the general state of the art which is not considered to be of particular relevance		*E	earlier document published on or after the international filing date	*X	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone	*L	document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Y	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art	*O	document referring to an oral disclosure, use, exhibition or other means	*G	document member of the same patent family	*P	document published prior to the international filing date but later than the priority date claimed		
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*E	earlier document published on or after the international filing date	*X	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone																					
*L	document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Y	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art																					
*O	document referring to an oral disclosure, use, exhibition or other means	*G	document member of the same patent family																					
*P	document published prior to the international filing date but later than the priority date claimed																							
Date of the actual completion of the international search 07 SEPTEMBER 1994		Date of mailing of the international search report 26 SEP 1994																						
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230		Authorized officer JEFFREY STUCKER Telephone No. (703) 308-0196																						

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US94/05739

## Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 2  
because they relate to subject matter not required to be searched by this Authority, namely:  
that the claimed subject matter is directed to mental processes.
2. ☒ Claims Nos.: 13-16 and 42-49  
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:  
because the sequences have not been submitted to the International Searching Authority in electronic form.
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.  
☐ No protest accompanied the payment of additional search fees.